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Dephosphorylation of the thylakoid membrane light-harvesting complex-II by a stromal protein phosphatase

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Abstract

Light-harvesting complex-II (LHC-II) phosphatase activity has generally been examined in the intact thylakoid membrane. A recent report of peptide-phosphatase activity associated with the chloroplast stromal fraction (Hammer, M.E et al. (1995) *Photosynth Res* 44:107-115) has led to the question of whether this activity is capable of dephosphorylating membrane-bound LHC-II. To this end, heat-treated thylakoid membranes were examined as a potential LHC-II phosphatase substrate. Following incubation of the thylakoid membrane at 60 °C for 15 min, the endogenous protein phosphatase and kinase activities were almost eliminated. Heat-inactivated phosphomembranes exhibited minimal dephosphorylation of the light harvesting complex-II. Peptide-phosphatase activities isolated from the thylakoid and stromal fraction were able to dephosphorylate LHC-II in heat-inactivated phosphomembranes. The stromal phosphatase showed highest activity against LHC-II at pH 9. Dephosphorylation of the LHC-II by the stromal enzyme was not inhibited by molybdate, vanadate or tungstate ions, but was partially inhibited by EDTA and a synthetic phosphopeptide mimicking the LHC-II phosphorylation site. Thus, the previously identified stromal phosphatase does appear capable of dephosphorylating authentic LHC-II in vivo.

Keywords: chloroplast, LHC-II, protein phosphorylation

Abbreviations: CPP – chymotryptic phosphopeptides; LHC-II – light-harvesting complex of Photosystem II; MP – protein phosphatase fractionated from the thylakoid membrane; P_{2Thr} – synthetic phosphopeptide MRK-SAT(p)TKKVW; SP – protein phosphatase fractionated from the stromal compartment

Introduction

Chloroplast thylakoid membranes contain a number of proteins which are capable of being phosphorylated, with the most abundant being those associated with the LHC-II (Bennett 1977). The phosphorylation of thylakoid membrane proteins is accomplished through one or more protein kinases (Bennett 1991; Allen 1992). The LHC-II kinase is light regulated; phosphorylation of the LHC II is activated by reduction of plastoquinone, which is coincident with the State 1 to State 2 transition in photosynthetic energy distribution (Allen 1995).

Dephosphorylation of thylakoid proteins appears to be catalyzed by at least one phosphatase located in the thylakoid membrane (Bennett 1980). Isolation and characterization of a thylakoid protein phosphatase has been difficult due to its membrane location and the hydrophobic nature of the phosphoprotein substrates. Thylakoid membranes isolated from highly purified chloroplasts do not dephosphorylate general phosphatase substrates such as phosphorylase α , histone, β -glycerolphosphate, or 4-methylumbelliferylphosphate (MacKintosh et al. 1991; Sun and Markwell 1992). Use of synthetic phosphopeptides analogous the LHC-II N-ter-

minus phosphorylation site (Sun et al. 1993) or authentic phosphopeptides cleaved from the thylakoid proteins (Hammer et al. 1995) has recently allowed for the detection of solubilized protein phosphatase activity away from the membrane and the determination of initial rates of protein phosphatase activity. Synthetic phosphopeptides mimicking the LHC-II N-terminus have also been shown to inhibit the dephosphorylation rate of phosphoproteins in intact thylakoid membranes (Sun et al. 1993; Cheng et al. 1994).

Peptide substrates mimicking the LHC-II have allowed for the detection of a peptide phosphatase from the chloroplast stroma (Hammer et al. 1995). The stromal phosphatase and a detergent liberated membrane phosphatase were both inhibited by NaF and EDTA, but the stromal phosphatase had a more alkaline pH optimum and displayed much less inhibition by molybdate ions than did the membrane phosphatase. The total phosphopeptide phosphatase activity of the stromal phosphatase was similar to that of the membrane phosphatase, and thus could be a significant factor in thylakoid membrane dephosphorylation (Hammer et al. 1995).

To assess whether the stromal phosphatase could dephosphorylate authentic LHC-II apoproteins, we have developed an assay using heat-inactivated phosphomembranes as *in vitro* substrates for the protein phosphatase. Phosphorylated thylakoid membrane phosphoproteins, containing low endogenous protein phosphatase or kinase activity, were dephosphorylated by the stromal phosphatase, indicating that this enzyme is capable of LHC-II dephosphorylation *in vivo*.

Materials and methods

Chloroplast isolation

Pea (*Pisum sativum* L. cv. Alaska) plants were grown, and intact chloroplasts were isolated and purified on Percoll gradients as previously described (Sun et al. 1992). The intact chloroplasts were hypotonically lysed by resuspension in 25 mM Na-Tricine (pH 7.6), 5 mM MgSO₄ at a Chl concentration less than 0.2 mg ml⁻¹, with the thylakoid membrane pellet and the stromal supernatant fractionated by centrifugation at 20,000 g for 10 min. The membrane pellet and stromal fractions were stored in the dark for less than 1 h at 5 °C prior to subsequent analysis.

Chl concentrations were measured in 80% acetone extracts (Arnon 1949).

Preparation of heat-inactivated phosphomembranes

Heat-inactivated phosphomembranes were prepared from isolated thylakoid membranes following incubation in [γ -³²P]ATP to label the endogenous phosphoproteins. Membranes containing 10 mg Chl were resuspended in 10 ml total volume containing the following: 25 mM Na-Tricine (pH 7.6), 5 mM MgSO₄, 20 mM NaF, 200 μ M unlabeled ATP, and 1 mCi [γ -³²P]ATP (7000 Ci mmol⁻¹, ICN Biomedicals, Inc.). The mixture was incubated in the light (500 / μ mol photons m⁻² s⁻¹ in the 400 to 700 nm range) at 30 °C for 15 min. Following the incubation, the mixture containing the phosphorylated membranes was made to 20 mM EDTA and placed in a 60 °C water bath for 30 min. Membranes were then pelleted by centrifugation at 20,000 g for 10 min, suspended in 25 mM Na-Tricine (pH 7.6), 5 mM MgSO₄, and pelleted again as above. The final membrane pellet was suspended in 3 ml 100 mM Tris C1 (pH 8.5), 5 mM Mg acetate, 5 mM dithiothreitol and 20% glycerol. Heat-inactivated phosphomembranes were stored at -80 °C until use.

Protein phosphatase fractionation

The Brij 35-liberated membrane CPP-phosphatase, or the stromal CPP-phosphatase, were fractionated using Sephacryl S-200 chromatography as previously described (Hammer et al. 1995). Phosphatase containing fractions were dialyzed into 5 mM Tris-C1 (pH 8.5), 5 mM Mg acetate, 1 mM DTT prior to assay with heat-inactivated phosphomembranes.

Assay of exogenous phosphatases using heat-inactivated phosphomembranes

Heat-inactivated, labeled phosphomembranes were thawed, centrifuged at 15,000 g, and the pellet resuspended at 2 mg Chl ml⁻¹ with a buffer containing 100 mM Tris C1 (pH 8.5), 5 mM Mg acetate, 5 mM dithiothreitol (for the stromal phosphatase) or with a buffer containing 100 mM HEPES (pH 7.2), 5 mM Mg acetate, 5 mM dithiothreitol (for the membrane phosphatase). Phosphatase fractions isolated from the membrane or stroma containing ca. 5 pmol min⁻¹ ml⁻¹ CPP-phosphatase activity were mixed with an equal volume of heat-inactivated phosphomembranes and incubated at 30 °C for various lengths of time. Reactions were stopped with the addition of an equal volume of denaturing

sample buffer (125 mM Tris-Cl, pH 6.8, 10% SDS, 5% β -mercaptoethanol, 25% glycerol, 0.05% bromophenol blue), heated at 90 °C for 15 min, then separated using discontinuous SDS-PAGE (Laemmli 1970). Gels were stained with Coomassie brilliant blue, dried, and the ^{32}P imaged using a Phosphorimager SF (Molecular Dynamics, Sunnyvale, CA, USA). The LHC-II band from the control reactions generally contained ca. 6000 dpm radioactivity.

For the measurement of pH optima, heat-inactivated phosphomembranes and the stromal phosphatase were incubated in 250 mM Tris-Cl, adjusted to pH 7 through 9, containing 5 mM dithiothreitol, and 5 mM magnesium acetate.

Preparation of phosphopeptides and peptide phosphatase assay

To initially assay endogenous phosphatase activity, chymotryptic phosphopeptides (CPP) were used as substrates. The CPP were prepared from isolated thylakoid membranes as previously described (Hammer et al. 1995). Briefly, thylakoid membranes were incubated with [γ - ^{32}P]ATP to label the endogenous phosphoproteins. The membranes were then incubated with chymotrypsin to liberate phosphorylated peptides. Peptides were fractionated using C_{18} -silica chromatography (Hammer et al. 1995).

The synthetic peptide $\text{P}_{2\text{Thr}}$ (MRKSATTKKVV) was phosphorylated by incubation with isolated thylakoid membranes as previously described (Sun et al. 1993). Phosphorylation of this peptide by the endogenous thylakoid protein kinase activity results in a specific phosphorylation of Thr⁶ (Michel and Bennett 1989).

The assay used to measure CPP-dephosphorylation was previously described (Hammer et al. 1995). Following incubation of the CPP with the enzyme fraction, $^{32}\text{P}_i$ released was assayed by phase partitioning (Shacter 1984) and Cerenkov counting. The determination of phosphopeptide concentration and P_i released during the assay were based on the ^{32}P specific radioactivity in the labeling reaction.

Results and discussion

Heat-inactivated thylakoid phosphomembranes

To develop a phosphomembrane substrate for the chloroplast protein phosphatase that did not contain endogenous phosphatase activity, it was first necessary

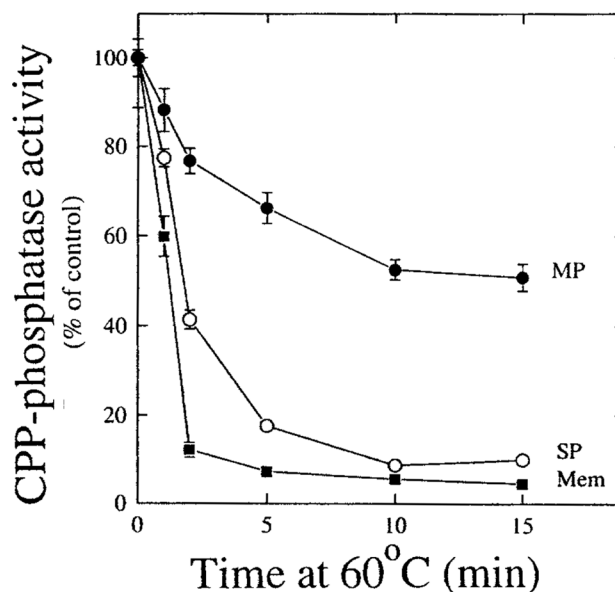


Figure 1. The effect of heating at 60 °C on phosphopeptide-phosphatase activity in pea chloroplast fractions. Activity was assayed from isolated thylakoid membranes (Mem), detergent-liberated, and Sephacryl-S-200 fractionated membrane phosphatase (MP), or Sephacryl-S-200 fractionated stromal phosphatase (SP). Following incubation at 60 °C for the indicated lengths of time, fractions were cooled to 30 °C and assayed for protein phosphatase activity using chymotryptic phosphopeptides (CPP) as a substrate. Specific activities at 100% for Mem, 8.7 pmol P_i released $\text{min}^{-1} \text{mg}^{-1}$ protein; MP, 23.7 pmol $\text{min}^{-1} \text{mg}^{-1}$ protein; SP, 37.8 pmol $\text{min}^{-1} \text{mg}^{-1}$ protein.

to examine conditions that would lead to phosphatase inactivation while largely maintaining membrane integrity. When thylakoid membranes were heated at various temperatures for 15 min, cooled to 30 °C, and assayed for CPP-phosphatase activity, the threshold temperature causing a significant decrease in activity of the thylakoid membranes was 60 °C. Incubation of membranes at this temperature caused a rapid loss of CPP-phosphatase activity to less than 5% of pre-incubation levels within 15 min (Figure 1). The 60 °C temperature treatment did not, however, cause any visible aggregation of the thylakoid membranes. To examine whether heat inactivation of phosphatase activity was similar when the enzyme was removed from the thylakoid membranes, detergent solubilized membrane phosphatase and the stromal phosphatase were also tested. Surprisingly, when removed from the membrane prior to the heating treatment, the membrane phosphatase did not show similar inactivation of CPP-phosphatase activity, and was inactivated less than 50% after a 15 min incubation (Figure 1). However, CPP-phosphatase activity of the stromal enzyme was reduced to 9% of pre-incubation levels af-

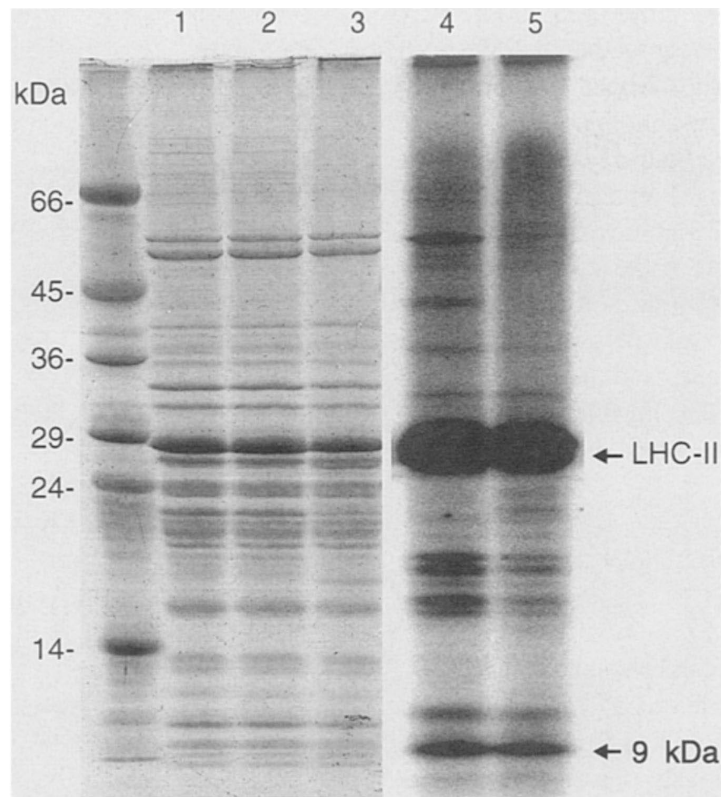


Figure 2. Effects of phosphorylation and 60 ° C temperature treatment on protein profiles (lanes 1-3) and ^{32}P incorporation (lanes 4-5) following SDS-PAGE. Thylakoid membranes (lane 1) were phosphorylated with the endogenous kinase in the presence of $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ (lane 2) then incubated at 60 ° C for 30 min (lane 3). Lanes 4 and 5 are a phosphorimage of lanes 2 and 3, respectively. The position of the LHC-II proteins and the 9 kDa protein are shown with arrows.

ter 10 min at 60 ° C (Figure 1). Detergent liberation of the membrane phosphatase may have caused some conformational or structural changes to the enzyme that made it less sensitive to heat denaturation. Heat-treated membranes were also examined for endogenous protein kinase activity. Following incubation at 60 ° C for 30 min, thylakoid membrane proteins were not phosphorylated under conditions optimal for kinase activity (data not shown).

Once it was established that the protein phosphatase activity endogenous to the thylakoid membrane could be inactivated over 95% following incubation at 60 ° C, the effects of heating on the phosphoproteins were examined. When the thylakoid membranes (Figure 2, lane 1) were phosphorylated by the endogenous kinase by adding $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ (Figure 2, lane 2) and then incubated at 60 ° C (Fig 2, lane 3), there was little change in the profile of Coomassie brilliant blue-stained proteins separated using SDS-PAGE. The addition of $[\gamma\text{-}^{32}\text{P}]\text{ATP}$ to unheated thylakoid membranes caused a number of

thylakoid membranes to become phosphorylated as visualized using phosphorimage analysis (Figure 2, lane 4). At least eleven proteins were noticeably phosphorylated, with the LHC-II apoproteins at 26 and 27 kDa, and the 9 kDa protein exhibiting the greatest amount of ^{32}P incorporation. A similar pattern of thylakoid membrane phosphorylation as been shown elsewhere (Markwell et al. 1984; Silverstein et al. 1993; Cheng et al. 1994). Heat treatment of the thylakoid membranes, in the presence of NaF and EDTA, did not cause a significant loss of ^{32}P from the LHC-II and the 9 kDa protein, while some minor thylakoid phosphoproteins did show a loss of radioactive label (Figure 2, lane 5). The dephosphorylation of some of these proteins may be due to an initial stimulation of the membrane phosphatase or an increase in membrane fluidity during heating. Since the LHC-II remained highly labeled following heat treatment, the validity of heat-inactivated thylakoid phosphomembranes as a substrate for exogenous LHC-II phosphatases was explored.

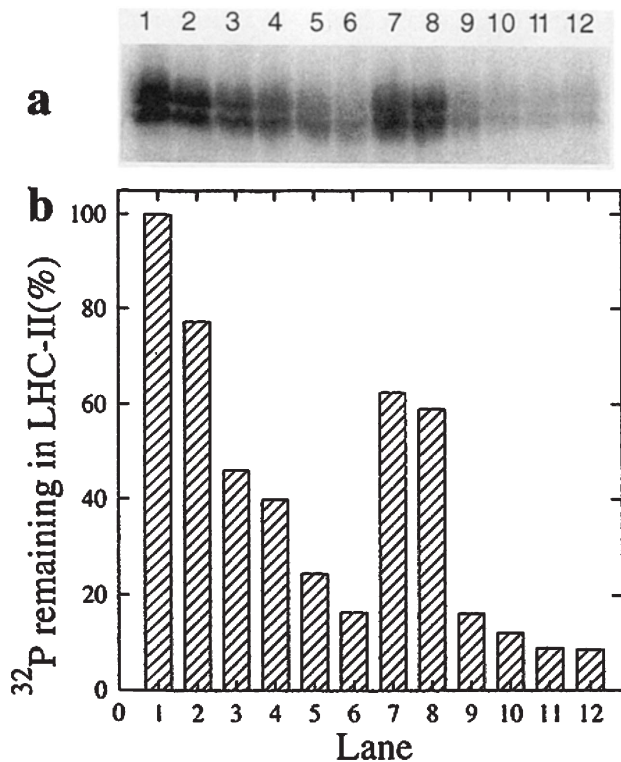


Figure 3. Dephosphorylation of the LHC-II in heat-inactivated thylakoid phosphomembranes following incubation with the membrane phosphatase (MP, lanes 1-6) or stromal phosphatase (SP, lanes 7-12). The phosphorimage of the SDS-PAGE separation (a) was integrated to measure ^{32}P labeling of LHC-II (b). Lanes 1 and 7 are at the start of exogenous phosphatase addition; lanes 2 and 8 are control reactions incubated for 120 minutes in the absence of exogenous enzyme. Times of incubation with the exogenous phosphatase were 15 min (lanes 3 and 9), 30 min (lanes 4 and 10), 60 min (lanes 5 and 11) and 120 min (lanes 6 and 12).

Heat-inactivated phosphomembranes as substrate for exogenous LHC-II phosphatases

The dephosphorylation of heat-inactivated phosphomembranes was examined using protein phosphatase activities associated with the stromal compartment (SP) or liberated from the thylakoid membranes by detergent treatment (MP). These phosphatases originated from Percoll-gradient purified pea chloroplasts. The phosphatases were further fractionated using Sephacryl S-200 chromatography. The MP and SP have similar elution profiles from gel permeation chromatography, but have been found to differ in their pH optima and response to molybdate ions (Hammer et al. 1995).

When MP or SP, having an equivalent CPP-phosphatase activity, were incubated with heat-inactivated phosphomembranes for various lengths of time, de-

phosphorylation of the LHC-II was apparent (Figure 3). In contrast, there was limited dephosphorylation of the LHC-II after 120 min. incubation at 30 °C in the absence of exogenous phosphatase (Figure 3, lane 2 and 8). Following 120 minutes of incubation, the MP dephosphorylated 83% of LHC-II associated ^{32}P while the SP dephosphorylated 86% of the LHC-II label, compared to the zero-time value for each enzyme fraction. Even though the Coomassie brilliant blue-stained gel indicated that all the lanes had similar amounts of LHC-II protein, there was a difference in LHC-II labeling intensities at time zero of the experiment (Figure 3b, lanes 1 and 7). This difference may be due to a more rapid dephosphorylation by the SP before the reaction could be stopped with the addition of denaturing sample buffer.

With longer phosphorimage exposures, the 9 kDa phosphoprotein showed a similar pattern of dephosphorylation as compared to that of the LHC-II (data not shown). Dephosphorylation was observed in other thylakoid membrane phosphoproteins but was difficult to quantify due to their diminished isotope labeling following the heat treatment (Figure 2, lane 5).

Validation of heat-inactivated thylakoid phosphomembranes as substrates for chloroplastic protein phosphatase, and demonstration that the SP will dephosphorylate thylakoid membrane associated LHC-II, allowed for the further characterization of this stromal activity.

Characterization of stromal LHC-II-phosphatase activity

Stromal LHC-II-phosphatase activity was characterized using heat-inactivated phosphomembranes as substrate. Following the dephosphorylation reaction, aliquots were separated on SDS-PAGE, visualized with the phosphorimager, and the ^{32}P associated with the LHC-II quantified.

Significant SP-catalyzed dephosphorylation of LHC-II was observed at pH values from 7.0 to 9.0, with a dephosphorylation optimum at pH 9. This was consistent with the CPP-phosphatase activity of the SP in response to assay pH (Hammer et al. 1995).

As previously reported for SP CPP-phosphatase activity (Hammer et al. 1995), dephosphorylation of LHC-II by SP was not inhibited by 10 mM molybdate ions but was partially inhibited by 10 mM EDTA. Additionally, 10 mM vanadate or tungstate ions were also found not to inhibit LHC-II dephosphorylation (data not shown).

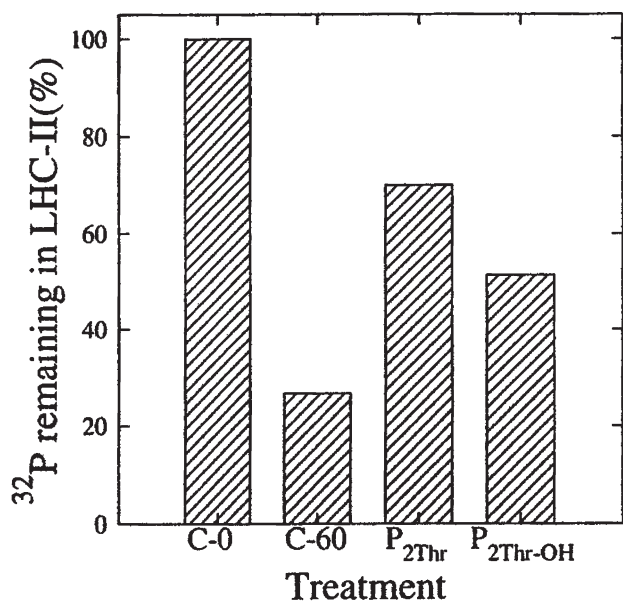


Figure 4. Effect of synthetic peptides on dephosphorylation of phospho-LHC-II by the stromal phosphatase fraction. Heat-inactivated thylakoid phosphomembranes were incubated with the stromal phosphatase for 0 or 60 min (C-0 or C-60, respectively) or incubated with the stromal phosphatase for 60 min in the presence of 7 μ M phosphorylated (P_{2Thr}) or unphosphorylated (P_{2Thr-OH}) peptides. Following incubation, samples were separated on SDS-PAGE, and the amount of ³²P labeling in LHC-II was measured by phosphorimaging.

To provide further evidence for the specificity of the stromal phosphatase, dephosphorylation of the heat-inactivated phosphomembranes was examined with the addition of synthetic phosphopeptides. The dephosphorylation of LHC-II by SP was inhibited by the phosphorylated synthetic peptide P_{2Thr}, an analog of the LHC-II phosphorylation site (Figure 4). Following a 60 minute reaction time, the control SP reaction dephosphorylated LHC-II by 73%, while with the addition of 7 μ M P_{2Thr} the LHC-II was dephosphorylated only by 30%. The addition of the non-phosphorylated peptide (P_{2Thr-OH}) did not cause the same extent of inhibition of dephosphorylation as the phosphorylated peptide (Figure 4). Similarly, in intact membranes, the endogenous membrane protein phosphatase is inhibited by the addition of a phosphopeptide analogues of the LHC-II N-terminus (Sun et al. 1993; Cheng et al. 1994).

Conclusions

Heat-inactivated phosphomembranes appear to be a suitable substrate with which to assay the ability of enzymes to dephosphorylate authentic LHC-II. The use

of such substrate, without significant endogenous protein phosphatase activity, is an important step to begin the isolation of LHC-II phosphatases from the thylakoid membrane. The use of such substrate has also allowed for the examination of potential LHC-II phosphatases not associated with the thylakoid membrane. The stromal fraction of isolated chloroplasts contains a protein phosphatase activity that is active on LHC-II in the membrane, and could be capable of LHC-II dephosphorylation *in vivo*. The total LHC-II peptide phosphatase activity in the stroma has been shown to be of equal magnitude to that of the thylakoid membrane (Hammer et al. 1995), indicating that the stromal activity is a significant component in the chloroplast. With the model of LHC-II phosphorylation and dephosphorylation currently proposed (Allen 1992), the LHC-II migrates from granal thylakoids to stromal-exposed thylakoids following phosphorylation (State 1 to State 2). A membrane-bound LHC-II phosphatase catalyzes the dephosphorylation which causes the LHC-II to migrate back to the granal thylakoids (State 2 to State 1). Would a stromal LHC-II phosphatase be additionally needed to provide more dynamic access to phosphorylated LHC-II? The exact role of a potential stromal LHC-II phosphatase is not known, but this activity should be considered in further investigations of LHC-II phosphorylation and dephosphorylation.

Acknowledgments

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