12-1-2003

A New Species of *Mathevotaenia* (Cestoda: Anoplocephalidae) and Other Tapeworms from Marsupials in Argentina

Scott Lyell Gardner  
*University of Nebraska - Lincoln*, slg@unl.edu

Mariel L. Campbell  
*University of New Mexico*, campbell@sevilleta.unm.edu

Graciela T. Navone  
*Centro de Estudios Parasitologicos y de Vectores*, gnavone@cepave.edu.ar

Follow this and additional works at: [http://digitalcommons.unl.edu/parasitologyfacpubs](http://digitalcommons.unl.edu/parasitologyfacpubs)  
Part of the *Parasitology Commons*

Gardner, Scott Lyell; Campbell, Mariel L.; and Navone, Graciela T., "A New Species of *Mathevotaenia* (Cestoda: Anoplocephalidae) and Other Tapeworms from Marsupials in Argentina" (2003). *Faculty Publications from the Harold W. Manter Laboratory of Parasitology*. Paper 20.  
[http://digitalcommons.unl.edu/parasitologyfacpubs/20](http://digitalcommons.unl.edu/parasitologyfacpubs/20)

This Article is brought to you for free and open access by the Parasitology, Harold W. Manter Laboratory of at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Faculty Publications from the Harold W. Manter Laboratory of Parasitology by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.
A NEW SPECIES OF MATHEVOTAENIA (CESTODA: ANOPLOCEPHALIDAE) AND OTHER TAPEWORMS FROM MARSUPIALS IN ARGENTINA

Mariel L. Campbell, Scott Lyell Gardner*, and Graciela T. Navone†

Museum of Southwestern Biology, Division of Mammals, University of New Mexico, Albuquerque, New Mexico 87131. e-mail: slg@unl.edu

ABSTRACT: Cestodes are reported from Didelphis albiventris Lund, 1840 and Micoureus cinereus Temminck, 1824 (Marsupialia: Didelphidae) in Argentina. These include a new species of Mathevotaenia Akhymian, 1946 (Cestoda: Anoplocephalata) as well as M. bivittata (Janicki, 1904) and an unknown hymenolepidid cestode. Mathevotaenia argentinensis n. sp. is characterized by a relatively narrow strobila, 18–37 mm in total length and 1.0–1.5 mm in maximum width, 135–163 craspedote proglottids, 19–27 testes, and a muscular genital atrium. This species differs from M. didelphidis (Rudolph, 1819) in the disposition of the genital ducts between the excretory canals and in the entrance of the vagina into the genital atrium posterior to the cirrus pouch; and from M. boliviana Schmidt and Martin, 1978 in the disposition of the genital ducts, absence of a seminal receptacle, and presence of an armed cirrus; and from M. boliviana Sawada and Harada, 1986 and M. pennsylvanica Chandler and Melvin, 1951 in the presence of an armed cirrus. Linstowinelines appear to be the dominant cestodes in New World marsupials, with M. bivittata representing the most prevalent and widely distributed species. The hymenolepidid is the first record of this family in Neotropical marsupials.

Materials and methods

Collection localities

Marsupials were collected from Moreno, Santiago del Estero Province, and from Orán, Salta Province, Argentina, by G.T.N. Moreno is located in the Chaco biogeographic province (Cabrera and Willink, 1973). The Chacoan thorn scrub plant community type occurs in semi-arid areas with a marked dry season between May and October and mean annual temperature is 21±23 C. Marsupials infected with cestodes were collected in a transition zone between Yungas and Chacoan vegetation.

Methods

Cestodes were preserved in cold 70% alcohol and were not relaxed before fixation. Specimens were stained in Grenacher’s acetic carmine, dehydrated in ethanol, cleared in terpineol and xylene, and mounted in Canada balsam. Drawings were made using a Leitz Ortholux II microscope with a drawing tube. Measurements and observations were made from whole mounts using a Zeiss Ultraphot II microscope with a drawing tube. Measurements and observations were made from whole mounts using a Zeiss Ultraphot II. Measurements are given in micrometers unless otherwise indicated; ranges are followed by means in parentheses. The holotype was deposited in the Colección Helminológica del Departamento Científico de Invertebrados (CHDCI), La Plata, Argentina; paratypes were deposited in the CHDCI and in the Harold W. Manter Laboratory of Parasitology, University of Nebraska–Lincoln, Lincoln, Nebraska. Mammalian symbiont (see Frey et al., 1992), when preserved, were deposited in the División Mastozoología, Museo Mar del Plata (MMPMA), Mar del Plata, Argentina.

RESULTS

Mathevotaenia argentinensis n. sp.

(Figs. 1–6)

Description

Relatively small cestodes, 18–37 mm (28 mm, n = 3) in total length, consisting of 135–163 (149) craspedote proglottids. Maximum width 1,000–1,500 (1,300) attained in gravid segments just past midline. Scolex unarmed, poorly demarcated from strobila, 305–462 (387) long by 392–536 (452) wide. Suckers oval, with thin muscular walls, 222–256 (235) wide by 152–197 (169) long, each recessed into a pocket opening externally through a longitudinal slit down sucker midline. Opening at sucker anterior margin highly muscular and frequently constricted, so that sucker margin appears incomplete (Fig. 1). Neck 1,100–1,200 long by 320–370 wide; in some specimens, segmentation beginning immediately behind scolex. Developing genitalia first visible in 28th proglottid, 2–4 mm behind scolex; testes first visible in proglottid 64, 4–6 mm behind scolex. First mature proglottid approximately segment 58–70; first gravid proglottid segments 86–102. Strobila very slender for first one-fourth to one-third of the total length, increasing rapidly in relative width beginning in late immature proglottids. Immature proglottids much wider than long, length–width ratio of 1:4 to 1:7.5. Mature proglottids (Figs. 2, 5) 220–260 long...
by 780–1,000 wide. Gravid proglottids (Figs. 3, 4) 240–350 long by 1,080–1,490 wide at maximum width of strobila. Terminal gravid proglottids 640–1,330 long by 570–650 wide, barrel-shaped, packed with eggs. Female genitalia and testes involute in gravid proglottids, beginning in proglottids 100–110, where only cirrus pouch and sperm-filled fertilization duct are visible. Osmoregulatory system consisting of paired lateral canals, with narrow dorsal duct and broader ventral duct on each side. Genital ducts pass between osmoregulatory canals. Testes 19–29, in single field posterior and lateral to vitelline gland, reaching the posterior margin of ovary. Testes in poorly relaxed specimens irregular; in relaxed specimens oval to subspherical, 37–70 (53) long by 25–39 (34) wide. Cirrus pouch elongate, pyriform, with clear, thick external wall, 120–183 (142) long by 45–53 (50) wide, narrowing markedly at proximal end where vas deferens enters. Lateral osmoregulatory canals crossing cirrus pouch midpoint. Cirrus pouch extending medially to poral lobe of ovary, not reaching proglottis midline. Cirrus thick, highly coiled, armed with large spines readily visible at ×10. Prostate cells visible at distal region of cirrus pouch. Internal and external seminal vesicles absent. Vas deferens forms highly coiled glandular duct anterior to ovary. Genital atrium pronounced, sphincterlike, 48–84 (63) deep by 5–24 (15) wide, encircled by strong bands of circular and radial musculature. Genital pores alternate irregularly, opening near anterior margin of proglottid. Ovary 90–168 (118) long by 286–324 (299) wide, medial, in anterior half of proglottid, bilobed, lobulate, with 10–13 horizontal digitiform lobules on each side. Lobes of ovary not quite reaching lateral osmoregulatory canals. Vitelline gland medial, compact, in posterior half of proglottid, 72–96 (84) long by 126–163 (139) wide, with approximately 8 small rounded lobules oriented posterolaterally. Thin vaginal duct expanding slightly and becoming glandularized opposite poral lobe of ovary. Seminal receptacle absent. Oviduct a thick sinusous tube arising from ventral surface of ovarian isthmus, initially directed posteriorly, then curving dorsally and anteriod, receiving narrow vagina and becoming fertilization duct. Broad, sperm-filled fertilization duct curves posteriorly entering Mehlis’ gland dorsal and anterior to vitelline gland. Vitelline duct arising from dorsal surface of vitelline gland, entering posterior border of Mehlis’ gland and fusing with fertilization canal. Uterine duct passing forward from Mehlis’ gland dorsal to ovary and vagina, descending into uterus anterolateral to ovary. Eggs initially appearing clustered in uterine pouches along ventral surface of the medulla, with maturing eggs becoming distributed singly in egg capsules throughout medulla and into cortical parenchyma; late gravid proglottids appearing as thin-walled sacs bulging with eggs (Fig. 4). Onchospheres (Fig. 6) (frequently deformed because of clearing) 26–32 (29) wide by 15–23 (18) long, surrounded by wrinkled envelope. Outer shell darkly staining, 44–53 (49) wide by 30–49 (38) long. Egg capsules 63–74 (69) long by 55–62 (59) wide. Embryonic hooks 14.3–15 (14.7).

### Taxonomic summary

**Type host:** *Didelphis albiventris* Lund, 1840 (2 individuals).

**Site of infection:** Posterior half to one-third of the small intestine.

**Prevalence:** One hundred percent.

**Intensity:** Twenty to 40 worms per individual opossum studied.

**Type locality:** Moreno, Provincia Santiago del Estero, Argentina; 27°12′30″S, 63°02′30″W.

**Specimens deposited:** One Holotype, slide no. II 200 A, (CHDCI5283) and 3 paratypes, slide number C II 200 B, (CHDCI5284), slide no. C 180 A, (HWML17711) and slide no. C 180 B, (HWML17725).

**Etymology:** The species name refers to Argentina (*argentensis* = of Argentina), the country in which it was found.

### Diagnosis

Of the species of *Mathevotaenia* reported previously from neotropical marsupials, *M. argentinensis* n. sp. is most similar to *M. didelphidis* (Rudolphi, 1819) in having approximately 20 testes but differs in the following characteristics: (1) genital ducts pass between rather than dorsal to the excretory canals, (2) the cirrus pouch is oriented transversely rather than anteriorly, and (3) the vagina enters the genital atrium posterior rather than anterior to the cirrus pouch.

Flores-Barroeta et al. (1961) described cestodes from marsupials in Costa Rica, which they referred to *Mathevotaenia* (syn. *Oochoristica*) *didelphis* (Rudolphi, 1819); however, their specimens differ from the original description of *M. didelphidis* in the characteristics of the cirrus pouch and vagina mentioned above and probably represent a distinct species. *Mathevotaenia argentinensis* differs from these Costa Rican specimens in having a much smaller scolex (0.3 by 0.4 mm vs. 1.2 mm in diameter), a much longer cirrus pouch (120–180 vs. 70–96) and, most importantly, mature eggs that are distributed evenly throughout the proglottids rather than concentrated along the lateral margins.

Flores-Barroeta et al. (1961) reported cestodes with 19–36 testes from *D. marsupialis* in Costa Rica and referred the species to *M. pennsylvanica*, originally described by Chandler and Melvin (1951) from North American shrews. *Mathevotaenia argentinensis* differs from these Costa Rican cestodes in having a much smaller scolex (387 by 452 vs. 760 by 1,170), a longer cirrus sac (120–180 vs. 660–1,060), an armed versus unarmed cirrus, and genital ducts passing between rather than dorsal to the osmoregulatory canals. Both *M. argentinensis* and the Costa Rican specimens described by Flores-Barroeta et al. (1961) differ from the original description of *M. pennsylvanica* by Chandler and Melvin (1951) in possessing a smaller strobila, fewer testes, testes distributed in a single field posterior to the female genitalia rather than in 2 lateral fields, a bilobed versus reniform ovary, and gravid segments that are longer than wide versus wider than long.

South American species of *Mathevotaenia* occurring in mammals other than marsupials and possessing only 20–30 testes include *M. paraguayae* Schmidt and Martin, 1978 from a South American armadillo and *M. boliviana* Sawada and Harada, 1986 from Bolivian bats. *Mathevotaenia argentinensis* differs from *M. paraguayae* in possessing genital ducts that extend between rather than dorsal to the osmoregulatory canals, by the absence of a seminal receptacle, and in having an armed cirrus highly coiled within the cirrus pouch. *Mathevotaenia argentinensis* is distinguished from *M. boliviana* by an armed versus unarmed cirrus, a much larger strobila, and a smaller scolex.
Several individuals of *M. argentinensis* did not exceed 2–4 mm in length, appearing to have accelerated maturation of the strobila such that the first mature and first gravid proglottids occurred in segment numbers 23–32 and 30, respectively, of a total of 45 proglottids. In these specimens, the scolex measured 248 long by 284 wide with suckers 135 long by 103 wide, and the maximum width of the strobila was 119 to 162. These specimens occurred in the same host individual as the type specimens and appear to be a forma minor of *M. argentinensis* (see discussion under *M. bivittata*).

**REDESCRIPTION**

*Melcaevonoea bivittata* (Janicki, 1904) Yamaguti, 1959

(Figs. 7–9)

**Synonyms:** Oochochistica bivittata Janicki, 1904, 772; Linstowia [Opossumia] bivittata (Janicki, 1904) Spasskii, 1951, 498, figure 225; Opossumia bivittata Spasskii, 1981.


Small cestodes, 13–15 mm in length by 960–1,350 in maximum width (n = 10). Maximum width in gravid proglottids in the posterior half or quarter of the strobila. Scolex (Fig. 7) small, 360–390 long (measured to sucker posterior margin) by 460–600 wide, not globular or distinctly demarcated from strobila; width of strobila immediately behind scolex 280–620. Suckers 225–291 long by 201–228 wide. Sucker pockets present. Neck present or absent; genital anlagen visible immediately behind scolex or within 0.2–0.4 mm. Thick bands of longitudinal muscle visible in whole mounts originating in scolex and running length of strobila. Proglottids craspedote, numbering 37–49, trapezoidal with laterally expanded posterior edges. Immature proglottids wider than long, length–width ratio of approximately 1:7. Mature proglottids (Fig. 9) wider than long in unrelaxed specimens, 194–197 by 400–601 (501); length–width ratio of 1:1.5–1:3.1. First mature proglottid segment 21–23; first gravid proglottid segment 24–28. Gravid proglottids (Fig. 8) wider than long to longer than wide; length–width ratio of 1:2.7 in early gravid proglottids to 1:0.4–1:1.4 in terminal gravid proglottids, measuring 400–900 long by 600–900 wide. Genital atrium small, 24–43 deep by 5–6 wide. Genital pores irregularly alternating, opening along anterior one-fourth of the lateral margin of proglottid. Cirrus pouch small, 71–108 by 34–49, appearing as rounded pouch not quite reaching lateral osmoregulatory canals. Vas deferens forming numerous loops and convolutions medial to cirrus pouch, anterior and portal to ovary. Testes 10–13, posterior to female genitalia at midline, sub-spherical, 23–31 long by 30–41 wide. Vagina opening into genital atrium posterior to cirrus pouch. Details of the vagina, including the presence of a seminal receptacle, were not distinguishable. The disposition of the genital ducts relative to the osmoregulatory canals was unclear in both whole mounts and sections because of the deformation of the strobila. Ovary 98–209 × 48–123, medial, in anterior half of proglottid, not bilobed; a total of 10–15 short lobules are present oriented both dorsoventrally and anteroposteriorly. Vitelline gland compact, medial, 42–105 by 31–74, posterior to ovary, slightly lobed. Mature eggs (visible as darkly staining onchospheres) concentrated along the lateral margins of the proglottids and embedded in the cortical parenchyma (Figs. 8, 10). Female genitalia persistent throughout strobila, visible in the relatively clear area at the center of gravid proglottids. Terminal gravid proglottids contain very few eggs. Onchosphere 19–23 by 15–17; darkly staining outer shell 26 by 23; egg capsule 31–41 by 23–29. Hooks 11.8 in length.

**Taxonomic summary**

**Hosts:** *Micoureus cinereus* Temminck, 1824, symbiotypes MMPMa 2524 and 2525.

**Site of infection:** Small intestine.

**Intensity:** Eleven to 34 cestodes per host.

**Locality:** Orán, Salta Province, Argentina; 22°59′40″S, 64°22′05″W.

**Specimens:** (#24 a.e, and #25 b CHDCI5285) and (#24 B, #25 A HWML17712).

**Remarks**

These specimens are similar to the original description and illustrations given in Janicki (1904), although the number of testes is greater than 5–8 and the strobila is much smaller than 55 mm. Baer (1927a) reported individuals of this species from Brazil not exceeding 10 mm in length, which he referred to as a forma minor. Gomes (1979) also described small (4–6 mm) yet reproductively mature individuals with 8–10 testes from Brazil, observing that smaller individuals tended to occur in hosts infected with large numbers of cestodes. All specimens examined in this study were taken from hosts infected with an average of 23 individuals, lending some support to Gomes’ hypothesis that small body size in these cestodes may result from a “crowding effect” in heavy infections. However, we cannot rule out the possibility that these size differences may reflect underlying population or species-level genetic differences.

**ADDITIONAL OBSERVATIONS**

Three strobilar fragments of an unknown hymenolepidid cestode (specimen #33, HWML17726) in poor condition were recovered from the small intestine of a single individual *M. cinereus* Temminck, 1824 (MMPMa 2533) from Orán, Salta Province, Argentina. These fragments appear to represent 3 individual cestodes, each lacking the scolex.

The only other report of a hymenolepidid from New World marsupials is a record of *Hymenolepis sp.* from *D. virginiana virginiana* from North America (Leigh, 1940). Leigh (1940) did not describe the morphology, noting only that “hooks were gone from rostella” (p. 192). It is possible that both the above reports represent accidental infections; however, several unique species of hymenolepis have been reported from marsupials in Australia. The Australian species include *H. peramelidarum* Nybelin, 1917, *H. bradleyi* Beveridge & Barker, 1975, *H. aklei* Beveridge & Barker, 1975, *H. cercarteti* Vaucher et al., 1984, *H. antechini* Vaucher et al., 1984, *H. bettongiae* Vaucher et al., 1984, *H. isoodonitis* Vaucher et al., 1984, *H. potoroi* Vaucher et al., 1984, and *H. peroryctis* Jones & Anderson, 1990 (see...
Vaucher et al., 1984). These species were transferred into the genus *Vampirolepis* Spasskii, 1954 by Jones and Anderson (1990) because of the presence of an armed rostellum. However, Vaucher (1992) rejected this transfer, suggesting that the hymenolepids of marsupials belong in their own genus (p. 301). Despite the lack of a scolex available for comparison, the unknown species from Argentina closely resembles *H. cercarteti* in the arrangement of the genitalia. The possibility that marsupials in the neotropics and in Australasia share similar species of hymenolepidid cestodes awaits verification.

ACKNOWLEDGMENTS

This work was funded in part by United States National Science Foundation grants DEB-9496263, BSR-9024816, and DEB-0097019 to S.L.G.

LITERATURE CITED


**BEVERIDGE, I., AND I. K. BARKER.** 1975. Acuariid, capillariid and hydella santeri. **DELLA SANTA, E.** 1956. Revision du genre *Vampirolepis* Vaucher et al., 1984). These species were transferred into the genus *Vampirolepis* Spasskii, 1954 by Jones and Anderson (1990) because of the presence of an armed rostellum. However, Vaucher (1992) rejected this transfer, suggesting that the hymenolepids of marsupials belong in their own genus (p. 301). Despite the lack of a scolex available for comparison, the unknown species from Argentina closely resembles *H. cercarteti* in the arrangement of the genitalia. The possibility that marsupials in the neotropics and in Australasia share similar species of hymenolepidid cestodes awaits verification.

ACKNOWLEDGMENTS

This work was funded in part by United States National Science Foundation grants DEB-9496263, BSR-9024816, and DEB-0097019 to S.L.G.

LITERATURE CITED


