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Diarrhea associated with *Enterococcus durans* in calves

Douglas G. Rogers, David H. Zeman, E. Denis Erickson

Enterococci in the alimentary tract of animals are believed to be commensal bacteria. However, *Enterococcus durans* has been associated with diarrhea in foals, piglets, and puppies. The possibility that *E. durans* is associated with diarrhea in calves has not been established, although gram-positive cocci adhered to villi, occasionally together with Cryptosporidia, have been seen histologically in the small intestine of diarrheic calves. This report describes the association of *E. durans* and diarrhea in 4 calves.

Samples from 3 1-3-week-old diarrheic mixed breed beef calves (calves 1, 2, and 3) were submitted for laboratory evaluation. The samples submitted varied with each case but included fresh and formalin-fixed jejunum (3), ileum (2), colon (1), abomasum (1), spleen (3), liver (2), and lung (1) and 1 liter of milky-white intestinal content (1). Calf 4 was a 1-week-old mixed breed beef calf that was presented dead for necropsy. The owner reported that this calf had died without clinical signs. At necropsy, the presence of watery

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Table 1. Results of laboratory evaluations of tissues from 4 diarrheic calves.

<table>
<thead>
<tr>
<th>Calf no.</th>
<th>Small intestine</th>
<th>Colon</th>
<th>Bacteriology*</th>
<th>Virology†</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>hyperemia, mild villus atrophy, moderate numbers gram-positive cocci adhered to villi, desquamated apical enterocytes, small numbers of neutrophils in lamina propria and crypts</td>
<td>no lesions</td>
<td>large numbers of <em>Enterococcus durans</em></td>
<td>negative</td>
</tr>
<tr>
<td>2</td>
<td>hyperemia, mild villus atrophy, moderate numbers of gram-positive cocci adhered to villi, desquamated apical enterocytes, small numbers of neutrophils in lamina propria and crypts</td>
<td>not submitted</td>
<td>large numbers of <em>E. durans</em></td>
<td>negative</td>
</tr>
<tr>
<td>3</td>
<td>hyperemia, mild villus atrophy, moderate numbers of gram-positive cocci adhered to villi</td>
<td>not submitted</td>
<td>large numbers of <em>E. durans</em></td>
<td>BVDV positive (small intestine); small numbers of hemolytic <em>Escherichia coli</em></td>
</tr>
<tr>
<td>4</td>
<td>hyperemia, moderate villus atrophy, large numbers gram-positive cocci and cryptosporidia adhered to villi, multifocal cryptal necrosis</td>
<td>multifocal cryptal necrosis</td>
<td>large numbers of <em>E. durans</em></td>
<td>BVDV positive, direct immunofluorescence (small intestine); rotavirus positive, electron microscopy (intestinal contents)</td>
</tr>
</tbody>
</table>

* Isolates from small intestine.
† Appropriate samples were evaluated for rotavirus, coronavirus, and bovine viral diarrhea virus (BVDV) by direct immunofluorescence and for BVDV by virus isolation; intestinal contents (when available) were evaluated for virus by electron microscopy.
‡ Brown-Brenn Gram stain.
ingesta in the distal jejunum, ileum, and colon was the most significant finding. Appropriate tissue samples from the 4 calves were processed for microscopic evaluation, aerobic and anaerobic bacteriologic culture, virologic evaluation using direct immunofluorescence, virus isolation, and, when feasible, electron microscopic examination of intestinal contents. In addition, samples of formalin-fixed small intestine from 1 calf were immersed in 3% glutaraldehyde in 0.1 M sodium phosphate buffer (pH 7.2) and processed for transmission electron microscopy.

Enterococcus durans was isolated in large numbers from the small intestine of each of the four calves (Table 1). Isolates of E. durans produced alpha hemolysis on 5% sheep blood agar plates and were identified by conventional and commercial methods. Enterococcus durans was considered to be strongly associated with diarrhea in calves 1, 2, and 3, and the histologic findings from these calves correlated with those cases involving foals. Bacteria were adhered to intestinal villi (Fig. 1). Ultrastructural examination of samples of small intestine from calf 1 revealed cocci adhered to or closely associated with microvilli on villus enterocytes. Cocci were 0.5-0.7 µm in diameter and often were surrounded by a rim of flocculent electron-dense material (Fig. 2). Detailed ultrastructural examination was not possible because of mild post-mortem decomposition in the samples. The significance of the isolation of bovine viral diarrhea virus (BVDV) and small numbers of hemolytic Escherichia coli from the small intestine of calf 3 could not be determined; there were no lesions suggestive of BVDV infection. The possibility that BVDV caused cryptal necrosis in the small intestine and colon from calf 4 was considered. The cause of diarrhea in calf 4 was attributed to multiple pathogens, including Elaterococcus durans.

The pathogenesis of diarrhea associated with E. durans adherence is unknown. The bacterium is believed to adhere to villus enterocytes by fimbriae-like structures. Isolates of E. durans from foals do not produce enterotoxin in the isolated loops of pig intestine but they do depress brush border enzyme activity, possibly causing malabsorption. It is unknown whether treatment with antibiotics predisposes the small intestine to adherence by E. durans. Calf 1 was the only calf known to have been treated with an antibiotic prior to death.

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Sources and manufacturers

a. Rapid Strep, API Analytical Products, Plainview, NY.

References