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Chloroviruses Encode a Bifunctional dCMP-dCTP Deaminase That Produces Two Key Intermediates in dTTP Formation

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The chlorovirus PBCV-1, like many large double-stranded DNA-containing viruses, contains several genes that encode putative proteins involved in nucleotide biosynthesis. This report describes the characterization of the PBCV-1 dCMP deaminase, which produces dUMP, a key intermediate in the synthesis of dTTP. As predicted, the recombinant protein has dCMP deaminase activity that is activated by dCTP and inhibited by dTTP. Unexpectedly, however, the viral enzyme also has dCTP deaminase activity, producing dUTP. Typically, these two reactions are catalyzed by proteins in separate enzyme classes; to our knowledge, this is the first example of a protein having both deaminase activities. Kinetic experiments established that (i) the PBCV-1 enzyme has a higher affinity for dCTP than for dCMP, (ii) dCTP serves as a positive heterotropic effector for the dCMP deaminase activity and a positive homotropic effector for the dCTP deaminase activity, and (iii) the enzymatic efficiency of the dCMP deaminase activity is about four times higher than that of the dCTP deaminase activity. Inhibitor studies suggest that the same active site is involved in both dCMP and dCTP deaminations. The discovery that the PBCV-1 dCMP deaminase has two activities, together with a previous report that the virus also encodes a functional dUTP triphosphatase (Y. Zhang, H. Moriyama, K. Homma, and J. L. Van Etten, J. Virol. 79:9945–9953, 2005), means that PBCV-1 is the first virus to encode enzymes involved in all three known pathways to form dUMP.
FIG. 1. The three known pathways for synthesis of DUMP, an intermediate in dTP synthesis. The enzymes shown in the diagram in bold italics are encoded by chlorovirus PBCV-1 and are known to be functional, except for the ribonucleotide reductase.

Center, New Haven, CT) and BL21(DE3)pLysS (Novagen, Madison, WI) were grown in Luria-Bertani (LB) medium (34).

Cloning and expression of the dCMP deaminase gene and purification of the enzyme. PBCV-1 open reading frame (ORF) A596R was cloned from PCR-amplified viral DNA by use of the following oligonucleotide primers: 5' primers 5'-GGTGTGCATATGCAAAAGCAGGAAG-3' and 5' TCCCT CGGATTGAGTTAGTGGTCTAT-3'. The 5' primer contained an NdeI restriction site, and the 3' primer contained an Xhol restriction site. The A596R gene was amplified with KOD Hot Start DNA polymerase (Novagen) in 50-μl reaction mixtures which contained 100 ng of virus DNA, 15 pmol of each primer, 0.2 mM each of dATP, dGTP, dCTP, and dTTP, and 1 mM MgSO4 by 35 cycles of heating and cooling (15 s at 94°C for denaturing, 30 s at 60°C for annealing, and 1 min at 68°C for elongation). The PCR products were purified from agarose gels by using a QIAEX II gel extraction kit (QIAGEN, Valencia, CA), digested with NdeI and XhoI, and inserted into the NdeI/XhoI sites of the pET23a+ expression vector (Novagen). This process produced a six-His tag at the C terminus of the target protein. The construction of the recombinant expression plasmid, named pET-dCD, was confirmed by DNA sequencing. Expression of the dCMP deaminase protein was carried out in E. coli BL21(DE3)/pLysS, which contains the T7 RNA polymerase gene under the control of the isopropyl-β-D-thiogalactopyranoside (IPTG)-inducible lacUV5 promoter and a plasmid, pLysS, constitutively expressing the T7 lysozyme that is the inhibitor of T7 RNA polymerase. Cells were transformed with plasmid pET-dCD and grown in LB medium at 37°C. Flasks containing 400 ml LB medium which had been supplemented with 0.5 mM IPTG (Sigma, St. Louis, MO) was added to a final concentration of 0.5 mM, and incubation was continued for 2 h until the absorbance at 595 nm reached 0.6 to 0.8. IPTG (Sigma, St. Louis, MO) was applied to the plates as standards. The plates were developed stepwise in the following solutions: (i) 1 M acetic acid for ~1 min, (ii) 0.9 M acetic acid and 0.3 M LiCl to 3 cm above the starting line, and (iii) 1 M acetic acid and 1.3 M LiCl to 14 cm above the starting line. After development, the plates were washed in 100% methanol for 5 min and dried with hot air, and the nucleotide spots were identified by UV light. The retardation factor values for dCTP, dCMP, dUMP, and dTMP were 0.3, 0.74, 0.83, and 0.63, respectively.

Inhibition assay of the enzyme activities. The inhibition reaction rates were monitored continuously with time by determining the decrease in absorbance at 290 nm after addition of enzyme at 30°C by use of a Gilford 250 spectrophotometer, as described previously (20). For inhibition of dCMP deaminase activity, the reaction mixtures contained 1 mM dCMP, 50 mM CAPSO, pH 9.5, 1 mM MgCl2, 2 mM 2-mercaptotethanol, 0.1 mM dCMP, and 2.2 μg of the enzyme in a final volume of 1.0 ml. For inhibition of dCTP deaminase activity, the reaction mixtures contained 1 mM dCTP, 50 mM Tris-HCl, pH 7.0, 1 mM MgCl2, 2 mM 2-mercaptotethanol, and 2.2 μg of the enzyme in a final volume of 1.0 ml. Each inhibitor, pyrimidin-2-one deoxyribodyste (PDRP) (2) and H2dUMP (14), was present at concentrations from 2 μM to 10 μM.

Sedimentation velocity studies. The PBCV-1 enzyme in NPI-250 buffer was dialyzed against two changes of 50 mM pH 7.0 buffer, each containing 50 mM potassium phosphate, pH 7.5, 100 mM NaCl, and 2 mM dithiothreitol at 4°C and then diluted to an A290 of 0.32. The viscosity and density of the buffers and partial specific volume (Vps) of the protein were obtained from SEDNTERP software (http://www.jpihlo.mailway.com). The Vps of the enzyme was calculated from the amino acid content to be 0.7313 ml/g at 25°C or 0.7291 ml/g at 20°C. The protein (0.4 ml) and 0.42 ml dialysis buffer were centrifuged at 50,000 rpm in 12-mm Epon carbon-filled double-sector centerpieces in a Beckman XL-1 analytical ultracentrifuge and an An-60 Ti rotor at 25°C. Absorption measurements were made at 280 nm. A single sample was run with zero time between scans; the R20w was set at 6.0, and the samples were scanned from the earliest time until the boundaries were at the cell bottom so as to obtain a large number of scans. Scans were set at a spacing of 0.003-cm radial step size in a continuous scan mode. Care was taken to have the samples at thermal equilibrium before starting. The centrifuge was accelerated directly to the speed of the experiment. The data were analyzed using the c(s) method found in SEDFIT, a program developed by P. Schuck (http://www.analyticalultracentrifugation.com). The experimentally calculated sedimentation coefficients from the SEDFIT program were converted to s20w values by use of the transform within the SEDFIT software.

Northern and dot blot analyses. Chlorella cells (1 × 106 cells) were collected at various times after PBCV-1 infection, frozen in liquid nitrogen, and stored at −80°C. RNA was extracted with TRIzol reagent and guanidinium vanillin, Carlsbad, CA), denatured with formaldehyde, separated on a 12% agarose denaturing gel, and transferred to nylon membranes (Micron Separations, Inc., Westborough, MA), as described previously (10). The membrane was subsequently photographed under UV illumination to visualize transferred RNA. The RNA was hybridized mixing. Enzyme activity was monitored at A290 with 5-s acquisition intervals in a temperature-equilibrated reaction chamber by use of a Peltier temperature control module (Beckman).
with a double-stranded \(^{32}\)P-A596R gene probe at 65°C in 50 mM sodium phosphate, 1% bovine serum albumin, and 2% sodium dodecyl sulfate (SDS), pH 7.2. The probe was labeled with \(^{32}\)P by use of a random-primer DNA labeling system (Invitrogen). After hybridization, radioactivity bound to the membranes was detected and quantified using a Storm 840 PhosphorImager and ImageQuant software (Molecular Dynamics, Inc., Sunnyvale, CA). To monitor possible loading differences between samples, the relative amount of the 3.6-kb rRNA in each lane was determined by converting the photographs of stained membranes to digital images with a Hewlett-Packard ScanJet 4C scanner and analyzing the images with ImageQuant software.

Viral DNAs used for dot blots were denatured, applied to nylon membranes fixed by UV cross-linker, and hybridized with the same probes used for the Northern analysis.

Phylogenetic analyses. A BLASTP search with the amino acid sequence of PBCV-1 A596R (NCBI NP_048952) was conducted using default settings in the Biology Workbench (http://workbench.sdsc.edu). Twenty-one taxa, including archaea, prokaryotes, and eukaryotes, as well as several viruses, were selected. The 22 taxa, which included PBCV-1, were aligned with ClustalW by use of the default setting in the Biology Workbench. The alignment was changed to NEXUS format and imported into PAUP 4.0b10 (36) for phylogenetic analysis. Trees were constructed using the following three methods: maximum-parsimony heuristic, neighbor joining, and maximum-parsimony bootstrap (1,000 replicates). The DNA binding protein from Streptococcus thermophilus (NCBI YP_142074), which has some sequence common to dCMP deaminase, was used as the outgroup to root the trees.

Other procedures. DNA and putative protein sequences were analyzed with the University of Wisconsin Computer Group version 10.1 package of programs (Genetics Computer Group, 2000).

RESULTS

PBCV-1 ORF A596R encodes a dCMP deaminase. Chlorella virus PBCV-1 encodes a 142-codon ORF (A596R) that has 30 to 40% amino acid identity with dCMP deaminases from eukaryotes, prokaryotes, and T4 phage. The predicted dCMP deaminase has an inferred molecular mass of \( \sim 16 \text{ kDa} \). Like dCMP deaminases from other organisms, A596R contains a conserved domain that includes a putative zinc-binding domain, an allosteric regulator binding site, and residues that form the catalytic site (1, 22, 26) (Fig. 2).

FIG. 2. Amino acid alignment of dCMP deaminases. The amino acid sequences from human (P32321), Lactococcus lactis (E86767), Pyrococcus abyssi (CAB49495), Saccharomyces cerevisiae (S46762), T4 phage (AAD42546), and PBCV-1 (O41078) were aligned by use of the Wisconsin GCG Pileup program. (Accession numbers are from the NCBI database.) The most conserved domain of dCMP deaminases, which forms the catalytic site, is boxed. The amino acid residues indicated with a solid square correspond to one zinc ion-binding site of T4 phage dCMP deaminase; the amino acid residues indicated with a solid inverted triangle correspond to the regulator (dCTP and dTTP) binding site of T4 phage dCMP deaminase. A black box around an individual residue indicates identity; a gray box indicates similarity.
protein could be stored for at least 1 year in 50% glycerol without precipitation. As described below, the isolated protein deaminates dCMP to dUMP and, unexpectedly, the protein also deaminates dCTP to dUTP. Therefore, we named the protein bifunctional dCMP-dCTP deaminase (bi-DCD). Elution of the His-tagged recombinant protein produced a single Coomassie brilliant blue staining band on SDS-polyacrylamide gels (Fig. 3A), which contained both the dCMP and the dCTP deaminase activities. Empty vector control cells (pET-23a) produced no measurable dCMP or dCTP deaminase activity.

Typically, dCMP deaminases function as hexamers (1), and a sedimentation velocity analysis yielded an $S_{20,w}$ of 6.06 for bi-DCD (results not shown). Based on a subunit mass of 17 kDa, the PBCV-1 enzyme sedimented as a hexamer.

Characterization of the bi-DCD dCMP deaminase activity. As shown in Fig. 3B, lane 2, TLC experiments established that bi-DCD deaminates dCMP to dUMP. Characterization of the enzyme using dCMP as the substrate was conducted at different temperatures, pHs, and cations. The enzyme is active from 25°C to 55°C, with an optimum of 42°C, which is higher than the 25°C optimum temperature for growing the host and the virus. The enzyme is active from pH 6.0 to 11.0, with an optimal pH at 9.5. The enzyme prefers Mg$^{2+}$ but retains some activity in the presence of Ca$^{2+}$, Mn$^{2+}$, and Ni$^{2+}$. Although the protein contains a putative zinc-binding site, its enzymatic activity in Zn$^{2+}$ is less than 6% of the activity obtained with Mg$^{2+}$. Assays for Zn$^{2+}$ carried out using the metallochromic indicator 4-(2-pyridylazo) resorcinol, as described previously for T4 dCMP deaminase (27), revealed little or no Zn$^{2+}$ in the protein. Like activities of dCMP deaminases from other organisms (24), the bi-DCD dCMP deaminase activity is activated by dCTP and feedback inhibited by dTTP, which is the end product of dTTP synthesis. The activity increased ~7-fold in the presence of 0.005 to 0.1 mM dCTP (Fig. 4A) and was reduced ~12-fold in the presence of 1 mM dTTP (Fig. 4B). Without dCTP, the enzyme exhibited very weak activity at high concentrations of dCMP (results not shown).
Characterization of the bi-DCD dCTP deaminase activity. Unexpectedly, TLC revealed that bi-DCD also has dCTP deaminase activity that converts dCTP to dUTP (Fig. 3C, lane 2). Therefore, dCTP may serve as either a substrate or a positive heterotropic effector (or activator) of the enzyme. Like the dCMP deaminase activity, the dCTP deaminase activity was highest at 42°C and enzyme activity occurred from 25°C to 55°C. The enzyme prefers Mg\(^{2+}\) and retains some activity in the presence of Ca\(^{2+}\), Mn\(^{2+}\), or Ni\(^{2+}\); essentially no activity occurred with Zn\(^{2+}\). However, the optimum pH for the dCTP deaminase activity was 7.0, which differs from the pH optimum of 9.5 for dCMP deaminase activity. The dCTP deaminase activity was also feedback inhibited by dTTP, decreasing about 10-fold in the presence of 2 mM dTTP (Fig. 5).

Kinetic analyses of bi-DCD. To characterize the kinetic properties of bi-DCD, initial rates for both dCMP and dCTP deaminase activities were measured at various substrate concentrations. dCMP deaminase activity exhibited a small sigmoidal relationship between the rate of dCMP deamination and the concentration of substrate (dCMP), with a Hill number of 2.5 (Table 1). These results imply that dCTP deamination and the concentration of substrate (dCTP), with a Hill number of 2.5 (Table 1). These results suggest that the same active site is involved in both dCMP and dCTP deaminations.

Expression of the A596R gene in PBCV-1-infected chlorella cells. RNA was extracted from virus-infected cells at various times after infection and hybridized to an A596R gene probe to determine when the gene is transcribed during PBCV-1 replication. The probe bound to an ~1.1-kb RNA transcript extracted from cells beginning at ~45 min p.i. The intensity of the hybridization increased until 90 min p.i., after which it decreased rapidly (Fig. 6A). PBCV-1 DNA synthesis begins 60 to 90 min after virus infection (37), indicating that the bi-DCD gene is expressed as an early-late gene. The 1.1-kb RNA transcript was slightly larger than expected for a PBCV-1-encoded 142-amino-acid protein.

Occurrence of the bi-DCD gene in other chlorella viruses. To determine if the bi-DCD gene is common among the chlorella viruses, genomic DNAs from 47 chlorella viruses isolated from diverse geographical regions, as well as the host chlorella, were hybridized to the PBCV-1 A596R gene probe used in the Northern analyses (Fig. 6B). DNAs from 36 of the viruses that infect Chlorella NC64A (NC64A viruses) hybridized strongly with the probe, while no hybridization was observed with the other six NC64A viruses, NYs-1, IL-5-2s1, MA-1D, NY-2B, NY-2A, and Ar158. Typically, the nucleotide sequences of gene homologs in these six viruses differ sufficiently from that of PBCV-1, such that hybridization signals are barely detectable.

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**TABLE 1. Kinetic constants of bi-DCD for two substrates**

<table>
<thead>
<tr>
<th>Substrate</th>
<th>$K_m$ (mM)</th>
<th>$k_{cat}$ (s(^{-1}))</th>
<th>$V_{max}$ (μmol min(^{-1}) mg(^{-1}))</th>
<th>$k_{cat}/K_m$ (10(^5) M(^{-1}) s(^{-1}))</th>
<th>Hill coefficient</th>
</tr>
</thead>
<tbody>
<tr>
<td>dCMP(^a)</td>
<td>0.76</td>
<td>24.9</td>
<td>87.9</td>
<td>32.8</td>
<td>1.8</td>
</tr>
<tr>
<td>dCTP</td>
<td>0.64</td>
<td>5.7</td>
<td>20.2</td>
<td>8.9</td>
<td>2.5</td>
</tr>
</tbody>
</table>

\(^a\) With 0.1 mM dCTP.

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**TABLE 2. Effects of inhibitors on bi-DCD dCMP and dCTP deaminase activities**

<table>
<thead>
<tr>
<th>Substrate</th>
<th>Amt of inhibitor (μM)</th>
<th>% Inhibition</th>
<th>PDRP</th>
<th>H(_4)dUMP</th>
</tr>
</thead>
<tbody>
<tr>
<td>dCMP(^a)</td>
<td>0</td>
<td>100</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>94</td>
<td>97</td>
<td>77</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>82</td>
<td>85</td>
<td>77</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>77</td>
<td>60</td>
<td>77</td>
</tr>
<tr>
<td>dCMP(^b)</td>
<td>0</td>
<td>100</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>36</td>
<td>7</td>
<td>9</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>64</td>
<td>38</td>
<td>38</td>
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<tr>
<td></td>
<td>10</td>
<td>77</td>
<td>60</td>
<td>60</td>
</tr>
<tr>
<td>dCTP</td>
<td>0</td>
<td>100</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>56</td>
<td>49</td>
<td>49</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>82</td>
<td>85</td>
<td>85</td>
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<tr>
<td></td>
<td>10</td>
<td>94</td>
<td>97</td>
<td>97</td>
</tr>
</tbody>
</table>

\(^a\) With 0.1 mM dCTP.

\(^b\) Without dCTP.
able (e.g., see the RNase III gene [47]). No hybridization was
detected with DNA from the Chlorella NC64A host or DNAs
from five viruses that infect Chlorella Pbi (Pbi viruses). This
lack of hybridization is not surprising since the nucleotide
identity of homologous genes between the NC64A and Pbi
viruses is typically 60 to 65% (37). The genomes of viruses
NY-2A (an NC64A virus) and MT325 (a Pbi virus) have been
sequenced recently (6, 7), and computer analysis indicates that
both viruses encode a dCMP deaminase ORF, with 81% and
63% amino acid identity, respectively, to PBCV-1 bi-DCD. Fur-
thermore, recombinant proteins from the two viruses have both
dCMP and dCTP deaminase activities (results not shown).

Phylogenetic analysis. Maximum-parsimony (heuristic),
neighbor-joining (distance method), and maximum-parsimony
using bootstrap (1,000 replicates) analyses produced nearly
identical tree topologies. The tree shown in Fig. 7 was con-
structed using the neighbor-joining algorithm in PAUP (36).
The values on the branches are the percentages of bootstrap
support (1,000 replicates). In order to root the tree, the se-
quence of a Streptococcus thermophilus late competence pro-
tein required for DNA binding (contains a dCMP deaminase
domain) (NCBI YP_142074) was used as the outgroup in the
phylogenetic analyses. The taxon closest to PBCV-1 was from
another virus, bacteriophage phiJL001, whose host is a marine
proteobacterium that associates with a sponge (19). The next
two most closely related taxa in the same clade are the pro-

The ClustalW amino acid sequence alignment of the 22 taxa
(see the legend for Fig. 7 for a listing of taxa used in the
alignment) showed high divergence among taxa, with only 11
of 142 amino acids being identical. The taxon closest to
PBCV-1 was bacteriophage phiJL001 (NCBI YP_223954),
with 63 identical and 27 similar amino acids out of 142. To-
gether, the four taxa in the PBCV-1 clade differed consider-
ably, with only 33 identical and 24 conserved amino acids.
Except for another phycodnavirus infecting the alga Emili-
ani huxleyi (44), the gene is absent in other viruses infecting eu-
karyotes.

DISCUSSION

In deoxynucleotide metabolism, dUMP is the immediate
precursor in the endogenous de novo synthesis of dTTP (Fig.
1). There are three known pathways to synthesize dUMP: (i)
deamination of dCMP to dUMP by dCMP deaminase, (ii)
deamination of dCTP to dUTP by dCTP deaminase followed
by hydrolysis of dUTP to dUMP by dUTPase, and (iii) reduc-
tion of UDP or UTP to dUDP or dUTP, respectively, by
ribonucleotide reductase followed by hydrolysis of dUTP to
dUMP by dUTPase. The dCMP deaminase route is the most
common pathway for synthesizing dUMP in eukaryotes and
prokaryotes (21, 31). However, dCMP deaminase is not ubiq-
tuous. In the gram-negative bacteria E. coli and Salmonella
enterica serovar Typhimurium, dUMP is synthesized from
dCTP that is produced sequentially by dCTP deaminase and
dUTPase (12, 28, 29).
Rapidly multiplying DNA viruses place a huge demand on their host to supply deoxynucleotides for genome replication; the problem is even more acute in nonproliferating cells. To overcome this problem, many large DNA viruses encode some nucleotide synthetic enzymes, including both subunits of ribonucleotide reductase. Chlorella virus PBCV-1 follows this pattern and encodes at least 13 putative enzymes involved in nucleotide metabolism (46). Recombinant proteins have been made from 5 of these 13 genes (and shown to be functional): aspartate transcarbamylase (18), glutaredoxin (Y. Zhang et al., unpublished results), dUTPase (48), thymidylate synthase X (11), and, as described here, dCMP deaminase. Unexpectedly, however, the PBCV-1 dCMP deaminase is bifunctional and has both dCMP and dCTP deaminase activities. Thus, assuming that the PBCV-1-encoded ribonucleotide reductase is active, the virus encodes enzymes in all three known pathways to...
synthesize dUMP, indicating that dTTP synthesis is well supported during PBCV-1 DNA replication (Fig. 1). Presumably the dTTP pool size in host cells (the chlorella host DNA is ~33% A+T) is not large enough to support rapid growth of PBCV-1, whose DNA is 60% A+T. In addition, the DNA concentration in PBCV-1-infected cells increases at least fourfold by 4 h p.i.; this amount cannot be accounted for simply by recycling deoxynucleotides from degraded host DNA (37). The bi-DCD gene is transcribed shortly after infection (30 min p.i.), and the transcripts are present during the early phase of DNA synthesis (60 to 90 min p.i.) (Fig. 6A). During this time, the virus is preparing a cellular environment for optimal viral DNA synthesis, and as an allosteric regulatory enzyme, bi-DCD probably plays a central role in adjusting the nucleotide pools. The fact that all four PBCV-1-encoded enzymes that contribute to dTTP synthesis are present in five more sequenced chlorella viruses (6–8) supports the importance of dTTP synthesis to chlorella replication.

The discovery that the PBCV-1 dCMP deaminase also has dCTP deaminase activity was a major surprise because, although their substrates are similar, dCMP deaminase and dCTP deaminase belong to separate enzyme classes that have different mechanisms of deamination, i.e., the former requires Zn$^{2+}$ and the latter does not. dCMP deaminase is most closely related to cytidine deaminase, which catalyzes the same reaction but uses unphosphorylated cytidine as a substrate. Both dCMP and cytidine deaminases have similar molecular topologies and have two similar domains; the second domain contains the residues that form the catalytic site, suggesting similarities in the catalytic mechanisms of the enzymes (1, 33). In contrast, dCTP deaminase and dUTPase share conserved motifs, one of which is the uridine-binding site that is also common in pseudouridine syntheses, suggesting that these functionally connected enzymes have evolved from a common ancestor (17).

An amino acid sequence analysis of bi-DCD indicates that it contains motifs characteristic of dCMP deaminases from other organisms (Fig. 2). In contrast, bi-DCD has no amino acid similarity to E. coli dCTP deaminase (15). A comparison of the PBCV-1 enzyme with dCMP deaminases from human and T4 phage, which have been studied extensively and represent two distinct classes of the enzyme, indicates that PBCV-1 bi-DCD has more similarity to human dCMP deaminase than to the T4 phage enzyme. Human dCMP deaminase, which has no dCTP deaminase activity (F. Maley, unpublished results), contains a single zinc ion per monomer that is required for catalytic activity. In contrast, the T4 enzyme contains two zinc ions per monomer and both are required for activity, but only one appears to be involved in the catalytic reaction mechanism (1). PBCV-1 bi-DCD has one putative zinc-binding site that could be formed by amino acid residues Cys95, Cys98, and His67 (Fig. 2); however, the activity of bi-DCD obtained with Zn$^{2+}$ is less than 6% of the activity obtained with Mg$^{2+}$, suggesting the enzyme does not require Zn$^{2+}$ for activity. Furthermore, analysis of bi-DCD for Zn$^{2+}$ indicated that Zn$^{2+}$ is not bound to the protein.

Like activities of dCMP deaminases from other organisms, the bi-DCD dCMP deaminase activity is activated by dCTP and inhibited by dTTP. Thr75 and Arg78 correspond to the dCTP and dTTP binding sites in T4 phage dCMP deaminase (Phe112 and Arg115) (16). PBCV-1 bi-DCD, like human dCMP deaminase, retains some activity in the absence of dCTP, while T4 dCMP deaminase is completely dependent on dCTP for activity. As a bifunctional enzyme, PBCV-1 bi-DCD has similar reaction requirements for the two substrates, except for pH; dCMP deaminase activity is highest at pH 9.5, and dCTP deaminase activity is highest at pH 7.0. Two well-known dCMP deaminase inhibitors, PDRP and H4dUMP, inhibited both bi-DCD activities, suggesting that the same active site is involved in dCMP and dCTP deamination. Taken together, the results suggest that bi-DCD uses the same mechanism to deaminate both dCMP and dCTP. bi-DCD may have a more flexible active site than other dCMP deaminases; this flexibility could allow the site to interact with either dCMP or dCTP. This hypothesis is supported by the finding that bi-DCD also converts dCDP to dUDP (results not shown). The conformation of the enzyme may change slightly at different pHs and allow the active sites to accommodate the three substrates.

As an activator, dCTP increased dCMP binding to bi-DCD and increased dCMP deaminase activity, suggesting that dCTP alters the conformation of the enzyme to make the active site of the enzyme more suitable for dCMP and more efficient for the deamination reaction. This result suggests that activator dCTP may bind to a putative regulatory site that causes a conformation change in the enzyme. However, when dCTP serves as a substrate at high concentrations, dCTP is deaminated at the active site of the enzyme. Therefore, the enzyme presumably has two dCTP binding sites, one involved in activation and the other serving as a substrate in the catalytic site.

Accumulating evidence suggests that the chlorella viruses have a long evolutionary history (4), possibly dating back to the time that prokaryotic and eukaryotic organisms separated, ca. 2.0 to 2.7 billion years ago (3, 5, 9, 13). Phylogenetic analysis of DNA polymerases places the phycodnavirus enzymes near the root of all eukaryotic δ DNA polymerases (42, 43). A couple of observations in the current manuscript are consistent with the proposed ancient history of the chlorella viruses. bi-DCD is the second known bifunctional enzyme encoded by PBCV-1. Previously, it was reported that the PBCV-1-encoded ornithine decarboxylase decarboxylates arginine more efficiently than ornithine (35). Maybe progenitor enzymes, like the two PBCV-1 bifunctional enzymes, are more precocious than the highly evolved enzymes in present-day organisms, where two separate enzymes carry out the function of one PBCV-1 enzyme. (Note: the bifunctional activity does not result from a fusion between two separate catalytic domains.) The versatility of bi-DCD is also reflected in its ability to function in a broad range of environmental conditions, e.g., temperatures (25 to 55°C) and pHs (6 to 11). Finally, the phylogenetic analyses indicate that PBCV-1 bi-DCD is more closely related to a bacteriophage and two proteobacteria than it is to enzymes from eukaryotes (Fig. 7). One thing in common among these four organisms is their aquatic origins. PBCV-1’s host, Chlorella, is common in freshwater, as is Magnetospirillum magnetotacticum. Vibrio parahaemolyticus is found in brackish water, and the host for bacteriophage phiL001 is a marine bacterium.

In summary, (i) chlorella virus PBCV-1 encodes a bifunctional enzyme (bi-DCD) that deaminates dCMP and dCTP to dUMP and dUDP, respectively; (ii) dCTP can serve as either a substrate or an activator of bi-DCD; (iii) dTTP inhibits both
bi-DCD activities; (iv) although the enzyme has a putative 
Zn^{2+} binding site, no detectable Zn^{2+} is associated with the 
protein (the best enzyme activity occurs with Mg^{2+}); (v) the 
native structure of bi-DCD is a hexamer; (vi) transcription of the 
bi-DCD gene begins 45 min after PBCV-1 infection, and the 
mRNAs disappear quickly after 120 min; (vii) the bi- 
DCD gene is widely distributed in the chlorella viruses; and 
(viii) to date, the gene has been found only in viruses infecting 
algae.

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