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Occurrence of Cyanazine Compounds in Groundwater: Degradates More Prevalent Than the Parent Compound

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Introduction

Cyanazine has historically been one of the most heavily used herbicides in the country, with roughly 9 million kg being used annually in the United States (10). Cyanazine use in the United States, however, is scheduled to end in the year 2000 (http://www.epa.gov/pesticides/citizens/triazine.htm). The degradation pathway for cyanazine is shown in Figure 1. The effect of these degradates on human health, however, are generally unknown. Because these cyanazine degradates are still chlorinated (Figure 1), it is anticipated that these degradates could be similar to the chlorinated degradates for atrazine (deethylatrazine and deisopropylatrazine) that have been found to be phytotoxic (11, 12). Indeed, cyanazine amide (CAM) has been shown to retain some of the herbicidal activity of the parent compound (13).

Research has suggested that because of the persistence of the s-triazine ring that degradates of cyanazine will be found in groundwater once they are looked for (14). Indeed, research has documented the presence of CAM and DIA in groundwater (3, 4). Little data have been obtained on the environmental occurrence of other cyanazine degradates due to the inherent difficulty in measuring these compounds (15). Recently, however, a sensitive and reliable method has been developed to measure cyanazine and its major degradates using liquid chromatography/mass spectrometry (LC/MS) (16, 17).

This paper describes the results of a study to determine the occurrence of cyanazine and its degradates (CAC, CAM, deethylcyanazine (DEC), and deethylcyanazine acid (DCAC)), in groundwater. Results from the sampling of 64 municipal wells across Iowa during the summer of 1999 are presented (Figure 2). To our knowledge, this is the first large-scale investigation for CAC, DEC, and DCAC in water resources and is one of the most detailed studies on the occurrence of cyanazine compounds in groundwater to date. This research is an extension of the multi-agency Iowa Ground Water Monitoring (IGWM) Program (18, 19).

Methods

In 1992, a network of 90 sampling sites were randomly selected from an inventory of over 2000 Iowa municipal wells representing all major aquifer types (alluvial, glacial drift,
bedrock/karst region, and bedrock/nonkarst region). The number of selected sampling sites from the major aquifer types was designed to represent the distribution of all municipal wells in Iowa. The annual sampling schedule, however, was such that wells suspected to be minimally impacted by land-applied chemicals were sampled less frequently (every 2–4 yr rather than annually). The water samples collected from the 64 wells (Figure 2) represent the sampling carried out for the IGWM during 1999. Wells thought to be least susceptible to land-applied chemicals were not sampled during this time period.

The sampling protocol for this study has been reported previously (20). All wells were pumped for at least 30 min before dissolved oxygen, pH, specific conductance, and water temperature were measured. Once the values for these parameters stabilized, the water samples were collected.

Water samples were filtered through a 0.7-μm glass fiber filter into 125-mL amber baked-glass bottles, immediately chilled, and sent by overnight air express to the U.S. Geological Survey Organic Research Laboratory in Lawrence, KS. Concentrations of cyanazine and its degradates CAC, CAM, DEC, and DCAC were analyzed from the 125-mL groundwater samples using a newly developed method using LC/MS. Details of this method have been described previously (16, 17). The analytical reporting limit for this method was 0.01 μg/L.

Total cyanazine residue (CYTOT) was determined by summation of cyanazine, CAC, CAM, DEC, and DCAC concentrations. Measured concentrations that were less than the analytical reporting limit for any of the individual concentrations were treated as zero in the calculation of CYTOT. Because deisopropylatrazine can also be derived from the degradation of atrazine (17), it was not included in the calculation of CYTOT.

The Kruskal–Wallis test (21) is a nonparametric statistical method that was used to test for differences among groups of data. Test results were evaluated at the 0.05 level of significance.

**FIGURE 1. Pathways for degradation of cyanazine (17).**

![Pathways for degradation of cyanazine](image)
Results

Similar to previous pesticide investigations (4, 22), cyanazine was infrequently detected in groundwater for this study (Figure 3). Several of the cyanazine degradates, however, were much more frequently detected in groundwater than its parent compound (Figure 3). The more frequent presence of CAC and CAM over cyanazine is in agreement with the reported greater stability of these degradates after transformation from cyanazine (13, 15, 16). The cyanazine compounds with the greatest frequency of detection (CAC and DCAC) are organic acids and as such are negatively charged at the pH of groundwater. Thus, these negatively charged species are soluble and more likely to be transported to groundwater than cyanazine. These results are similar to that for alachlor and metolachlor, which each form an oxanilic acid (17) that is transported to groundwater much more readily than their parent compounds (4). The detection of total cyanazine residue (CYTOT) increased more than 12-fold over that of just the parent compound, with maximum concentrations of CYTOT (3.3 μg/L) 165 times that for cyanazine (0.02 μg/L) (Figure 3).

Of the total measured concentration of cyanazine, only 0.2% was from its parent compound (Figure 4). For comparison, parent compounds also comprised a minority of the total measured concentrations for acetochlor (0%), alachlor (0%), atrazine (40.0%), and metolachlor (6.9%) during this investigation. Two cyanazine degradates (DCAC and CAC) comprised 92.5% of the measured cyanazine concentrations for this study (Figure 4). Although DCAC and CAC have similar frequencies of detection (Figure 3), DCAC was generally present in higher concentrations. Conversely, it appears that cyanazine and DEC are relatively unstable with little transport to groundwater (Figures 3 and 4). As hypothesized previously (15, 23), cyanazine may degrade through various pathways to deisopropylatrazine and deethylatrazine (Figure 1), compounds that are also derived from the degradation of atrazine.

No concentrations of cyanazine compounds for this study exceeded water-quality criteria for the protection of human health. Only cyanazine however has such a criteria established—having an unenforceable health advisory level set at 1.0 μg/L. Making the gross assumption that the chlorinated degradates may have similar toxicity as the parent compound—similar to what has been found with the
The results of this study documented that data on the degradates for cyanazine are critical for understanding its fate and transport in the hydrologic system, similar to what has been found with acetochlor, alachlor, atrazine, and metolachlor (3, 4, 25, 26). Furthermore, the prevalence of the chlorinated degradates of cyanazine found in groundwater suggests that to accurately determine the overall effect of cyanazine on human health and the environment its degradates should also be considered.

Although there is no apparent geographical pattern to the presence of CYTOT in groundwater (Figure 5), the data did vary hydrogeologically. A significant difference ($p = 0.014$, Kruskal–Wallis test) in the occurrence and concentration of CYTOT was determined among the major aquifer types sampled (Figure 6). The occurrence of CYTOT was much more prevalent in alluvial aquifers, being found about 2–5 times more frequently than the other major aquifer types sampled. In contrast, cyanazine occurrence rates did not significantly differ ($p = 0.517$, Kruskal–Wallis test) among the major aquifer types sampled.

The relative infrequency with which CYTOT was detected in the bedrock/karst aquifers (Figure 6) is in sharp contrast to that determined from an earlier study of herbicides and herbicide degradates where 77.4% of the bedrock/karst aquifers were found to have these compounds present (4). Most of these herbicide detections were from select degradates of alachlor and metolachlor (4). The lower frequency of detection found for CYTOT as compared to the earlier study of herbicide degradates in groundwater might be an artifact of the smaller number of bedrock/karst wells sampled in 1999.

Because of the strong interaction between groundwater and surface water within alluvial systems (27), the prevalence of CYTOT in alluvial aquifers has potential long-term implications for the occurrence of CYTOT in streams. Cyanazine compounds can be transported to alluvial aquifers through a variety of processes such as infiltration through the unsaturated zone following application to soils being farmed in the flood plain (28, 29), episodic stream flooding (30), bank storage of streamwater (31), and contaminated streams losing water to the groundwater system (32). Many streams across the Midwest receive significant groundwater contributions to streamflow—particularly during base-flow conditions. Therefore, once an alluvial aquifer becomes contaminated with cyanazine compounds, it can provide a long-term source of CYTOT to the corresponding streams. Thus, even though the use of cyanazine is nearing its end in the United States, the legacy of that use, as reflected in the presence of CYTOT in groundwater, may persist for many years because of the lag-time between changes in chemical use at the land surface and concentrations in groundwater.

Consequently, it is anticipated that low-level concentrations of CYTOT will continue to be found in streams for years after the use of cyanazine has stopped—primarily through its movement from groundwater into streams during base-flow conditions. Groundwater has been previously documented to be a source of herbicide concentrations to streams (33) and has been shown that its contributions to stream contamination of herbicide compounds can be long-term in nature. For example, a degrade of alachlor was one of the most frequently detected herbicide compounds in alluvial aquifers across Iowa (4) even though alachlor had little use documented during the year of sample collection. Alachlor has progressively decreased from being the second most heavily used pesticide in 1990, to the 12th in 1996, to essentially zero use in 1999 (34). Few detections of the parent compound for alachlor were found during this study. In addition, a regional study of 70 streams across the Illinois, Iowa, and Minnesota during base-flow conditions in 1997 found the same alachlor degrade as one of the most frequently detected compounds (35). No detections of the parent compound for alachlor were found during this stream sampling. The evidence of prevalent detections of the same alachlor compounds in both alluvial aquifers and streams during base-flow conditions combined with there being multiple years since the extensive use of alachlor supports the hypothesis that alluvial aquifers can act as a long-term source of herbicide compounds to streams.

Because the wells in this study were derived from a variety of aquifers and hydrogeologic settings across Iowa, groundwater age varies significantly among these wells. Previous research has shown that the frequency of herbicide detection generally increases with decreasing water age (36, 37). Although no direct measures of groundwater age were obtained for this study, two indirect estimates of age were available—dissolved-oxygen concentrations and well depth.
and low-vulnerability classes. Because the lag time between the youngest groundwater than samples from the intermediate- the high vulnerability class (wells estimated to contain the CYTOT was detected about 3
degradates are accounted for (Figure 8). The occurrence of detection for each group. An explanation of a boxplot is provided in Figure 3.

changes in chemical use at the land surface and concentrations in groundwater should become shorter as groundwater age decreases, it is anticipated that CYTOT concentrations will first decrease in the wells within the high vulnerability class following the termination of cyanazine use.

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FIGURE 7. Summation by aquifer type of well depth for the sites sampled and dissolved-oxygen concentration for the corresponding water samples collected (ALLUV = alluvial, GD = glacial drift, BK = bedrock/karst region, and BNK = bedrock/nonkarst region). An explanation of a boxplot is provided in Figure 3.

Acknowledgments
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