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## Conservation Efforts and Natural History of a Prairie Habitat at Jennings Environmental Education Center, Pennsylvania

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#### **Abstract**

The habitat considered in this study is a 19.8-acre (8.0-hectare) prairie relict, which is part of the 296-acre (120-hectare) park (Jennings Environmental Education Center), and it is located in northwest Pennsylvania (Butler County). This prairie remnant is one of the easternmost existing relicts of the once widespread post-glacial prairies. The focus of this paper is to present an overview of the natural history of the Jennings Prairie with emphasis on preservation practices that have occurred since its discovery, in 1905, and management approaches aimed at the conservation of this habitat. The maintenance of its 225 native plant species requires human intervention in order to avoid its succession into a forest ecosystem, which is typical for this bioregion. Pedologic data are presented briefly as a framework used to justify decision-making in common land management practices. The conservation of the Jennings Prairie (the only one remaining of a few prairies dislocated through the landscape of western Pennsylvania) remains a unique habitat unit to demonstrate ecosystem diversity in the region.

**Keywords:** prairie, adaptive management, conservation, Jennings Environmental Education Center, relict prairie, western Pennsylvania

Although deciduous forests cover almost entirely the state of Pennsylvania, prairie habitats have been described in the western part of the state for decades (Aaron 1974a, 1974b). A consistent establishment of early European settlements in the second half of the eighteenth century initiated major physical changes on the landscape. Smith (2001) argued that prairie extirpation from the Midwestern landscape occurred primarily because of agricultural expansion and urban development. A similar fate affected prairie habitats in western Pennsylvania and has been attributed to both deforestation and aggressive mining operations (Taylor and others 2003). In addition to this, the great oil boom of the 1800s led to the establishment and development of towns all over northern Butler County, although no oil was found in the Jennings area (M. Dunn, personal communication 2002). Perhaps, it was to save this rare testimony of its ancient natural history that the Western Pennsylvania Conservancy purchased Jennings Prairie from private landowners in 1951.

Today this relict habitat is part of Jennings Environmental Education Center, which is located in Butler County (northwest Pennsylvania) and comprises 296 acres (120 ha) managed by the Pennsylvania Department of Conservation and Natural Resources (DCNR), Bureau of State Parks. The nature reserve includes a relict prairie of the once Slippery Rock cluster (Aaron 1974a), which is adjacent

to U.S. Route 8, 19.3 km north of Butler, Pennsylvania. Here, in 1905, a 19.8-acre (8-ha) prairie ecosystem was discovered by Dr. Otto Emery Jennings, a botanist with the Pittsburgh Carnegie Museum (DCNR brochure). According to Aaron (1974a, 1974b) prairie habitats evolved in western Pennsylvania at the end of the glacial era (about 14,000 years ago). Eleven of them were discovered in the 1960s in Butler County.

The Jennings Prairie is valued for its dominant growth of forbs, and among these the dense blazingstar (*Liatris spicata*) is particularly appreciated. The purpose of this paper is to describe this rare, wet-mesic prairie habitat in a typically forested region and to present its natural history. Additionally, our paper focuses on discussing conservation strategies and challenges in prairie maintenance by managers and researchers at Jennings Environmental Education Center with the aim of achieving adaptive and sustainable management practices for the long-term prosperity of this, unique prairie habitat.

#### **Physical Features of the Land**

Jennings Environmental Education Center and its relict prairie are part of the Appalachian Plateau Province and stand at altitudes ranging between 1,247 and 1378 ft

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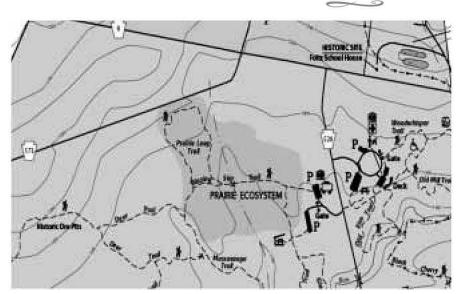


Figure 1. Jennings Prairie map. Adapted from DCNR brochure

(380–420 m) above sea level (State Park Management Manual 1993). The topographic features of this landscape have been shaped through long periods of orogeny, glaciation, and erosion. Geologic data indicate that the region was above sea level during the Mesozoic era and the Tertiary period so that the only continental deposits belong to the Pleistocene ice age and these lie directly on the Pennsylvanian rocks, which are almost horizontally layered through the land profile (Deevey 1949).

Continental glaciers advanced toward the Jennings area three times between 120,000 and 23,000 years ago, and geologic history relevant to present day conditions of Jennings Prairie occurred at similar times, during a concurrent establishment of prairie habitats in southern Ohio (Braun 1928b). At earlier times however (from 360 to 330 million years before present), all of western Pennsylvania was part of a shallow, inland sea, bordered on the west by deep waters, and on the east by a then newly formed ridge which lasted about 30 million years (Transeau 1935, Mehrhoff 1997). Erosion and sedimentation contributed to the establishment of rock strata between the late Paleozoic and early Mesozoic eras and shaped the characteristics of its soils, which derived primarily from sediments of glacial lakes (Transeau 1935, Deevey 1949). The moderate permeability of most soils in western Pennsylvania supports a high, seasonal water table and makes them unfeasible for cashcrop farming, or development. They are mostly gently sloping and acidic (Soil Survey of Butler Co. 1989), favoring an agricultural production of corn, soy bean and fodder crops that are grown in rotation, to suffice the needs of the local dairy industry. Pedogenesis at Jennings goes back to 14,000 years, when the glaciers that until then covered the landscape, began to recede. An additional 7,000 years determined favorable climatic changes, which enhanced prairie establishment, as the environment evolved through a xerothermic phase (Braun 1928a, Braun 1955, Mehrhoff 1997). However, according to Cowles (1928) this time frame, although consistent, was not sufficient to develop prairie soils

in the grasslands of northeastern North America, as evidenced also by the clayey profile with only 1.6–2.4 inches (4–6 cm) of topsoil, which characterizes the substrate of the prairie habitat under study. The climate of the region is classified as a humid-continental type with rain precipitation that is evenly distributed throughout the year. Therefore, although accurate phenology data are not known for this site, climatic conditions for this area favor maximum blooming between June and early September.

## Jennings Prairie and its Flora

The prairie at Jennings represents a unique community of significant botanical interest, which although discovered in the early

1900s began to be formally protected only five decades later (Aaron, 1974a). Comprising approximately the 6% of the park total acreage, the prairie houses a variety of plants and animals (State Park Management Manual, 1993), including endangered species. The area is predominantly covered with forbs that range between 2 and 4 ft (0.6-1.2 m) in height with scattered shingle oaks (Quercus imbricaria) that are adapted extremely well to the environmental conditions of this unique prairie ecosystem. Two hundred and twenty five native plant species (Appendix 1) were identified at this site in 1964 (State Park Management Manual 1993). The presence of certain prairie plant indicators such as whorled rosin weed (Silphium integrifolium), ox-eye sunflower (Heliopsis helianthoides), Culver's root (Veronicastrum virginicum), mad dog scullcap (Scutellaria laterifolia) and dense blazingstar, including a variety of native grass species characterize this habitat as a relict prairie in the region. Forty-four wildflowers have been described belonging to the family Asteraceae, tickseed (Coreopsis tripteris) being the tallest of all. A small, open water stand and associated wetland of 0.1 acre (0.04 ha) is also part of the relict prairie ecosystem. It divides the Jennings Prairie longitudinally in two, almost equal halves (Figure 1) and is located near the center junction of the four primary quadrants comprising this habitat. Here, some arboreal growth encroachment is present (Quercus spp., Prunus spp., Salix spp., Cornus spp.), in addition to more typical wetland vegetation (Typha spp., Juncus spp., Carex spp.). This wetland area serves also as a refugium and hibernaculum for an endangered animal species: the massassauga rattlesnake (Systrurus catenatus catenatus). Particular attention is paid to routine management practices at the Jennings Prairie to preserve the habitat of this indigenous reptile.

#### Conservation Strategies and Approaches at the Jennings Prairie

Three trails are maintained through the prairie (Blazing Star, Massassauga, and Prairie Loop) in order to facilitate access to the site by visitors and also for obvious managerial reasons (Figure 1). The holistic goal of prairie ecosystem management at Jennings focuses on protecting, maintaining and enhancing a unique ecosystem, in an effort of preserving also rare species, in a biologically diverse community (State Park Management Manual 1993). However, a pursuit toward this primary and broad goal entails interaction with a variety of other goals supporting specific areas. Inevitable conflicts may arise occasionally, when certain goals conflict with one another, or detract from the primary goal.

At Jennings Prairie these conflicts are common and they are resolved with an approach that often demands flexibility in routine decision-making, as simplistic solutions applied to complex systems normally fail to fulfill expected outcomes (Pilione 1998). Maintaining a safe and durable system that will not irreversibly damage the prairie through visitors' impact constitutes an extremely important secondary goal for the Jennings Prairie. More goals include the protection of the prairie from alien vegetation, maintain species composition, control soil water saturation and water based hibernaculum area, manage aggressive tree growth invasion. Maintaining current research programs about management prescription effects is another legitimate goal (State Park Management Manual 1993), as often decisions about land management and use are made without considering implications for ecological disturbances and impacts created by a linear modus operandi (Dale and others 2000). We remain convinced that land management unavoidably demands a decision-making solidly grounded in ecological and ethical principles and theory. A viable framework to prepare land managers to operate in a more sustainable manner is presented by the Land Ethic (Leopold 1949), and we advocate an application of its principles in conservation efforts at Jennings and elsewhere.

#### Prescribed Fires as a Management Tool at Jennings Prairie

Controlled burning is an effective technique in prairie restoration and conservation (Pauly 1997), but in order to effectively fulfill the managerial goals of a specific environment, it may be integrated with other methods to maximize its beneficial effects. At Jennings, vegetation control with prescribed fires takes place usually in the spring, between mid–March and early May and these are coordinated with the Bureau of Forestry and County Control. Copeland and colleagues (2002) discussed how prescribed prairie fires during the late growing season enhance species richness of both forbs and warm-season tallgrasses. A spring burn contemplated by the fire schedule in place at Jennings is conceived to generate

minimum disturbance to the massassauga rattlesnake, which is still hibernating at this time of the year. A 10-foot fire line surrounds the prairie on its western side whereas existing trails serve as a fire line for the eastern boundary (Figure 1). The respective fire lines are mowed semi-annually and when soils are deeply frozen, depending upon which quadrants will be burned on a yearly basis. Therefore, these operations can be conducted during the winter months if heavy, mechanical equipment is used.

The prairie has never been burnt all at once in order to avoid excessive disturbance to the hibernating rattlesnake and, more importantly, to minimize the risk for the surrounding forest to catch on fire. Rosburg (2001) pointed out that late spring fires can be used effectively to control non-native species in prairies without negatively affecting forb communities. Winter (end of January at Jennings) is also conducive to accomplishing manual operations on the prairie, aimed primarily at controlling the propagation of tree seedling community (Aspen spp.), which pose great challenges to prairie preservation at Jennings. Manual operations such as this one could be postponed into late winter, especially if geared also toward seed dispersal, as Jackson (1999) demonstrated the beneficial effects of trampling during seeding with prairie perennials to establish permanent pastures in northern Iowa. Immediate and long-term effects of fire on complex, vegetative, assemblages depends on fire severity, extent and timing as well as on the characteristics of the area (Reichman 1987, Vickery and Dunwiddie 1997). Although forb assemblage fires differ from tallgrass prairies and even more from woodland fires, burns constitute standard maintenance procedures to fit eastern grasslands (Winne 1997). However, frequent burning has been proven to reduce species heterogeneity at the advantage of grasses, if mowing, and/or grazing do not complement vegetation management (Collins and others 1998). Finally, Vickery (2002) demonstrated that the effect of prescribed fires enhances seed viability and protection from insect predators on the northern blazingstar (Liatris scariosa var. novae-angliae). Therefore, it is plausible to surmise that besides positively affecting the plant phenology and propagation, prescribed fires may exert a beneficial control of various pathogens and parasites. The challenge remains to verify with more research that a combination of practices (including burning and mowing), properly alternated, can best fulfill the managerial needs of the site.

#### Conclusion

It remains challenging to verify the environmental features of Butler County, Pennsylvania and its relict prairie at Jennings prior to European settlement. Thus, preserving a habitat that accurately reflects those earlier botanical attributes becomes quite an arduous venture. Allison (2002) argued that even surveys of old tallgrass prairies in the Midwest aimed at pursuing the most precise restoration plans are difficult to achieve in full. Inevitable environmental changes, succession and various anthropogenic factors (including interactions among plants and soil communities) are consistent with the

dynamism occurring in natural systems. A reliable body of literature remains available to learn about prairie habitats in various regions of continental United States (Whitford 1958, Reichman 1987, Van Bruggen 1992, Vickery and Dunwiddie 1997) and assess conservation efforts over time (Bever 1994, Vidrine and others 2001). Concurrently, a growing interest by the general public of adopting a prairie habitat as a model to convert lawns into more sustainable landscapes, inhabited primarily by perennial plant species over annual cultivars is becoming more popular and better accepted through educational initiatives as those programmed by Jennings Environmental Education Center. Recent efforts in Louisiana demonstrate the enthusiastic momentum of the restoration movement in the southern United States, where, until a few years ago, such a paradigm was often unconceivable (M.F. Vidrine, public communication 2001). Since 1995, similar efforts have been occurring at the Robert A. Macoskey Center of Slippery Rock University, an institution for higher education that is located a few miles north of the Jennings Prairie (Doherty and others 2001, Chesto and others 2002). This site has provided for several years outstanding educational opportunities for its students in restoration ecology, while strengthening collaboration in education and research efforts with Jennings Environmental Education Center and other conservation parties in western Pennsylvania. Manuals and other materials have become available recently to learn more about

the botanical heritage of our region and thus, enhance

concerns among local citizens for preserving rare and endan-

gered plant species (Haywood and Monk 2001, Rushing

Several implications to guide future research have emerged from this initial study beside a need to continue with education efforts in our area in order to instill among the general public sound principles of ecosystem conservation, as natural habitats become scarcer and more fragmented. An adaptation in the use of fire while attempting to preserve the endangered massassauga rattlesnake constitutes a clear challenge at the Jennings Prairie. Therefore, a better understanding of the biology of this reptile will allow for a more efficient use of fire in the management of the prairie flora. Alternative methods to fire use studies (mechanical, chemical, manual) to control arboreal population encroachment is another open field for future investigations, which requires great attention at Jennings. Finally, soil and hydrological investigations will broaden the knowledge of the ecology of plant communities at Jennings and of its associated entomofauna. Vickery and Dunwiddie (1997) pointed out for example, how limited is the knowledge about invertebrates (particularly insects), and their ecological interactions with prairie plants. Thus, a better understanding about prairie insects and flora could expand and improve opportunities in achieving also sustainability in modern farming systems (Vidrine and Borsari 1998, Vidrine and Borsari 1999) feasible for this part of the country. We refer primarily to pollination services and biological control strategies by beneficial insect species, which studied at the Jennings Prairie could eventually, become employable in local agroecosystems, as

Pennsylvania remains primarily a vast agrarian region. Also, we remain convinced that a better understanding of the evolutionary consequences of management practices about the plant species being managed at the Jennings Prairie is of paramount importance. The achievement of this objective will require more in-depth knowledge about the ecological processes that shaped the Jennings Prairie and its taxa. Therefore, we advocate a need for more quantitative research framed around current management practices that will generate opportunities to learn more about this unique environment and, ultimately, help us to include new research findings into more improved management practices.

#### **Acknowledgments**

We are grateful to Philip Tramdack and his staff at the Bailey Library of Slippery Rock University for their help with the retrieval of most of the references that were consulted to write this manuscript. We also thank anonymous reviewers and attendees of the 19<sup>th</sup> North American Prairie Conference. Dr. Malcolm F. Vidrine was very supportive and encouraged us enthusiastically throughout this work. Giovanni B. Borsari was helpful with the transcription and organization of the plants list.

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# Appendix 1. Herbaceous, angiosperm list of taxa by major plant group, and then by family from Jennings Prairie in western Pennsylvania.

(Adapted from: State Park Management Manual 1993). The symbol (\*) after the plant scientific name stands for: Introduced (non-native).

	Common Name	Scientific Name	
MONOCOTYLEDONS			
ARACEAE	Northern jack-in-the-pulpit	Arisaema stewardsonii	
	Small jack-in-the-pulpit (L.) Schott	Arisaema triphyllum	
DIOSCOREACEAE	Wild yamroot	Dioscorea villosa	
RIDACEAE	Blue-eyed grass	Sisyrinchium angustifolium P. Mill	
LILIACEAE	White clintonia	Clintonia umbellulata Michx.	
EIEII (OE) IE	Day lily Hemerocallis fulva L.		
	Canada lily	Lilium canadense L. ssp. canadense	
	Large-flowered trillium	Trillium grandiflorum (Michx.) Salisb.	
	Wild oats	Uvularia sessilifolia L.	
	Indian poke	Veratrum viride Ait.	
ORCHIDACEAE	Nodding ladies'-tresses	Spiranthes cernua (L.) L. C. Rich.	
POACEAE			
CACEAE	Slender wheat grass Agropyron trachycaulum		
	Hairgrass	Agrostis hiemalis (Walt.) B.S.P.	
	Redtop	Agrostis stlonifera	
	Big bluestem	Andropogon gerardii Vitman	
	Broom sedge	Andropogon virginicus (L.)	
	Sweet vernal grass	Anthoxanthum odoratum	
	Bearded short husk	Brachyelytrum erectum	
	Common hairy chess	Bromus commutatus	
	Smooth bromegrass	Bromus inermis (Leyss.)	
	Wild chess	Bromus kalmii Gray	
	Wood reed grass	Cinna arundinacea	
	Poverty oats	Danthonia spicata	
	Wide-leaved panicum	Dichanthelium laterifolium	
	Crabgrass	Digitaria sanguinalis	
	Canada ryegrass	Elymus canadensis	
	Bottlebrush grass	Elymus hystrix	
	Riverbank ryegrass	Elymus riparius (Wieg.)	
	Virginia wild rye	Elymus virginicum	
	Nodding fescue	Festuca obtusa Biehler	
	Fowl manna grass	Glyceria striata	
	Velvet grass	Holcus lanatus	
	Bottlebrush grass	Hystrix patula Moench	
	Whitegrass	Leersia virginica (Willd.)	
	Tall millet grass	Milium effusum	
	Nimblewill	Muhlenbergia schreberii	
	Witch grass	Panicum capillare	
	Wide low panicum	Panicum latifolium	
	Linear leaved panicum	Panicum linearifolium	
	Switchgrass	Panicum virgatum L.	
	Canada bluegrass	Poa compressa	
	Foul bluegrass	Poa palustris	
	Yellow foxtail	Setaria lutescens	
	Green foxtail	Setaria viridis	
	Little bluestem	Schizachyrium scoparium (Michx.) Nash	
	Indiangrass	Sorghastrum nutans (L.) Nash	
	Prairie wedge grass	Sphenopholis obtusata	
SMILACACEAE	Carrion flower	Smilax herbacea	
DIVITACACEAE	Common greenbrier	Smilax rotundifolia	
TYDLLACEAE			
ГҮРНАСЕАЕ	Narrow-leaved cattail	Typha angustifolia L.	



#### DICOTYLEDONS

ARACEAE

ASTERACEAE

**ASCLEPIADACEAE** 

ANACARDIACEAE Poison ivy Toxicodendron radicans (L.) Kuntze

APIACEAE Water hemlock Cicuta maculata L.

Honewort Cryptotaenia canadensis (L.) DC

Sweet cicely Osmorhiza claytonii
Anise-root Osmorhiza longistylis
Sanicle Sanicula gregaria

APOCYNACEAE Spreading dogbane Apocynum androsaemifolium L.

Indian hemp Apocynum cannabinum L.
Skunk cabbage Symplocarpus foetidus L.
Poke milkweed Asclepias exaltata L.
Swamp milkweed Asclepias incarnata L.
Common milkweed Asclepias syriaca L.

Butterflyweed Asclepias tuberosa L. Yarrow Achillea millefolium L. (\*)

Lesser ragweed Ambrosia artemisiifolia L.
Field pussytoes Antennaria neglecta
Calico aster Aster lateriflorus (L.) Britt.
New England aster Aster novae-angliae L.
Downy aster Aster sagittifolius
Flat-topped white aster Aster umbellatus P. Mill

Ox-eye daisy Chrysanthemum leucanthemum L. (\*)

Tall thistle Cirsium altissimum
Canada thistle Cirsium arvense (L.) Scop.
Swamp thistle Cirsium muticum Michx.
Bull thistle Cirsium vulgare (Savi) Tenore

Tall coreopsis Coreopsis tripteris Daisy fleabane Erigeron annuus (L.) Pers. Philadelphia fleabane Erigeron philadelphicus L. Robin plantain Erigeron pulchellus Boneset Eupitorium perfoliatum L. Hollow Joe-pye-weed Eupatorium fistulosum White snakeroot Eupitorium rugosum Flat-topped goldenrod Euthamia galetorum Thin-leaved sunflower Helianthus decapetalus Giant sunflower Helianthus giganteus

Ox-eye Heliopsis helianthoides (L.) Sweet

Sneezeweed Helenium autumnale L.
Orange hawkweed Hieracium aurantiacum L. (\*)
Yellow hawkweed Hieracium floribundum
Rough hawkweed Hieracium scabrum Michx.
Two-flowered Cynthia Krigia biflora (Walt.) S. F. Blake
Wild lettuce Lactuca canadensis L. var. canadensis

Dense blazingstar Liatris spicata (L.) Willd.
Black-eyed Susan Rudbeckia hirta L. var. hirta
Green-headed coneflower Rudbeckia laciniata L.
Golden ragwort Senecio aureus L.

Whorled rosinweed Silphium trifoliatum L. var. trifoliatum

Blue-stemmed goldenrod Solidago caesia L. var. caesia

Canada goldenrod Solidago canadensis var. canadensis L.

Late goldenrod Solidago gigantea L.
Early goldenrod Solidago juncea Ait.
Spreading goldenrod Solidago patula

Rough-stemmed goldenrod Solidago rugosa Ait. Var. rugosa

Showy goldenrod Solidago speciosa
Common dandelion Taraxacum officinale L.
Coltsfoot Tussilago farfara L. (\*)

New York ironweed Vernonia noveboracensis (L.) Michx.

BALSAMINACEAE Spotted touch-me-not Impatiens capensis Meerb.
BERBERIDACEAE May-apple Podophyllum peltatum L.

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BRASSICACEAE	Winter cress	Barbarea vulgaris R. Br.
	Pennsylvania bitter cress	Cardamine pennsylvanica
	Cow-cress	Lepidium campestris (L.) R. Br.

Field pennycress Thlaspi arvense L. CAMPANULACEAE Indian tobacco Lobelia inflata L. Lobelia siphilitica L. Great lobelia Pale-spike lobelia Lobelia spicata Lam.

CAPRIFOLIACEAE Glaucous honeysuckle Lonicera dioica var. glauscens Triosteum perfoliatum Tinker's weed

CARYOPHYLLACEAE Cerastium fontanum Baump. Mouse-ear chickweed Deptford pink Dianthus armeria L. Grassleaf chickweed Stellaria graminea L.

Common chickweed Stellaria media (L.) Vill. Dwarf St. Johnswort Hypericum muticum Spotted St. Johnswort Hypericum punctatum Hedge bindweed Convolvulus sepium Upright bindweed Convolvulus spithamala

CUSCUTACEAE Dodder Cuscuta gronovii Willd ex. Schultz

**ERICACEAE** Deerberry Vaccinium stamineum L. **EUPHORBIACEAE** Euphorbia corollata Flowering spurge **FABACEAE** Ground nut Apios americana Medic. Medicago lupulina Black medic Low hop clover Trifolium campestre Alsike clover Trifolium hybridum

Red clover Trifolium pratense White clover Trifolium repens L.

Showy tick trefoil Desmodium canadense (L.) DC. Dillen's tick trefoil Desmodium glabellum

Panicled tick trefoil Desmodium paniculatum Gentiana andrewsii Closed gentian American columbo Frasera caroliniensis Walt. Wild geranium Geranium maculatum

Downv wood mint Blephelia ciliata (L.) Benth. Wild basil Clinopodium vulgare L. Bee-balm Monarda didyma Monarda fistulosa Wild bergamot Monarda media Purple bergamot Downy scullcap Scutellaria incana Mad-dog scullcap Scutellaria laterifolia Collinsonia canadensis L. Horse balm

Glechoma hederacea L. (\*) Ground ivv Henbit Lamium amplexicaule L. (\*) Cut-leaved water-horehound Lycopus americanus L. Lycopus virginicus L. Bugle weed Heal-all Prunella vulgaris L. (\*) Mountain mint Pycnanthemum flexuousum Pycnanthemum virginianum Virginia mountain mint Monotropa uniflora L.

Indian pipe Basil Satureja vulgaris (L.) Fritsch. (\*)

Circaea lutetiana L. **ONAGRACEAE** Enchanther's nightshade

> Purple-leaved willow herb Epilobium coloratum Biehler Seed box Ludwigia alternifolia L. Sundrops Oenothera fructicosa L.

Yellow wood-sorrel Oxalis europaea L. **OXALIDACEAE** Yellow wood-sorrel Oxalis stricta L.

Violet wood-sorrel Oxalis violacea L. Common plantain Plantago major Plantain Plantago rugelii Claytonia virginica L. Spring beauty

PORTULACACEAE Blue phlox **POLEMONIACEAE** Phlox divaricata L. Wild sweet william Phlox maculata L. Greek valerian Polemonium reptans L.

CLUSIACEAE

CONVOLVULACEAE

**GENTIANACEAE** 

**GERANIACEAE** 

MONOTROPACEAE

**PLANTAGINACEAE** 

LAMIACEAE

**POLYGALACEAE** Field milkwort Polygala sanguinea Whorled milkwort Polygala verticillata POLYGONACEAE Hairy Solomon's seal Polygonatum pubescens Long bristled smartweed Polygonum caespitosum Mild water pepper Polygonum hydropiperoides Arrow-leaved tearthumb Polygonum sagittatum L. Climbing false buckwheat Polygonum scandensis L. Polygonum virginianum Jump seed Sheep sorrel Rumex acetosella L. Bitter dock Rumex obtusifolius Fringed loosestrife Lysimachia ciliata L. **PRIMULACEAE** Whorled loosestrife Lysimachia quadrifolia L. RANUNCULACEAE Wood anemone Anemone quinquefolia Thimble anemone Anemone virginiana Marsh marigold Caltha palustris Black cohosh Cimicifuga racemosa Clematis virginiana L. Virgin's bower Kidney leaf buttercup Ranunculus abortivus L. var. abortivus Yellow water crowfoot Ranunculus fascicularis Hairy crowfoot Ranunculus recurvatus Creeping buttercup Ranunculus repens Swamp buttercup Ranunculus septentrionalis Tall meadow-rue Thalictrum pubescens Pursh. Thalictrum thalictroides L. Rue anemone ROSACEAE Agrimonia gryposebala Agrimony Agrimonia parviflora Ait. Swamp agrimony Common strawberry Fragaria virginiana P. Mill ssp. virginiana Geum canadense White avens Potentilla canadensis Dwarf cinquefoil Potentilla simplex Michx. Common cinquefoil Pasture rose Rosa carolina L. RUBIACEAE Cleavers Galium aparine Rough bedstraw Galium asprellum Shining bedstraw Galium concinuum Wild licorice Galium kamtschaticum Stiff bedstraw Galium obtusum Sweet-scented bedstraw Galium triflorum Houstonia caerulea Bluets Partridgeberry Mitchella repens SAXIFRAGACEAE Heuchera americana L. Alumroot Chelone glabra L. SCROPHULARIACEAE Turtle head Square-stemmed monkey-flower Mimulus ringens L. Foxglove beardtongue Penstemon digitalis Nutt ex. Sims Veronica officinalis L. Common speedwell Thyme-leaved speedwell Veronica serpyllifolia L. Culver's root Veronicastrum virginicum (L.) Farw. **UMBELLIFERAE** Queen Anne's lace Daucus carota L. URTICACEAE Stinging nettle Urtica dioica L. VERBENACEAE Blue vervain Verbena hastata L. White vervain Verbena urticifolia L. VIOLACEAE American dog violet Viola conspersa Reichnb. Wooly blue violet Viola hirsutula Brainerd Northern white violet Viola macloskeyi spp. Pallens Common blue violet Viola papilionacea Smooth yellow violet Viola pennsylvanica Michx. Downy yellow violet Viola pubescens Ait. Arrow-leaved violet Viola sagittata Ait.

Wooly blue violet Viola sororia Willd.

VITACEAE Virginia creeper Parthenocissus quinquefolia L.

Fox grape Vitis labrusca New England grape Vitis novae-angliae Frost grape Vitis riparia