

1-1-2013

Polyproline tetramer organizing peptides in fetal bovine serum acetylcholinesterase

Kevser Biberoglu

Hacettepe University, Kevser.biberoglu@gmail.com

Lawrence M. Schopfer

University of Nebraska Medical Center, lmschopf@unmc.edu

Ashima Saxena

Walter Reed Army Institute of Research, ashima.saxena@us.army.mil

Ozden Tacal

Hacettepe University, otacal@yahoo.com

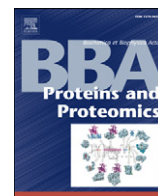
Oksana Lockridge

University of Nebraska Medical Center, olockrid@unmc.edu

Follow this and additional works at: <http://digitalcommons.unl.edu/usarmyresearch>

Biberoglu, Kevser; Schopfer, Lawrence M.; Saxena, Ashima; Tacal, Ozden; and Lockridge, Oksana, "Polyproline tetramer organizing peptides in fetal bovine serum acetylcholinesterase" (2013). *US Army Research*. Paper 183.
<http://digitalcommons.unl.edu/usarmyresearch/183>

This Article is brought to you for free and open access by the US Department of Defense at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in US Army Research by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.



Polyproline tetramer organizing peptides in fetal bovine serum acetylcholinesterase[☆]

Kevser Biberoglu^a, Lawrence M. Schopfer^{b,*}, Ashima Saxena^c, Ozden Tacal^a, Oksana Lockridge^b

^a Department of Biochemistry, School of Pharmacy, Hacettepe University, 06100 Ankara, Turkey

^b University of Nebraska Medical Center, Eppley Institute, Omaha, NE 68198-5950, USA

^c Walter Reed Army Institute of Research, Silver Spring, MD 20910, USA

ARTICLE INFO

Article history:

Received 20 November 2012

Received in revised form 26 December 2012

Accepted 14 January 2013

Available online 23 January 2013

Keywords:

Fetal bovine serum acetylcholinesterase

Polyproline peptide

Mass spectrometry

PAGE gel electrophoresis

Tetramer organization

ABSTRACT

Acetylcholinesterase (AChE) in the serum of fetal cow is a tetramer. The related enzyme, butyrylcholinesterase (BChE), in the sera of humans and horse requires polyproline peptides for assembly into tetramers. Our goal was to determine whether soluble tetrameric AChE includes tetramer organizing peptides in its structure. Fetal bovine serum AChE was denatured by boiling to release non-covalently bound peptides. Bulk protein was separated from peptides by filtration and by high performance liquid chromatography. Peptide mass and amino acid sequence of the released peptides were determined by MALDI-TOF and LTQ-Orbitrap mass spectrometry. Twenty polyproline peptides, divided into 5 families, were identified. The longest peptide contained 25 consecutive prolines and no other amino acid. Other polyproline peptides included one non-proline amino acid, for example serine at the C-terminus of 20 prolines. A search of the mammalian proteome database suggested that this assortment of polyproline peptides originated from at least 5 different precursor proteins, none of which were the ColQ or PRiMA of membrane-anchored AChE. To date, AChE and BChE are the only proteins known that include polyproline tetramer organizing peptides in their tetrameric structure.

© 2013 Elsevier B.V. All rights reserved.

1. Introduction

Vertebrate acetylcholinesterase (AChE; EC 3.1.1.7) belongs to the serine-esterase protein family [1]. AChE terminates the action of acetylcholine at the post-synaptic membrane in the neuromuscular junction. AChE is also found in a variety of other tissues where its function is unclear. For example, AChE is abundant in fetal bovine serum, but not in the serum of the adult cow where its concentration is only 10% of that in fetal serum [2]. In the neuromuscular junction, tetrameric forms of AChE are anchored to the basal lamina via a collagenous subunit referred to as ColQ [3]. In the brain, tetrameric forms of AChE are bound to membranes via a proline-rich membrane anchor (PRiMA) [4]. Both

ColQ and PRiMA contain a proline-rich domain (PRAD) that is credited with organizing AChE monomers into tetramers. The crystal structure of the 40-residue AChE tetramerization domain in complex with the 15-residue PRAD peptide shows four highly conserved tryptophan amphiphilic α -helices from the C-terminus of AChE wrapped around an antiparallel left-handed PRAD helix resembling polyproline II [5]. The essential feature of PRAD is the presence of long stretches of consecutive proline residues. Even synthetic polyproline can function to organize AChE monomers into tetramers [6].

AChE can also be found in soluble form. Co-expression of AChE with the proline-rich fragment from ColQ promotes secretion of stable AChE tetramers in cell culture. In the absence of the proline-rich fragment, only monomers and dimers are secreted, indicating that the proline-rich fragment organizes assembly of the soluble tetramer [7]. It was found that soluble, tetrameric butyrylcholinesterase in human serum is stabilized by a proline-rich peptides derived from the lamellipodin protein [8] in addition to several other proteins. This strongly suggests that the tetramer-organizing peptides used for soluble cholinesterases are different from those employed by membrane-bound forms, but also suggests that they share a common polyproline motif. However, the identity of the proline-rich peptides present in naturally-occurring, soluble AChE has not been determined.

Fetal bovine serum (FBS) is a particularly rich source of soluble AChE. FBS AChE is a symmetric tetrameric globule with a molecular mass of 340,000 Da. The four subunits are associated in a dimer-of-dimers motif. That is, two pairs of disulfide crosslinked monomers

Abbreviations: AChE, acetylcholinesterase; BChE, butyrylcholinesterase; MALDI-TOF-TOF, matrix assisted laser desorption/ionization time-of-flight tandem mass spectrometry; ColQ, a collagen like protein; PRiMA, proline rich membrane; PRAD, proline-rich attachment domain; FBS, fetal bovine serum; MS, mass spectrometry; MSMS, tandem mass spectrometry; BLAST, basic logical alignment search tool; LC/MSMS, liquid chromatography/mass spectrometry including a peptide fragmentation stage; LTQ-Orbitrap, linear ion-trap quadrupole mass spectrometer; NCBI, National Center for Biotechnology Information non-redundant database

[☆] None of the authors have any conflict of interest or potential conflict of interest regarding this paper.

* Corresponding author at: University of Nebraska Medical Center, Eppley Institute, Omaha, NE 68198-5950, USA. Tel.: +1 402 559 6305; fax: +1 402 559 4651.

E-mail addresses: Kevser.biberoglu@gmail.com (K. Biberoglu), lmschopf@unmc.edu (L.M. Schopfer), ashima.saxena@us.army.mil (A. Saxena), otacal@yahoo.com (O. Tacal), lockrid@unmc.edu (O. Lockridge).

are held together by hydrophobic interactions [9]. Based on the foregoing discussion it is to be expected that the tetrameric assembly of FBS AChE is organized around a proline-rich peptide.

In the present study, we extracted and characterized the polyproline peptides of FBS AChE. The protein was denatured by boiling to release the peptides. Released peptides, separated from AChE subunits by HPLC or by filtration, were analyzed by mass spectrometry. We identified polyproline-rich peptides none of which matched the PRAD sequence from either ColQ or PRiMA.

2. Methods and materials

2.1. Materials

AChE from fetal bovine serum was purified from serum pooled from multiple animals as described [10] and stored at -80°C in 50% glycerol until use. The FBS AChE was 99% pure as determined by Coomassie blue staining of an SDS gel and confirmed by measurement of specific activity. α -Cyano-4-hydroxy cinnamic acid (α -CHCA, catalog number 70990) from Fluka (a member of the Sigma-Aldrich group, St. Louis, MO) was prepared as a saturated solution (10 mg/ml) in 50% acetonitrile/water plus 0.3% trifluoroacetic acid. Acetonitrile was of DNA sequencing grade from Fisher (Pittsburgh, PA, catalog number BP-1170), and trifluoroacetic acid was of sequencing grade (>99.9%) from Beckman (Brea, CA, catalog number 290204). 5,5'-dithiobis-(2-nitrobenzoic acid) was from Sigma-Aldrich (catalog number D8130). Formic acid (puriss p.a. for mass spectrometry, catalog number 94318) was from Sigma-Aldrich. All other chemicals were of biochemical grade.

2.2. AChE activity

Activity was assayed with 1 mM acetylthiocholine in 0.1 M potassium phosphate buffer (pH 7.0) at 25°C , in the presence of 0.5 mM 5,5'-dithiobis-(2-nitrobenzoic acid) [11]. One unit of activity is defined as 1 μmol of acetylthiocholine hydrolyzed per minute. The specific activity of 99% pure FBS AChE at pH 7.0, 25°C is 2160 units/mg where protein concentration is calculated from absorbance at 280 nm. The same preparation has a specific activity of 3000–3300 units/mg when activity is assayed at pH 8.0 [9].

2.3. HPLC purification of tetramer organizing peptides

In preparing a sample for HPLC purification of the organizing peptides, the buffer for 14.1 ml of FBS AChE (350 U/ml) was changed from 50 mM sodium phosphate pH 8.0 containing 50% glycerol to 10 mM ammonium bicarbonate pH 8.1 plus 0.01% sodium azide by concentrating the sample to 1 ml in a 10-ml, Amicon stirred-cell (Millipore, Billerica, MA, catalog # 5121) using a YM 30 membrane (Millipore, catalog #13612) and then diluting it to 9 ml. This was repeated 3-times. Finally, the sample was concentrated to 1.4 ml ($A_{280}=2.4$ with a measured activity of 2800 U/ml). Assuming 100% purity and using an A_{280} value of 1.1 for 1 mg/ml FBS AChE, a protein concentration of 2.2 mg/ml could be calculated for this preparation.

The polyproline peptides were released from the FBS AChE preparation (1.4 ml at 1.3 mg AChE/ml in 10 mM ammonium bicarbonate pH 8.1 plus 0.01% azide) by heating the AChE in a boiling water bath for 5 min. The boiled AChE was acidified to pH 2 with trifluoroacetic acid, filtered through a 0.2 μm syringe filter, and injected onto a Zorbax 300 SB C-18 reverse-phase column (Agilent Technologies, Santa Clara, CA, U.S.A.) attached to a Waters 625 HPLC system (Milford, MA, U.S.A.). The HPLC was operated at room temperature (22°C) at a flow rate of 0.5 ml/min. Buffer A was 0.1% trifluoroacetic acid in water; buffer B was 0.09% trifluoroacetic acid in acetonitrile. Peptides were eluted with a gradient of 0–60% buffer B in 60 min. The absorbance was monitored at 210 nm. The HPLC eluent was collected into 1 min fractions (0.5 ml each). Each fraction was concentrated to approximately 50 μl

in a vacuum centrifuge (Savant SpeedVac), and saved at 4°C for mass spectrometry analysis.

2.4. Filtration to separate tetramer organizing peptides from FBS AChE

In preparing a sample for direct application of the organizing peptides onto the MALDI target plate or for Orbitrap analysis, the buffer for 1.5 ml of FBS AChE (2070 U/ml) was replaced with water using an Amicon stirred cell as described above (process was repeated 4-times). The sample volume was reduced to 0.4 ml. The sample was boiled for 5 min to release the polyproline peptides, and then filtered through a Microcon ultracel YM 10 spin filter (MW cutoff 10,000 Da) to separate the released peptides from residual protein. The volume of the filtrate was concentrated to 0.1 ml making the final concentration of peptides equivalent to that expected from a 25 mg/ml sample of AChE.

2.5. MALDI-TOF-TOF mass spectrometry

All MALDI-TOF-TOF experiments were performed on an Applied Biosystems MALDI-TOF-TOF 4800 mass spectrometer equipped with a 355 nm Nd-YAG laser (Applied Biosystems, Framingham, MA). The samples (0.5 μl) were spotted on a MALDI target plate (Opti-TOF 384 well Insert from Applied Biosystems), air-dried and overlaid with 0.5 μl of α -cyano-4-hydroxy cinnamic acid matrix. Mass spectra were acquired in positive ion reflector mode, under delayed extraction conditions (500 ns), using an acceleration voltage of 20 kV, with laser intensity of 4000–6000 V, a mass range of 1000–4000 Da, a detector voltage multiplier of 0.75, with low mass gate on and low mass gate offset equal to zero. Mass spectra shown are the average of 500 laser shots collected from randomly selected locations on the target spot (50 pulses per location). Selected ions were fragmented by collision-induced dissociation, using a 1 kV method, with air as the collision gas at 2×10^{-6} Torr, a precursor mass window of ± 1 Da, metastable suppression on, a detector voltage multiplier of 0.95, using factory calculated delayed extraction values (DE1 = 370 ns, DE2 = 37,271 ns), and with timed ion selector activated. The identity of the fragments in the MSMS spectra were assigned manually using the Data Explorer software (version 4.9 from Applied Biosystems) with the aid of the Proteomics Toolkit (<http://db.systemsbio.net>) and the MS Product algorithm in Protein Prospector v 5.9.4 from the University of California, San Francisco mass spectrometry facility (prospector.ucsf.edu/prospector/mshome.htm).

2.6. LC/MSMS Orbitrap mass spectrometry

The sample was prepared for the Orbitrap by diluting a 3.6 μl aliquot of the Microcon YM 10 filtrate from boiled FBS AChE (made as described above: 25 mg/ml in water) with 6.4 μl of 0.1% formic acid to make a 116 μM solution of peptides.

LC/MSMS analysis was performed on an LTQ Orbitrap quadrupole mass spectrometer (Thermo Scientific a part of Thermo Fisher Scientific, Rockland, IL) using electrospray ionization. Five microliters of the sample were loaded onto a C18 reverse phase trap column (CapTrap Peptide from Michrom BioResources, Auburn, CA, catalog # TRI/25109/32) and washed with 2% acetonitrile/water plus 0.1% formic acid, on an Eksigent nanoLC 2D HPLC system. Then, the peptides were transferred to a C18 reverse phase separation column (Pico frit BioBasics C-18 from New Objective, Woburn, MA, catalog # PF 7515-100H052) and eluted with a 45 min linear gradient starting with 98% solvent A (2% acetonitrile/water plus 0.1% formic acid) and 2% solvent B (98% acetonitrile/water plus 0.1% formic acid) and ending with 35% solvent A and 65% solvent B. The flow rate was 250 nl/min and the effluent was electrosprayed directly into the mass spectrometer. Data were collected in a data dependent manner with each cycle consisting of one high-resolution mass spectrum (over a 300 to 2000 Da mass range) taken with the Orbitrap and five MSMS fragmentation spectra taken with the LTQ ion trap. Collision-induced dissociation was used for fragmentation with helium

as the collision gas (at 1×10^{-3} Torr) and a normalized collision energy of 35 (or about 35% of the maximum collision energy of 5 V). The activation time was 30 ms, and the activation Q was 0.25. Parent ions of any charge state with a minimum signal intensity of 50,000 cps were accepted for fragmentation. After two data collections for a given mass were completed, that mass was excluded from further analysis for 60 s. Fragmentation spectra were analyzed manually using the Qual Browser feature of Xcalibur software (v 4.9 from Thermo Scientific).

3. Results

3.1. MS analysis

The peptides released from heat-denatured FBS AChE are proline-rich. We found that freezing in the absence of glycerol also caused release of proline-rich peptides from FBS AChE. Fig. 1 is a MALDI TOF mass spectrum showing the masses of the released peptides. They range in mass from 1037 to 2764 Da. The three most prominent peaks (1949.9, 1971.9 and 1993.9 Da) represent the same peptide in 1) the protonated form, 2) the single-sodium adduct form and 3) the double-sodium adduct form, respectively. All major peptides from this sample were subjected to collision-induced fragmentation in the MALDI mass spectrometer. Qualitatively, the same information was obtained from the samples before and after HPLC purification, however better quality fragmentation spectra were obtained from the HPLC purified peptides.

For the peptides with the more intense signals, collision-induced dissociation in either the MALDI or Orbitrap mass spectrometers provided enough fragmentation information to determine their amino acid sequences. These peptides are listed in Table 1. Fragmentation series of the y-, b- and a-ion types were observed. In most cases, the peptides observed with the MALDI mass spectrometer were in complex with a single sodium atom (sodium replacing the added proton which makes the peptide positively charged) while those in the Orbitrap were sodium free.

Five different families of peptides are shown in Table 1, all of which contain extended stretches of proline residues. Family 1 has nine peptides composed solely of proline (from 13 to 25 residues long). Families 2, 3, and 4 contain a non-proline residue at the N-terminus (alanine, leucine/ isoleucine/hydroxyproline, or glutamine/lysine), while Family 5 has serine at the C-terminus. The parent ions in families 1 and 2 include peptides with different numbers of prolines. Different length peptides could arise either naturally via exopeptidase action on the tetramerization peptide while it was attached to AChE in the circulation or via in-source fragmentation of the liberated peptide during

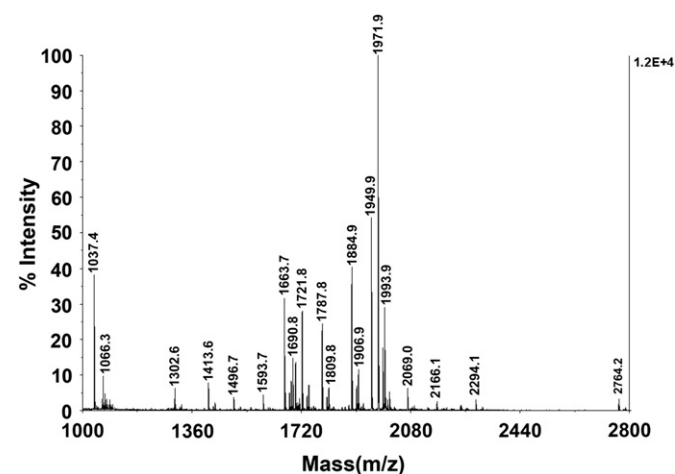


Fig. 1. MALDI mass spectrum of the peptides released from FBS AChE by boiling. Peptides were separated from bulk protein by filtration through a YM10 membrane (10,000 MW cutoff). The right-hand y-axis indicates the signal intensity in counts-per-second.

Table 1

Summary of the peptides released from FBS AChE. Peptides were extracted from serum acetylcholinesterase by simple denaturation and identified by mass spectrometry. Sequences were assigned by manual evaluation of the mass spectral fragmentation patterns.

Sequence ^a	# of residues	Series type ^b	Instrument	Mass, Da
Family 1				
(PPP)PP P(PP)	13	y-, b-ion	Orbitrap	1280.7
(PPPP)P P(PP)P	14	b-ion	Orbitrap	1377.7
(PPPP)P P(PP)P	15	y-ion	Orbitrap	1474.8
(PPPP)P P(PP)P P(PP)P	16	y-, b-, a-ion	Orbitrap	1571.0
(PPPP)P P(PP)P P(PP)P	17	b-, a-ion	Orbitrap	1668.9
PPPP P(PP)P P(PP)P Na ⁺	18	y-, b-, a-ion	MALDI	1788.3
PPPP P(PP)P P(PP)P Na ⁺	19	y-, b-, a-ion	MALDI	1885.0
(PP)PPP P(PP)P P(PP)P P(PP)P	24	y-ion	MALDI	2348.3
(PP)PPP P(PP)P P(PP)P P(PP)P P(PP)P	25	y-ion	MALDI	2445.2
Family 2				
(APP)P P(PP)P P(PP)P	14	y-, b-ion	Orbitrap	1351.7
(APPP)P P(PP)P P(PP)P	16	y-ion	Orbitrap	1545.8
Family 3				
(L/I/Hyp) P(PP)P P(PP)P P(PP)P	16	y-ion	Orbitrap	1587.9
Family 4				
PPPQ/KP P(PP)P P(PP)P P(PP)P Na ⁺	17	y-, b-ion	MALDI Orbitrap	1721.9
Family 5				
PPPP P(PP)P P(PP)P P(PP)P S Na ⁺	21	y-, a-ion	MALDI	2069.1

^a Peptide sequences are written with the N-terminus on the left and the C-terminus on the right. Residues enclosed in brackets indicate that separate signals for the masses of each residue were not resolved in the mass spectrum. However, masses of the bracketed fragments exactly fit the indicated sequence. Leucine, isoleucine, and hydroxyproline (L/I/Hyp) cannot be distinguished because they have the same mass. Glutamine and lysine (Q/K) cannot be distinguished because they have the same mass.

^b The "Series type" column lists the major fragment series observed in the MSMS spectrum of each peptide. Peptide sequences to the left of each series type indicate the specific sequence associated with each type.

introduction into the mass spectrometer. We have seen in-source fragmentation of model polyprolines, in the MALDI mass spectrometer, at laser energies of 4000–6000 V, such as were used in the current experiments (LMS unpublished observations). However, the electrospray ionization energies employed to introduce samples into the Orbitrap are substantially lower making it less likely that in-source fragmentation is responsible for the variation in parent ion length seen in the Orbitrap experiments. Thus, it is likely that both possibilities for shortening the peptide are operative.

Examples of MSMS spectra for these peptides are given in Figs. 2, 3 and 4. The principal ion series detected were a-, b- and y-ion series. The a-series were seen exclusively from peptides in complex with sodium. The b-ion series were found exclusively in the Orbitrap data. There were also major peaks at high mass due to sodium-promoted C-terminal fragmentation. There were stretches of minor peaks corresponding to series of prolines that were not associated with a-, b- or y-ion series. Finally, it was common to find a series of fragments associated with internal fragmentation at proline. Internal fragmentation at the relatively labile proline linkage is a well established phenomenon in mass spectrometry and would be expected under the current circumstances where many prolines are present in the peptides. Most of the unannotated peaks in the MSMS spectra could be accounted for by one-or-another of these types of fragmentation.

3.2. MSMS fragmentation analysis, the first example

Fig. 2 shows MSMS data for the 1885.0 Da peptide, taken with the MALDI mass spectrometer. The parent ion mass is consistent with the presence of 19 prolines plus a sodium atom. Presence of the sodium

the adjacent amino acid and subsequent loss of the residue mass of the C-terminal amino acid [97 Da for proline] leading ultimately to the production of a new alkali cationized peptide lacking the original C-terminal residue" [17]. Additional fragmentation can occur for the newly formed alkali cationized peptide resulting in ions which are two and even three amino acids shorter than the original peptide. The signal intensity of the ion from the second fragmentation can be comparable to that from the first, but the signal intensity for the ion from third fragmentation is generally much smaller [18].

In Fig. 2, the first cycle of sodium-promoted fragmentation is consistent with formation of the intense, 1788.1 Da fragment. A second cycle of sodium-promoted fragmentation would be consistent with the intense signal at 1691.0 Da. However, assigning subsequent fragments to sodium-promoted fragmentation is complicated by overlapping fragments from the weaker, y-ion series. The complication arises from two facts. First, the delta mass for loss of the N-terminal residue from a y-ion series (the high mass end of the spectrum) is equal to the dehydro-mass of the residue. The delta mass for sodium-promoted fragmentation of the C-terminal residue is also equal to the dehydro-mass. Second, the peptides in this study have proline residues at both the C-terminus and the N-terminus (refer to Table 1). Consequently, the delta mass for release of the N-terminal residue for a y-ion series is the same as the delta mass for release of the C-terminal residue by sodium-promoted fragmentation. Discrimination between the two pathways relies on the relative intensities of the fragments. The principal contribution to high intensity signals can be ascribed to sodium-promoted fragmentation, with minor contribution coming from y-ion fragments when the masses from the two pathways overlap.

Additional sodium-promoted fragmentation can result in a prominent loss of 44 Da from the parent ion (probably due to loss of CO₂) [15]. A prominent mass at 1840.9 Da in Fig. 2 corresponds to this fragmentation. Another loss of 44 Da is associated with the 1788.1 Da fragment. Finally, for polyproline peptides, a novel mass, 113 Da smaller than the parent ion (equal to 97 Da plus 16) is typically seen in sodium-promoted fragmentation spectra (L.M. Schopfer, unpublished observations). The 1772.1 Da fragment in Fig. 2 corresponds to this ion.

3.4. MSMS fragmentation, additional examples

Fig. 3 shows MSMS data for the 1721.9 Da peptide, taken with the MALDI mass spectrometer. The parent ion mass is consistent with the presence of 16 prolines plus a glutamine (or lysine), and a sodium atom. Presence of the sodium atom is confirmed by the mass at 23 Da. Fragmentation is again dominated by sodium ion promoted processes. Masses at 1677.7, 1624.9, 1608.9, 1581.0 and 1527.9 Da are consistent with 1) loss of CO₂ from the parent ion, 2) loss of a proline from the C-terminus, 3) loss of 16 Da from the 1624.9 Da ion, 4) loss of a CO₂ from the 1624.9 Da mass, and 5) loss of a proline from the 1624.9 Da mass, respectively. An extensive y-ion series (that includes 13 consecutive prolines) can be extracted from the minor signals. Once again, the y-ion series overlaps the cation promoted fragments. A sequence interval of 128 Da for the difference between 1302.8 and 1430.8 Da corresponds to either a lysine or a glutamine residue. Y-ion and b-ion sequences from two analogous peptides (obtained with the Orbitrap mass spectrometer, see Table 1) indicated the presence of a glutamine/lysine residue on the N-terminus of a long string of prolines confirming the presence of this non-proline residue in the sequence (data not shown). We have assigned the residue to glutamine on the basis of BLAST search results (see the section entitled "BLAST search").

The high intensity of the 1302.8 Da peak is also consistent with the presence of a non-proline residue at this position. The 1302.8 Da mass arises from cleavage on the N-terminal side of proline. The well known "Proline Effect" [19] is renowned for enhancing

the intensity of fragments on the N-terminal side of proline. Finally, a distinct peak for the proline immonium ion was observed at 70 Da [12].

Fig. 4 shows MSMS data for a doubly-charged peptide ($[M+2H]^{+2}=640.85$ Da) taken with an LTQ Orbitrap mass spectrometer. The parent ion mass is consistent with the presence of 13 prolines. Singly-charged peaks for both y-ion and b-ion series are annotated. In addition, there are prominent peaks for doubly-charged b-ions and a y-ion. All of these ions are consistent with a peptide containing only prolines. It is noteworthy that there are strong signals for both a singly-charged y-ion series and a singly-charged b-ion series, and that there is no evidence for fragmentation favoring the loss of two prolines at a time, for either series. This can be compared to studies by Unnithan et al. on Pro_n-NH₂ peptides (n=4–10) who observed that "even numbers of Pro residues were lost from the N-terminal end of the peptide" to form a singly-charged y-ion series. B-ions were not detected [20]. Paired loss of prolines from proline-rich regions was supported by Wang et al. in studies of the proline-rich, 79-residue polypeptide, prophenin 1 [21]. A possible explanation for the difference in fragmentation patterns is that a much higher gas pressure was used in the fragmentation chamber of the Orbitrap (helium at 1×10^{-3} Torr) compared to that used by Unnithan et al. in the Bruker Esquire 3000 plus quadrupole ion trap (helium at 3×10^{-5} Torr).

3.5. BLAST search on the proline-rich peptides

A total of fourteen proline-rich peptides were identified from FBS AChE (Table 1). All of these are suitable candidates for tetramer-organizing peptides. These peptides could be divided into five families. The origin of these peptides is unclear. A logical assumption would be that they begin as part of another protein. In order to test this assumption, the longest representative from each family was submitted for a BLAST (Basic Logical Alignment Search Tool) search of the NCBI database (November 2011) using the mammalian taxonomy (40674). Matches were found for each family. We had anticipated that the necessity for 15 or more prolines in a row would have imposed a severe limitation on the number of matching candidates. However, this was not the case. For example, we found 25 separate proteins that matched the APPPP P P P P P P sequence from family 2. Attempting to sort through this abundance of candidates for those that interact specifically with AChE is beyond the scope of the current study. Therefore, we have selected representative examples for purposes of discussion, which are shown in Table 2. When possible, a protein from the *Bos taurus* taxonomy was chosen for Table 2. When no candidates appeared in the *B. taurus* taxonomy, a candidate from the *Homo sapiens* taxonomy was used. The proteins listed in Table 2 should not be assumed to be the actual source of the corresponding peptide found in AChE tetramers. This issue is addressed in greater detail in the Discussion, Section 4.

Nine of the peptides from Table 1 consisted of only proline residues (family 1). A Blast search using the 25-residue proline peptide found seven different proteins that contained a continuous stretch of 25 prolines. There was no match from the bovine proteome so "UDP-N-acetyl glucosamine transferase subunit homology domain-containing protein 1" from the *H. sapiens* taxonomy was selected for use in Table 2.

The second family was represented by APPPP P P P P P P. There was some ambiguity in the position of the alanine in the N-terminus (see Table 1). Consequently, a BLAST search was made with the alanine at positions 1, 2, 3 and 4 from the N-terminus. All four searches yielded matches to multiple proteins. With the alanine in the N-terminal position (as shown in Table 2) 25 proteins matched the sequence. Five of these proteins were found in the *B. taurus* taxonomy (Predicted R3H domain-containing protein 1; Leiomodin-2; Predicted formin-like 1; Large proline-rich protein BAG6; and Zinc finger homeobox 4).

derive from the membrane-bound forms. For human and horse plasma BChE the source of the polyproline peptide appears to be a number of proteins including lamellipodin [8,22]. In our first report, we identified lamellipodin as the source of the tetramer organizing polyproline peptide [8] however subsequent investigations found that tetramer organizing peptides could be traced to several proteins [22]. Reasons for the differences in the results are discussed in Biberoglu et al. [22]. For FBS AChE there also appear to be multiple sources. Therefore, no single gene encodes for all of the polyproline peptides identified in either soluble BChE or soluble AChE.

4.2. Noncovalent binding

The polyproline peptides are bound non-covalently to tetrameric FBS AChE. The peptides were easily released by denaturing the protein in boiling water, or by freezing the protein in the absence of a cryoprotectant. The interaction of polyproline with the tetramerization domain is very strong as the tetramer does not dissociate upon dilution down to nanomolar levels. Released peptides were visualized at 210 nm in an HPLC chromatogram and by mass spectrometry. However, the polyproline peptides were not detected on polyacrylamide gels stained with Coomassie blue or on gels reverse stained with imidazole-zinc [23].

4.3. A second gene product is required to make a tetramer

To date AChE and BChE are the only proteins known to require a polyproline peptide for assembly into tetramers. A search of the Protein Data Bank showed many tetrameric crystal structures but none included a polyproline peptide derived from a second gene. The common motif for tetramer organization is through interactions of side chains of the 4 subunits. AChE and BChE also have contacts between subunits but these are not sufficient to stabilize the tetramer. The cartoon in Fig. 5 indicates the location of a polyproline peptide in the center of the tetramer. This cartoon is presented to emphasize the overall organization of the AChE tetramer and to focus attention on the role of the tetramerization peptide in organizing this structure. The planar symmetry and large contact area between the polyproline peptide and the core of the AChE are not intended to realistically represent the details of the interaction.

Muscle and brain contain not only membrane bound forms of AChE but also a small amount of soluble tetrameric AChE. It is not known what type of proline-rich peptide is present in tetrameric

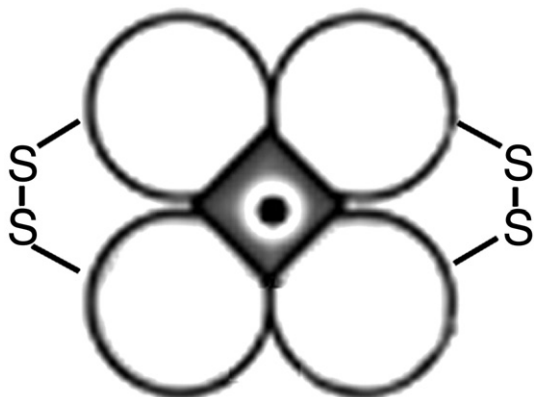


Fig. 5. Four identical subunits of fetal bovine serum AChE (accession number gi 108493 and gi 115497516) are linked by interactions with a polyproline peptide in the center of the tetramer. Dimers of AChE are formed by interchain disulfide bonds at Cysteine 580. The close contact between the globular subunits is intended to show that interactions between subunits contribute to the stability of the tetramer. Each AChE subunit contains 583 amino acids. The MW of the glycosylated tetramer is 340 kDa.

AChE in these tissues. We expect that future studies will identify additional tetrameric proteins that include a non-covalently bound polyproline peptide in the tetrameric structure.

4.4. The proline-rich peptides released from FBS AChE contain up to 25 residues

The crystal structure of the complex between synthetic human PRAD and 4 parallel chains representing the tetramerization domain showed that the coiled 15-residue ColQ PRAD peptide (LLTPP PPPLF PPPFF) has the same length as the helical 40-residue tetramerization domain of AChE [5]. The crystal structure of full-length tetrameric AChE complexed with a proline-rich peptide is not yet available. However, information from the crystal structure of synthetic peptides allowed Dvir et al. to produce a model of the AChE tetramer linked to the full length ColQ protein. Fig. 6 (adapted from Dvir et al.) shows that the ColQ protein traverses the entire AChE tetramer. The N-terminal region of ColQ interacts with the AChE C-terminal tetramerization domain in an anti-parallel head-to-tail configuration, passes through the center of the AChE tetramer and extends out the other side. This configuration will accommodate all sizes of polyproline peptides observed in our study, including the 25-residue polyproline peptide. A complete symmetry of the four subunit positions would not necessarily be expected since the tetramerization domains must accommodate to the helical structure of the polyproline.

4.5. Requirement for protease activity

Protease activity is not involved in the production of the membrane-anchored forms of AChE. In contrast, the polyproline peptides in plasma

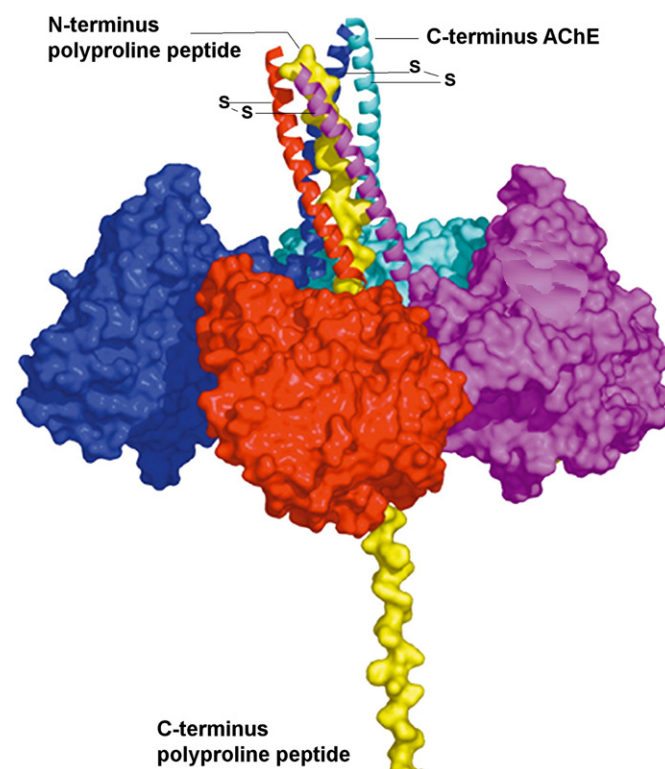


Fig. 6. Model of the AChE tetramer in the presence of the full-length ColQ protein. AChE monomers are displayed as globular units with their tryptophan-rich, C-terminal, amphiphilic α -helices protruding out from the top of each unit. The ColQ protein, or alternatively the tetramer organizing polyproline peptide (yellow), projects through the middle of the AChE tetramer complex. The N-terminus of ColQ interacts with the C-terminal tetramerization domain of AChE (at the top) and extends as a single chain through the bottom of the complex. Figure reproduced from Ref. [5].

AChE and plasma BChE derive from large precursor proteins that must undergo proteolysis to yield the proline-rich peptides. It is unknown whether the entire precursor protein is initially assembled together with the AChE tetramer followed by proteolysis, or whether proline-rich peptides are released from the precursor proteins before assembly with AChE subunits to yield AChE tetramers. The polyproline peptides released from FBS AChE range in size from 1280 to 2445 Da corresponding to 13 to 25 amino acid residues. The various sizes could originate from protease activity in plasma. Mass spectrometry analysis of the low molecular weight plasma proteome has demonstrated that proteases including aminopeptidase and carboxypeptidase convert large proteins to overlapping series of related peptides [24].

4.6. Assembly of the AChE tetramer

The presence of a set of different polyproline peptides extracted from the cores of soluble AChE tetramers raises a number of questions. First, could the variety of tetramer-organizing peptides reflect AChE expression from different tissues? Second, unlike membrane-bound AChE tetramers (which employ dedicated proteins that carry proline-rich tetramer organizing domains) the tetramer organizing peptides for soluble AChE appear to come from proteins designed for other purposes. How do these precursor proteins release the polyproline peptides for use in AChE? Is the polyproline peptide added to the oligomers prior to secretion or do these associations occur spontaneously in the bloodstream? Is there any evidence for the presence of putative precursor proteins in the endoplasmic reticulum and/or Golgi where interaction with developing AChE might be expected to occur? Third, since the AChE used in our experiments came from pooled serum, could the diversity in tetramer-organizing peptides that was observed represent variations between individuals?

Question 1. The origin of soluble AChE in fetal bovine serum is unknown. However, it is firmly established that soluble butyrylcholinesterase (BChE) in human serum originates in the liver [25]. It is widely accepted that the origin of mammalian serum cholinesterases, in general, is the liver. Though fetal bovine could differ, it would be unexpected to find serum AChE coming from tissues other than the liver.

Question 2. All of the bovine proteins that were found to match the tetramerization peptides were examined in detail with the goal of determining a mechanism by which the peptide could become associated with AChE. Their sequences were obtained from the NCBI nr Protein resource. Their functions, subcellular locations, and tissue distributions were obtained from UniProtKB protein knowledgebase.

In all, 11 bovine and 1 human protein(s), associated with the 5 peptide families, were examined. The proline-rich sequences from families 2, 3 and 5 are all located deep within the protein structure (97 to 1556 residues from the nearest terminus).

Nuclear proteins included: 1) zinc finger protein 318 (gi 194677641) an endocrine regulator protein expressed in endocrine tissue (containing a family 3 peptide), 2) lysine-specific demethylase 6B (gi 359076756) a histone demethylase with no specific tissue location (containing a family 3 peptide), 3) YLP motif-containing protein 1 (gi 297479917) involved in reduction of telomerase activity (containing a family 3 peptide), 4) zinc finger homeobox protein 4 (gi 296480588) a transcription regulator involved in neural and muscular differentiation expressed in brain, skeletal muscle, liver and stomach (containing both a peptide from family 2 and a peptide from family 5; different peptides), 5) R3H domain-containing protein 1 (gi 194664704) a nucleic acid binding protein (containing a family 2 peptide), and 6) myb-related p42POP (gi 359075807) a transcriptional repressor and DNA binding protein (containing a family 2 peptide).

Cytoplasmic proteins (all interacting with the cytoskeleton) included: 7) leiomodrin-2 (gi 157427900) an actin and tropomyosin binding

protein found in heart and skeletal muscle (containing a family 2 peptide), 8) protein diaphanous homolog 3, DIAPH3 (gi 296481927) a profilin and Rho binding protein that promotes actin polymerization with no tissue specificity (containing a family 3 peptide), and 9) formin-like 1 (gi 297487196) a regulator of cell morphology and cytoskeletal organization that binds to actin and Rho (containing a family 2 peptide).

Two of the remaining three protein matches (to families 1 and 2) are more encouraging in that they are associated with the endoplasmic reticulum and therefore could theoretically come into contact with AChE as it is being synthesized. UDP-N-acetylglucosamine transferase subunit ALG13 homology domain-containing protein 1 (gi 153791910) is involved with protein N-glycosylation in the endoplasmic reticulum. It contains the peptide from family 1. No proteins from the bovine taxonomy were matched to the family 1 peptide, therefore this protein from the human taxonomy has been taken as a credible substitute. The unfortunate feature of this protein is that the tetramerization peptide is located 192 residues from the C-terminus, and further from the N-terminus. The large proline-rich protein BAG6 (gi 161621275) is a chaperone associated with insertion of tail-anchored proteins into the endoplasmic reticulum. It contains a family 2 peptide. Like the previous candidate, the tetramerization peptide is located well from either terminus (464 residues from the C-terminus). It should be noted that both of these proteins are associated with the cytosolic, not the luminal face of the endoplasmic reticulum. Their topology presents a problem for interactions with AChE.

The most appealing candidate of the 12 is mesoderm-specific transcript homology protein (MEST) that is also referred to as paternally expressed 10 isoform 3 (gi 187761320) (to family 4). The tetramerization peptide is ideally suited for association with fetal bovine AChE in that it is located 5 residues from the C-terminus. One can easily envision the proline helix providing an anchor around which four AChE monomers could cluster. Once formed, the mature AChE tetramer could be released by proteolysis of the MEST C-terminus. A similar mechanism with other proteins similar to MEST could account for the other tetramerization peptides isolated from soluble AChE tetramers. The major drawback for this candidate is that it is found in the nucleus and cytoplasm.

Question 3. Because the AChE used in these experiments came from multiple animals, the possibility that the different polyproline peptides arose from variations between individual animals cannot be excluded.

More detailed investigation of these issues is a matter for future studies.

5. Conclusion

Soluble, tetrameric AChE molecules in plasma contain a polyproline peptide in their quaternary structure. Five types of polyproline peptide have been identified. These polyproline peptides are bound to the C-terminal tetramerization domain where they organize four AChE subunits into a tetramer. No single protein precursor is the source of these polyproline peptides.

Acknowledgements

This work was supported by a TÜBİTAK grant from the Scientific and Technological Research Council of Turkey to OT, a fellowship from Hacettepe University to KB, and NIH grant P30CA36727 to the Eppley Cancer Center.

The funding sources had no role in the study design; in collection, analysis and interpretation of the data; in writing of the report; or in the decision to submit the article for publication.

Mass spectra were obtained with the support of the Mass Spectrometry and Proteomics core facility at the University of Nebraska Medical Center.

References

- [1] J. Massoulie, L. Pezzementi, S. Bon, E. Krejci, F.M. Vallette, Molecular and cellular biology of cholinesterases, *Prog. Neurobiol.* 41 (1993) 31–91.
- [2] B. Li, J.A. Stribley, A. Ticu, W. Xie, L.M. Schopfer, P. Hammond, S. Brimijoin, S.H. Hinrichs, O. Lockridge, Abundant tissue butyrylcholinesterase and its possible function in the acetylcholinesterase knockout mouse, *J. Neurochem.* 75 (2000) 1320–1331.
- [3] G. Feng, E. Krejci, J. Molgo, J.M. Cunningham, J. Massoulie, J.R. Sanes, Genetic analysis of collagen Q: roles in acetylcholinesterase and butyrylcholinesterase assembly and in synaptic structure and function, *J. Cell Biol.* 144 (1999) 1349–1360.
- [4] A.L. Perrier, J. Massoulie, E. Krejci, PRiMA: the membrane anchor of acetylcholinesterase in the brain, *Neuron* 33 (2002) 275–285.
- [5] H. Dvir, M. Harel, S. Bon, W.Q. Liu, M. Vidal, C. Garbay, J.L. Sussman, J. Massoulie, I. Silman, The synaptic acetylcholinesterase tetramer assembles around a polyproline II helix, *EMBO J.* 23 (2004) 4394–4405.
- [6] S. Simon, E. Krejci, J. Massoulie, A four-to-one association between peptide motifs: four C-terminal domains from cholinesterase assemble with one proline-rich attachment domain (PRAD) in the secretory pathway, *EMBO J.* 17 (1998) 6178–6187.
- [7] S. Bon, J. Massoulie, Quaternary associations of acetylcholinesterase. I. Oligomeric associations of T subunits with and without the amino-terminal domain of the collagen tail, *J. Biol. Chem.* 272 (1997) 3007–3015.
- [8] H. Li, L.M. Schopfer, P. Masson, O. Lockridge, Lamellipodin proline rich peptides associated with native plasma butyrylcholinesterase tetramers, *Biochem. J.* 411 (2008) 425–432.
- [9] J.S. Ralston, R.S. Rush, B.P. Doctor, A.D. Wolfe, Acetylcholinesterase from fetal bovine serum. Purification and characterization of soluble G4 enzyme, *J. Biol. Chem.* 260 (1985) 4312–4318.
- [10] D. De la Hoz, B.P. Doctor, J.S. Ralston, R.S. Rush, A.D. Wolfe, A simplified procedure for the purification of large quantities of fetal bovine serum acetylcholinesterase, *Life Sci.* 39 (1986) 195–199.
- [11] G.L. Ellman, K.D. Courtney, V. Andres Jr., R.M. Featherstone, A new and rapid colorimetric determination of acetylcholinesterase activity, *Biochem. Pharmacol.* 7 (1961) 88–95.
- [12] C.W. Hung, A. Schlosser, J. Wei, W.D. Lehmann, Collision-induced reporter fragmentations for identification of covalently modified peptides, *Anal. Bioanal. Chem.* 389 (2007) 1003–1016.
- [13] D. Renner, G. Spiteller, Linked scan investigation of peptide degradation initiated by liquid secondary ion mass spectrometry, *Biomed. Environ. Mass Spectrom.* 15 (1988) 75–77.
- [14] R.P. Grese, R.L. Cerny, M.L. Gross, Metal ion-peptide interactions in the gas phase: a tandem mass spectrometry study of alkali metal cationized peptides, *J. Am. Chem. Soc.* 111 (1989) 2835–2842.
- [15] L.M. Teesch, J. Adams, Fragmentation of gas-phase complexes between alkali metal ions and peptides: metal ion binding to carbonyl oxygens and other neutral functional groups, *J. Am. Chem. Soc.* 113 (1991) 812–820.
- [16] T. Lin, G.L. Glish, C-terminal peptide sequencing via multistage mass spectrometry, *Anal. Chem.* 70 (1998) 5162–5165.
- [17] J.M. Barr, M.J. Van Stipdonk, Multi-stage tandem mass spectrometry of metal cationized leucine enkephalin and leucine enkephalin amide, *Rapid Commun. Mass Spectrom.* 16 (2002) 566–578.
- [18] X.J. Tang, W. Ens, K.G. Standing, J.B. Westmore, Daughter ion mass spectra from cationized molecules of small oligopeptides. 2: The proline effect in collision-induced dissociation of protonated Ala-Ala-Xxx-Pro-Ala (Xxx = Ala, Ser, Leu, Val, Phe, and Trp), *J. Am. Soc. Mass Spectrom.* 22 (2011) 1032–1039.
- [19] C. Bleiholder, S. Suhai, A.G. Harrison, B. Paizs, Towards understanding the tandem mass spectra of protonated oligopeptides. 2: The proline effect in collision-induced dissociation of protonated Ala-Ala-Xxx-Pro-Ala (Xxx = Ala, Ser, Leu, Val, Phe, and Trp), *J. Am. Soc. Mass Spectrom.* 22 (2011) 1032–1039.
- [20] A.G. Unnithan, M.J. Myer, C.J. Veale, A.S. Danell, MS/MS of protonated polyproline peptides: the influence of N-terminal protonation on dissociation, *J. Am. Soc. Mass Spectrom.* 18 (2007) 2198–2203.
- [21] Y. Wang, J. Johansson, W.J. Griffiths, Characterisation of variant forms of prophenin: mechanistic aspects of the fragmentation of proline-rich peptides, *Rapid Commun. Mass Spectrom.* 14 (2000) 2182–2202.
- [22] K. Biberoglu, L.M. Schopfer, O. Tacal, O. Lockridge, The proline-rich tetramerization peptides in equine serum butyrylcholinesterase, *FEBS J.* 279 (2012) 3844–3858.
- [23] C. Fernandez-Patron, E. Hardy, A. Sosa, J. Seoane, L. Castellanos, Double staining of Coomassie blue-stained polyacrylamide gels by imidazole–sodium dodecyl sulfate–zinc reverse staining: sensitive detection of coomassie blue-undetected proteins, *Anal. Biochem.* 224 (1995) 263–269.
- [24] J.M. Koomen, D. Li, L.C. Xiao, T.C. Liu, K.R. Coombes, J. Abbruzzese, R. Kobayashi, Direct tandem mass spectrometry reveals limitations in protein profiling experiments for plasma biomarker discovery, *J. Proteome Res.* 4 (2005) 972–981.
- [25] G.F. Khoury, J. Brill, L. Walts, R.W. Busuttill, Atypical serum cholinesterase eliminated by orthotopic liver transplantation, *Anesthesiology* 67 (1987) 273–274.
- [26] C. Donger, E. Krejci, A.P. Serradell, B. Eymard, S. Bon, S. Nicole, D. Chateau, F. Gary, M. Fardeau, J. Massoulie, P. Guicheney, Mutation in the human acetylcholinesterase-associated collagen gene, COLQ, is responsible for congenital myasthenic syndrome with end-plate acetylcholinesterase deficiency (Type Ic), *Am. J. Hum. Genet.* 63 (1998) 967–975.
- [27] E. Krejci, S. Thomine, N. Boschetti, C. Legay, J. Sketelj, J. Massoulie, The mammalian gene of acetylcholinesterase-associated collagen, *J. Biol. Chem.* 272 (1997) 22840–22847.
- [28] S. Bon, F. Coussen, J. Massoulie, Quaternary associations of acetylcholinesterase. II. The polyproline attachment domain of the collagen tail, *J. Biol. Chem.* 272 (1997) 3016–3021.
- [29] E. Krejci, F. Coussen, N. Duval, J.M. Chatel, C. Legay, M. Puype, J. Vandekerckhove, J. Cartaud, S. Bon, J. Massoulie, Primary structure of a collagenic tail peptide of Torpedo acetylcholinesterase: co-expression with catalytic subunit induces the production of collagen-tailed forms in transfected cells, *EMBO J.* 10 (1991) 1285–1293.
- [30] H. Noureddine, C. Schmitt, W. Liu, C. Garbay, J. Massoulie, S. Bon, Assembly of acetylcholinesterase tetramers by peptidic motifs from the proline-rich membrane anchor, PRiMA: competition between degradation and secretion pathways of heteromeric complexes, *J. Biol. Chem.* 282 (2007) 3487–3497.