The Role of Chitinase Production by Stenotrophomonas maltophilia Strain C3 in Biological Control of Bipolaris sorokiniana

Zhongge Zhang  
*University of Nebraska - Lincoln*

Gary Y. Yuen  
*University of Arizona*

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The Role of Chitinase Production by Stenotrophomonas maltophilia Strain C3 in Biological Control of Bipolaris sorokiniana

Zhongge Zhang and Gary Y. Yuen

ABSTRACT

Stenotrophomonas maltophilia strain C3 was selected originally as an antagonist of Rhizoctonia solani AG 1-IA, causal agent of brown patch disease in turf grass (6). In growth-chamber and field experiments on tall fescue (Festuca arundinacea) caused by Bipolaris sorokiniana, was investigated in vitro and in vivo. The filtrate of a broth culture of C3, with chitin as the carbon source, was separated into fractions. A high molecular-weight fraction (>8 kDa) was chitinolytic and more inhibitory than a low-molecular-weight, nonchitinolytic fraction to conidial germination and hyphal growth. The chitinolytic fractions caused swelling and vacuolation of conidia and discoloration, malformation, and degradation of germ tubes. When boiled, the chitinolytic fractions lost chitinase activity along with conidial germination and hyphal growth (>8 kDa) was chitinolytic and more inhibitory than a low-molecular-weight fraction to conidial germination and hyphal growth. A protein fraction derived by ammonium sulfate precipitation and a chitinase fraction purified by chitin affinity chromatography also were chitinolytic and highly anti-fungal. The chitinolytic fractions caused swelling and vacuolation of conidia and discoloration, malformation, and degradation of germ tubes. When boiled, the chitinolytic fractions lost chitinase activity along with conidial germination and hyphal growth.

Additional keywords: antagonism, antibiosis, microbial interactions, turf grass.


The role of chitinase production by Stenotrophomonas maltophilia strain C3 in biological control of leaf spot on tall fescue (Festuca arundinacea) caused by Bipolaris sorokiniana, was investigated in vitro and in vivo. The filtrate of a broth culture of C3, with chitin as the carbon source, was separated into fractions. A high molecular-weight fraction (>8 kDa) was chitinolytic and more inhibitory than a low-molecular-weight, nonchitinolytic fraction to conidial germination and hyphal growth. The chitinolytic fractions caused swelling and vacuolation of conidia and discoloration, malformation, and degradation of germ tubes. When boiled, the chitinolytic fractions lost chitinase activity along with conidial germination and hyphal growth.

The filamentous fungus Bipolaris sorokiniana (B. sorokiniana) is a major pathogen of numerous grass species and the causal agent of brown patch disease in turf grasses. An aqueous extract of leaves colonized by a chitinase-deficient mutant of C3, with and without added chitin, was no higher than the background, and the extracts lacked anti-fungal activity. Chitinolytic microbes upon addition of chitin to soil was associated with increased chitinase activity as measured by the release of N-acetylglucosamine. Chitinase production by applied biocontrol agents on plant tissues has not been demonstrated by direct detection or quantification.

Most of the antifungal properties. Two chitinase-deficient and two chitinase-reduced mutants of C3 were compared with the wild-type strain for inhibition of germination of B. sorokiniana conidia on tall fescue leaves and for suppression of leaf spot development in vivo. The mutants exhibited reduced antifungal activity but did not lose all biocontrol activity. An aqueous extract of leaves colonized by wild-type C3 had higher chitinase activity than that of noncolonized leaves and was inhibitory to conidial germination. The addition of chitin to leaves along with the wild-type strain increased both chitinase and antifungal activity. The chitinase activity level of extracts from leaves colonized by a chitinase-deficient mutant of C3, with and without added chitin, was no higher than the background, and the extracts lacked anti-fungal activity. Chitinolytic microbes upon addition of chitin to soil was associated with increased chitinase activity as measured by the release of N-acetylglucosamine. Chitinase production by applied biocontrol agents on plant tissues has not been demonstrated by direct detection or quantification.

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The objective of this study was to determine the relationship between chitinolytic and antagonism by strain C3 against B. sorokiniana. Two approaches were used, the first being the evaluation of chitinolytic factors excreted by strain C3 in vitro for effects on pathogen growth and disease development, and the second being a comparison between wild-type C3 and chitinase-deficient mutants with respect to chitinase production and pathogen inhibition in vivo. Preliminary reports have been published (29–31).
MATERIALS AND METHODS

General microbial and plant-culturing methods. Strain C3 and its derivatives were stored at −70°C in nutrient broth containing 10% glycerol. For each experiment, a fresh culture of a strain was grown on tryptic soy agar (TSA; Sigma Chemical Co., St. Louis) for 24 h at 25°C. Cells were suspended in sterile 0.01 M potassium phosphate buffer, pH 7.0, and the suspension was used for inoculating broth cultures and plant materials. Cell density in every suspension was adjusted to 10^8 CFU/ml by measuring turbidity in a spectrophotometer (Spectronic 20, Spectronic Instruments, Inc., Rochester, NY) and diluting with sterile phosphate buffer to the desired concentration. A spontaneous rifampicin derivative of strain C3 (6) was used in all experiments. Its population size was determined by dilution plating on TSA amended with rifampicin (Sigma) at 100 µg/ml.

*B. sorokiniana*, isolated in Nebraska from Kentucky bluegrass (*Poa pratensis*), was used for spore germination experiments and plant inoculations. Culturing conditions and procedures for producing inoculum in the form of conidial suspensions were described previously (32).

Tall fescue cv. Kentucky-31 was used in experiments on in vivo chitinase production and antagonism by bacterial strains. Seeds were planted in a steam-pasteurized potting medium (Sharpsburg silty-clay loam, vermiculite, and sand mixed in equal volumes) contained in 15-cm pots (200 seeds per pot). The plants were grown for over 45 days in a greenhouse under natural light at an average temperature of 25°C and watered daily with a nutrient solution (Peters 20-10-20, Scotts-Sierra Horticultural Co., Marysville, OH) containing N at 0.2 g/liter. Plants were cut to a height of 5 cm every 10 days.

Preparation of culture filtrate fractions. For experiments on chitinase production in vitro, strain C3 was cultured in a broth medium containing (NH₄)₂SO₄ at 1.0 g/liter, MgSO₄·7H₂O at 0.3 g/liter, and KH₂PO₄ at 1.36 g/liter (16). Colloidal chitin, prepared from practical-grade chitin (Sigma) by the method of Roberts and Selintrennikoff (20), was added to 5 g/liter as a carbon source. Yeast extract (Sigma) was autoclaved separately and added to sterilized broth to 0.5 g/liter. Four replicate cultures (50-ml volumes in 250-ml Erlenmeyer flasks) were incubated for 5 days at 25°C with shaking (180 rpm; Radial Shaker model 4710, Queue Systems, Parkersburg, WV), and centrifuged (J2-21, Beckman Instruments, Inc., Palo Alto, CA) at 15,000 × g for 20 min to remove cells and debris. The supernatants were sterilized by filtration through 0.22-µm cellulose nitrate membranes (Nalge Nunc International, Rochester, NY) and pH adjusted to 7.0 with 1 N HCl. Portions of each wash were used in determining population size of the applied strain. The remainder of the wash was centrifuged (15,000 × g, 20 min), and the supernatant was filtered through 0.22-µm membranes. The filtrates were stored at −20°C before being assayed for chitinase activity and antifungal properties. The washed leaves were dried at 60°C for 3 days and weighed so chitinase and population-size measurements could be expressed on a sample dry weight basis.

Enzyme activity assays. Chitinase production by bacterial strains in culture was detected by streaking or spotting strains on TSA amended with 0.2% colloidal chitin. The cultures were incubated at 25°C and examined periodically for clear zones of hydrolysis around colonies. Chitinase activity in cell-free liquids was assessed by two methods. In one method, 50-µl aliquots were pipetted into wells (0.4 cm diameter) made in water agar containing 0.2% colloidal chitin. The plates were examined for the clarity, shape, and size of hydrolytic zones after 3 days of incubation at 25°C. In the other method, chitinolysis was measured through the hydrolysis of *p*-nitrophenyl-β-1,3-N-acetylchitobiose (Sigma), releasing *p*-nitrophenol as a yellow product (20). Aliquots (10 µl) of test fluids were added to 60 µl of 0.2 mM *p*-nitrophenyl reagent in 50 mM potassium phosphate buffer, pH 6.0, in duplicate wells of a microplate. After the mixture was incubated at 37°C for 30 min, the reaction was terminated by adding 10 µl of 1 N NaOH to each well. The amount of *p*-nitrophenol produced was determined by measuring the absorbance of the fluids at 405 nm with an Optimax Microplate Reader (Molecular Devices Corp., Sunnyvale, CA).
TABLE 1. Chitinolysis by various fractions of filtrate from a culture of *Stenotrophomonas maltophilia* strain C3 and effects of fractions on germination of *Bipolaris sorokiniana* conidia in vitro and development of leaf spot on tall fescue

<table>
<thead>
<tr>
<th>Filtrate fraction¹</th>
<th>Heat treatment²</th>
<th>Chitinolysis³</th>
<th>Germination (%) ⁴</th>
<th>Lesions/10 cm of leaf⁵</th>
<th>Diseased leaf area (%) ⁶</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole filtrate</td>
<td>–</td>
<td>+</td>
<td>8 d</td>
<td>0 d</td>
<td>1 d</td>
</tr>
<tr>
<td>Whole filtrate</td>
<td>+</td>
<td>–</td>
<td>58 c</td>
<td>NT</td>
<td>NT</td>
</tr>
<tr>
<td>Protein fraction</td>
<td>–</td>
<td>+</td>
<td>9 d</td>
<td>16 c</td>
<td>9 c</td>
</tr>
<tr>
<td>Protein fraction</td>
<td>+</td>
<td>–</td>
<td>51 c</td>
<td>31 a</td>
<td>33 b</td>
</tr>
<tr>
<td>High molecular-weight fraction</td>
<td>–</td>
<td>+</td>
<td>9 d</td>
<td>1 d</td>
<td>2 d</td>
</tr>
<tr>
<td>High molecular-weight fraction</td>
<td>+</td>
<td>–</td>
<td>53 c</td>
<td>10 c</td>
<td>10 c</td>
</tr>
<tr>
<td>Low molecular-weight fraction</td>
<td>–</td>
<td>–</td>
<td>80 b</td>
<td>24 b</td>
<td>28 b</td>
</tr>
<tr>
<td>Phosphate buffer control</td>
<td>–</td>
<td>–</td>
<td>92 a</td>
<td>36 a</td>
<td>41 a</td>
</tr>
</tbody>
</table>

¹ Values in a column followed by the same letter are not significantly different (P = 0.01) according to Duncan’s multiple range test; NT = not tested.
² Whole filtrate was prepared from 5-day-old broth culture of C3 by removing cells by centrifugation and filter sterilization. The protein fraction was derived by ammonium sulfate precipitation and dissolution in phosphate buffer. Low and high molecular-weight fractions were prepared from whole filtrate by ultrafiltration through a 8,000-Da membrane.
³ + and – indicate presence and absence, respectively, of clear hydrolysis zones on water agar containing colloidal chitin.
⁴ Germination was determined after conidia were incubated in filtrate fractions for 20 h at room temperature. Values are means of two experiments, each with four replications per treatment.
⁵ Lesion frequency and diseased leaf area were measured 4 and 7 days, respectively, after inoculation. Values are means of two experiments, each with four replications per treatment.

Fig. 1. Nongerminated and germinated conidia of *Bipolaris sorokiniana* after 20 h of incubation in A and C, phosphate buffer and B and D, high molecular-weight fraction of a filtrate from a culture of *Stenotrophomonas maltophilia* strain C3. All images were photographed at 410×.
Fisher’s least significant difference (LSD) and Duncan’s multiple range tests were used for mean separation. Percentage data (i.e., conidial germination and disease severity) were analyzed with and without arc sine transformation, but only results involving non-transformed data are reported because the transformation did not affect the statistical analysis.

**RESULTS**

**Effects of C3 culture filtrate fractions on B. sorokiniana in vitro.** The raw culture fluid and the high molecular-weight and protein fractions of the culture fluid were chitinolytic, producing clear hydrolytic zones when they were applied to water agar containing chitin (Table 1). Boiling these fluids for 10 min resulted in loss of chitinolytic activity. The high molecular-weight and protein fractions were as inhibitory to germination of *B. sorokiniana* conidia as the raw culture filtrate. Heating of these fractions significantly reduced, but did not completely eliminate, their inhibitory properties. The low molecular-weight fraction was not chitinolytic and only slightly inhibitory to spore germination.

*B. sorokiniana* conidia that failed to germinate after exposure to the high molecular-weight or protein fractions appeared swollen and vacuolated. In some of the treated conidia that did germinate, the germ tubes stopped growing before their lengths equaled the width of the spores. In others, the hyphae exhibited swelling of the tips, highly irregular branching, and pronounced cell alterations, such as swelling, browning, and shortening, compared with hyphae in the nontreated controls (Fig. 1). The low molecular-weight fraction did not have these effects on the conidia or hyphal development (data not shown).

**Effects of C3 culture fractions on Bipolaris leaf spot disease.** When applied to tall fescue plants, the culture filtrate and all the fractions in nonheated form suppressed leaf spot disease compared with the control (Table 1). Only the high molecular-weight fraction reduced lesion frequency, measured on day 4, to the same extent as the whole filtrate. Both the high molecular-weight fraction and the protein fraction were similar to the whole filtrate in reducing disease severity by day 8. Lesion frequency and disease severity on plants treated with the low molecular-weight fraction were significantly higher than with the other two fractions. When boiled for 10 min, the protein and high molecular-weight fractions lost some, but not all, of their inhibitory capacity, because lesion frequency and disease severity levels in the treatments with boiled fractions were significantly lower than in the control.

The culture fluid fraction that was eluted from a chitin affinity column by acetic acid was highly chitinolytic, with nearly a 10-fold increase in specific activity compared with the raw culture fluid. That fraction inhibited *B. sorokiniana* conidial germination and germ tube elongation compared with the phosphate buffer control (Table 2). However, it was less inhibitory to *B. sorokiniana* than the raw culture fluid at the same protein concentration.

**Disease suppression and leaf colonization by chitinase-altered mutants.** Of the 6,000 chloramphenicol-resistant derivatives tested for chitinolysis on colloidal chitin-amended TSA, 6 did not produce hydrolysis zones, and 42 formed smaller clearing zones than the wild-type strain. Two strains lacking any detectable chitinase activity (1-53 and 7-41) and two with reduced chitinase activity (20-14 and 24-8) were selected for further investigation because they were similar to the wild-type strain in growth rate and colony morphology. Chloramphenicol resistance and altered chitinase activity were stable in the mutant strains through repeated transfers. All four mutant strains were reduced in lipase activity, and three were reduced in β,1,3 glucanase activity compared with the wild-type (Table 3). Protease production in the mutant strains was unaffected, and antagonism against other bacteria was unchanged or increased.

When applied to tall fescue plants, all of the mutants inhibited stages of Bipolaris leaf spot disease development compared with the phosphate control (Table 3), but none were as effective as the wild-type strain. Although the spore germination levels varied among the four mutant strains, with the chitinase-reduced strain 20-14 having the lowest, there was no difference in disease inhibition between the strains deficient in chitinase activity and those reduced in chitinase activity. The mutants, in general, did not colonize tall fescue leaves over the long term to the same level as the wild-type. In one experiment, population sizes of three mutants were ≥1 log unit lower than the wild-type (Fig. 2). Similar results were obtained in the repetition of the experiment (data not shown).

**Chitinase production and antifungal activity on leaves.** A chitinase-deficient mutant, 7-41, was compared with the wild-type for chitinase production in vivo. Significantly higher amounts of chitinase activity were detected in extracts of tall fescue leaves colonized for 4 days by strain C3 than in extracts of leaves without C3 (Table 4). The application of colloidal chitin with C3 fur-

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**TABLE 2.** Effects of filtrates of a culture of *Stenotrophomonas maltophilia* strain C3 and a chitinase fraction derived by chitin affinity chromatography on conidial germination and germ tube elongation by *Bipolaris sorokiniana*.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Chitinase activity</th>
<th>Germination (%)</th>
<th>Germ tube length (µm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raw culture filtrate</td>
<td>0.14</td>
<td>10 c</td>
<td>2 c</td>
</tr>
<tr>
<td>Chitinase fraction a</td>
<td>1.21</td>
<td>30 b</td>
<td>8 b</td>
</tr>
<tr>
<td>Phosphate buffer</td>
<td>0.00</td>
<td>81 a</td>
<td>19 a</td>
</tr>
</tbody>
</table>

a Values in a column followed by the same letter are not significantly different (P = 0.01) according to a least significant difference test.

b Chitinase activity was measured in units per milligram of protein by hydrolysis of p-nitrophenyl-N,N'-diacetylchitobiose.

c Germination was determined after conidia were incubated in culture fluid fractions or phosphate buffer for 20 h at room temperature. Each fraction was adjusted to a protein concentration of 0.25 mg/ml prior to antifungal assay. Values are means of four replications.

d Fraction eluted from chitin affinity chromatography column by 20 mM acetic acid (pH 3.0) and adjusted to pH 5.5.

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**TABLE 3.** Comparison of wild-type and chitinase-deficient mutants of *Stenotrophomonas maltophilia* strain C3 for lytic enzyme activity, bacterial antagonism, and effects on germination of conidia of *Bipolaris sorokiniana* and development of leaf spot on tall fescue.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Chitinase</th>
<th>β-1,3-glucanase</th>
<th>Lipase</th>
<th>Protease</th>
<th>Bacterial antagonism b</th>
<th>Germination (%) c</th>
<th>Lesions/10 cm of leaf c</th>
<th>Diseased leaf area (%) c</th>
</tr>
</thead>
<tbody>
<tr>
<td>C3</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>11 d</td>
<td>10 c</td>
<td>2 c</td>
</tr>
<tr>
<td>1-53</td>
<td>–</td>
<td>++</td>
<td>–</td>
<td>++</td>
<td>++</td>
<td>79 b</td>
<td>41 b</td>
<td>22 b</td>
</tr>
<tr>
<td>7-41</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>++</td>
<td>86 a</td>
<td>36 b</td>
<td>20 b</td>
</tr>
<tr>
<td>20-14</td>
<td>+</td>
<td>+</td>
<td>++</td>
<td>++</td>
<td>++</td>
<td>68 c</td>
<td>33 b</td>
<td>19 b</td>
</tr>
<tr>
<td>24-8</td>
<td>+</td>
<td>+</td>
<td>++</td>
<td>++</td>
<td>+++</td>
<td>81 b</td>
<td>38 b</td>
<td>25 b</td>
</tr>
<tr>
<td>None</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>87 a</td>
<td>52 a</td>
<td>38 a</td>
</tr>
</tbody>
</table>

a, –, +, ++, and +++ indicate none, reduced, same, and increased activity, respectively, relative to wild-type strain C3. Values in a column followed by the same letter are not significantly different at P = 0.05 for germination and P = 0.01 for lesion frequency and diseased leaf area, according to Duncan’s multiple range test.

b Antagonism was determined in a plate inhibition test against *Clavibacter michiganensis* subsp. *nebraskensis* and *Pseudomonas syringae* pv. *tomato*. Results were the same against both target species.

c Percent germination, lesion frequency, and percent diseased leaf area were determined 1, 4, and 8 days, respectively, after inoculation with conidia of *B. sorokiniana*. All values are means of two experiments, each with four replications per treatment.
ther increased chitinase activity. The presence of strain 7-41 on leaves, with or without chitin, did not affect chitinase activity levels when compared with the respective no-bacteria controls.

The population density of wild-type C3 measured in the leaf extract was significantly higher than that of the chitinase-deficient mutant (Table 4). This reflected the population size of the wild-type on grass leaves increasing by 0.5 log units, while that of 7-41 remained unchanged from the starting level over the 4-day experiment period. Neither strain was affected in colonization by addition of chitin.

The antifungal activity of the various leaf extracts, measured on the basis of their effect on germination of *B. sorokiniana* conidia, was related to the relative levels of chitinase activity (Table 4). Extracts from leaves colonized by 7-41 were the same as the controls in allowing high levels of conidial germination. Extracts from C3-colonized leaves, with and without chitin, significantly inhibited germination, with the chitin amendment producing the highest degree of inhibition.

**DISCUSSION**

We conclude that chitinolysis is one mechanism involved in the biological control of *B. sorokiniana* by strain C3. One line of evidence is the close association observed between chitinolytic factors produced by C3 in vitro and inhibition of spore germination and disease development. The major components in culture fluids active in fungal inhibition and disease suppression appeared to be heat-labile proteins rather than antibiotic compounds of low molecular weight. The observed alteration of germ tube and hyphal morphology by the chitinolytic high molecular-weight and protein fractions also support the hypothesis that chitinase was involved. Swelling, browning, branching, and degradation of germ tubes were reported as symptoms in fungi exposed to chitin or chitinolytic microorganisms (1,14,18). Kritzman et al. (12) showed that the active sites for chitinases are the tips, septa, and branches of hyphae.

As supportive evidence that chitinolysis is involved in antagonism by C3, chitinase-deficient and -reduced mutants of C3 were less disease suppressive than the wild-type. The mutants strains exhibited reduced colonization ability over the long term compared with the wild-type strain. However, pathogen inoculation was performed 1 day after bacterial treatment; therefore, the difference in population numbers at inoculation was small and could not account entirely for the large reduction in antifungal activity and disease suppressiveness observed in the mutants. Extracellular enzyme activity in strain C3 was globally regulated (D. Kobayashi, personal communication); we suspect that global regulator sites were affected in mutants because they also had reduced β-glucanase and lipase activity. Furthermore, strain C3 produced multiple chitinases in a preliminary study (33); thus, it was unlikely that the mutations in the chitinase-deficient strains involved structural genes. We isolated the mutants from a chloramphenicol-amended medium in the process of generating transposon mutants. Although it is not clear what the connection between selection for antibiotic resistance and mutation in global control sites is, spontaneous gac regulatory mutants in *P. aeruginosa* and *P. fluorescens* were isolated under similar circumstances (3,13).

Strong evidence for the importance of chitinase production by strain C3 is the detection of chitinase activity in extracts derived from C3-colonized leaves at levels that were well above that in non-colonized leaves. Furthermore, the extracts from C3-colonized leaves were antifungal, indicating that chitinase produced on leaves was sufficient in concentration to have a biological effect. The finding that colonization by a chitinase-deficient mutant did not result in chitin levels above the background strongly supports the hypothesis that strain C3 produced chitinases on leaf surfaces, but we cannot rule out the possibility that the chitinase might also have been of plant origin. Roby et al. (21) reported that soluble oligomeric products of chitin hydrolysis, rather than insoluble polymers of chitin, induced plant defense proteins such as chitinases and, thus, the highly elevated chitinase activity occurring when C3 was supplied with chitin could have been due in part to host chitinase production stimulated by chitin hydrolysis by C3. Western blot techniques using antibodies specific to bacterial chitinases or plant chitinases might be useful in answering the question as to the chitinase source.

We do not consider chitinolysis to be the sole mechanism by which strain C3 can affect *B. sorokiniana* and leaf spot development. Protease was an important factor in inhibition of *Pythium ultimum* by a rhizosphere strain of *Stenotrophomonas maltophilia* (5). Maltophilin, an antibiotic with antifungal activity, was reported from another rhizosphere strain (7), and xanthobaccins were implicated in the suppression of *Pythium* damping-off disease of sugar beet by a strain of *Stenotrophomonas* sp. (17). In this study, strain C3 excreted β-1,3-glucanase, protease, and lipase in vitro. Antifungal activity of culture fluid fractions, in general, was related to chitinase activity level, but fractions rendered nonchitinolytic and low molecular-weight components also had some degree of fungal activity.

**TABLE 4.** Chitinase activity and population sizes detected in extracts of tall fescue leaves colonized by wild-type *Stenotrophomonas maltophilia* strain C3 and a chitinase-deficient mutant (7-41) with and without chitin and effects of leaf extracts on germination of conidia of *Bipolaris sorokiniana* in vitro

<table>
<thead>
<tr>
<th>Treatment†</th>
<th>Chitinase units/ g of leaf‡</th>
<th>Log CFU/ g of leaf‡</th>
<th>Conidial germination (%)§</th>
</tr>
</thead>
<tbody>
<tr>
<td>Phosphate buffer</td>
<td>7.1 d</td>
<td>0.00</td>
<td>86 a</td>
</tr>
<tr>
<td>Chitin</td>
<td>8.6 c,d</td>
<td>0.00</td>
<td>83 a</td>
</tr>
<tr>
<td>Strain 7-41</td>
<td>10.4 c,d</td>
<td>7.19 b</td>
<td>82 a</td>
</tr>
<tr>
<td>Strain 7-41 + chitin</td>
<td>11.6 c</td>
<td>7.16 b</td>
<td>79 a</td>
</tr>
<tr>
<td>Strain C3</td>
<td>19.3 b</td>
<td>8.04 a</td>
<td>65 b</td>
</tr>
<tr>
<td>Strain C3 + chitin</td>
<td>41.6 a</td>
<td>8.08 a</td>
<td>41 c</td>
</tr>
</tbody>
</table>

† Extracts in phosphate buffer were made from leaves collected 4 days after treatment were applied to tall fescue plants. Values in a column followed by the same letter are not significantly different (P = 0.05) according to Duncan’s multiple range test.

‡ All treatments were suspended in phosphate buffer and applied to tall fescue leaves.

§ Chitinase activity was measured by hydrolysis of p-nitrophenyl-β,N,N′-diacetetylchitobiose. Values are means of two experiments with three replications each.

¶ C3 and 7-41 were applied to tall fescue plants to 7.5 log CFU/ g of leaf. Results from phosphate buffer and chitin treatments were not included in statistical analysis. Values are means of two experiments with three replications each.

*Germination of conidia was determined after incubation for 20 h in extracts. Values are means of two experiments with three replications each.*
inhibition. The chitin-affinity chromatography fraction containing only chitin-binding proteins was highly chitinolytic but was not as inhibitory as the raw culture fluid, which also points to the involvement of multiple factors. Finally, the mutant strains deficient in inhibitory as the raw culture fluid, which also points to the involvement of multiple factors. Finally, the mutant strains deficient in inhibitory activity could be important in using strain C3 in controlling these pathogens as well.

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