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GROWTH, CARCASS AND PALATABILITY TRAITS OF INTACT MALES AND STEERS IMPLANTED WITH ZERANOL OR ESTRADIOL EARLY AND THROUGHOUT LIFE

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ABSTRACT

This study was conducted to assess the impact of implanting intact beef males with protein anabolic agents at varying intervals throughout life. Ninety-six intact males were assigned to three implant treatments: 1) not implanted, 2) implanted at 9 wk of age, weaning and at 56-d intervals thereafter with a 36-mg zeranol implant or 3) estradiol implant at 9 wk of age and 68 d post-weaning. During the 118-d, post-weaning growing period, eight animals per treatment (one replication) were castrated. After a 114-d finishing period, cattle were slaughtered (average age of 13 to 14 mo). Feedlot performance, carcass and palatability data were obtained. Average daily gains and feed efficiency did not differ (P>.05) between zeranol and estradiol-implanted intact males. Regardless of implant treatment, steers had lighter carcass weights (P<.05) and higher (P<.01) quality grades than intact males. Implanting either intact males or steers with zeranol or estradiol resulted in higher (P<.05) numerical yield grades. Quality grades were higher in zeranol-implanted cattle than the non-implanted or estradiol-implanted cattle. Intact males implanted with zeranol were similar in carcass fatness to zeranol-implanted steers. No differences (P>.05) in tenderness or connective tissue were detected. Implanting intact males early and throughout life with zeranol made them similar to steers in fatness, while estradiol implantation had few effects on carcass and palatability traits of intact males or steers.

(Key Words: Growth Promoters, Bulls, Steers.)

Introduction

Considerable research has been conducted to examine the efficiency of feeding young intact males for slaughter. Generally, it has been reported that young intact males gain faster and more efficiently than do steers (Seideman et al., 1982). However, recent research also has emphasized that, when fed to the same compositional endpoint, steers can attain the desired endpoint more quickly and efficiently than can intact males (Crouse et al., 1984). In an effort to capitalize on the growth rates of young intact males and to produce meat of similar composition and palatability to steers, recent attention has focused on the use of growth-promoting implants in young intact males. In general, three regimens of implantation have been investigated: implanting at the beginning of the finishing phase (Gregory et al., 1983; Johnson et al., 1984; Schanbacher et al., 1984); at or near weaning and during the growing and finishing periods (Forrest, 1975; Johnson and Gee, 1982; Newland et al., 1984; Vanderwert et al., 1984); or implanting near birth and at pre-planned intervals throughout life (Ralston, 1978; Lamm et al., 1980; Brethour, 1982; Greathouse et al., 1983; Gray et al., 1984; Unruh et al., 1984). Most of this recent research deals with zeranol as a growth promotant. The utility of lifelong implantation with estradiol has not been investigated. Most research with estradiol deals with implanting animals after weaning, during the growing and(or) finishing periods. Results from the previously cited papers emphasize two concepts: first, the age of the animal and the duration of exposure to hormone-implant treatments are important in determining the magnitude of response; and second, that little agreement has been reached regarding the efficacy of implanting intact males with the protein anabolic agents zeranol or estradiol. With the current heightened interest in use of estradiol-17β as a growth promotant, and an absence of informa-

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tion regarding the effects of lifelong implantation of intact males with this hormone, our research was conducted to assess the impact of implanting young intact males, early and throughout life, with selected growth promoters on growth, carcass and palatability traits.

**Experimental Procedure**

Ninety-six intact males from primiparous Angus x Hereford dams and MARC II sires (one-fourth each of Angus, Gelbvieh, Hereford and Simmental) were assigned randomly to one of three treatments: 1) not implanted, 2) zereanol (Ralgro®, implanted five times with a 36-mg implant; at 9 wk of age, at weaning and at 56-d intervals thereafter) or 3) estradiol-17β (Compudose®, 200-d implant, implanted twice; at 9 wk of age and 68 d post-weaning). Following weaning, at about 6 mo of age, the calves were allotted to four replications of each treatment and fed a corn-and-silage based growing diet (calculated net energy for maintenance was 1.78 Mcal/kg of dry matter; net energy for gain was 1.18 Mcal/kg of dry matter) for 118 d. One replication of each treatment was castrated 70 d after the beginning of the growing period (about 9 mo of age) to provide a steer comparison for carcass data. The cattle were fed a high-energy finishing diet (calculated net energy for maintenance was 1.97 Mcal/kg of dry matter; net energy for gain was 1.35 Mcal/kg of dry matter) for 114 d prior to slaughter at 13 to 14 mo of age. At the start of the growing and finishing periods, a full weight was taken on each animal to facilitate calculation of average daily gain (ADG) and feed efficiency (kg of dry matter per kg of gain). Final weights were calculated from hot carcass weight on the basis of 62% dressing percent.

Blood samples were obtained from all animals on d 63 during the growing period and d 1, 49 and 114 during the finishing period. Serum testosterone concentrations were determined by the radioimmunoassay techniques of Kiser et al. (1978).

The cattle were shipped about 400 km to a commercial slaughter plant and slaughtered immediately upon arrival. After a 48-h chilling period, USDA quality and yield grade factors were obtained along with muscle color, firmness and texture scores. Wholesale ribs were obtained from the steer carcasses (n=8 per implant group) and from randomly selected carcasses from intact males (n=12 per implant group).

The ribs were marked for removal of the 9-10-11th rib sections according to the procedures of Hankins and Howe (1946) and shipped to the University of Nebraska meat laboratory. At 10 d postmortem, the ribs were cut into 9-10-11th rib sections, frozen, and later thawed and dissected into separable lean, fat and bone. The separable lean and fat fractions were analyzed separately for fat and moisture using standard AOAC procedures (1975). Following the procedures of Hankins and Howe (1946), the percentage of ether-extractable fat of the edible portion of the dressed carcass was calculated based on the percentage of fat in the rib section and upon the percentage of fat in the longissimus muscle.

At the time of rib section removal, individual longissimus muscle steaks were obtained for determination of total and heat soluble collagen by the procedures of Hill (1966) and Bergman and Loxley (1963).

Steaks were obtained from the wholesale ribs, frozen and later thawed for cooking. Steaks were cooked on roasting pans in a 177 C rotary hearth oven to an internal temperature of 70 C (monitored by small diameter, copper-constantan thermocouples). Steaks were turned over when they reached 35 C internal temperature. Samples were then evaluated by a trained (Cross et al., 1978) sensory panel for juiciness, muscle fiber tenderness and connective tissue amount. Ratings were made on a 15-cm unstructured line scale with the extremes labeled as tough, abundant connective tissue or dry on one end of the line and tender, no connective tissue or juicy on the opposite end. Warner-Bratzler shear force values were obtained on 1.3-cm cores using an Instron Universal Testing Machine.

Data were analyzed using the Statistical Analysis System (SAS, 1982). Analysis of variance was conducted using a model which contained replication, sex, implant treatment and sex x implant treatment as sources of variation. Experimental error and sampling error were pooled, where appropriate. Significant interactions or, in their absence, main effects were evaluated using the new Duncan's multiple-range procedure to test for differences among treatment means.

**Results and Discussion**

**Growth Traits.** Table 1 presents the production data for intact males. Since only one replication of steers was fed, production data of
TABLE 1. EFFECTS OF IMPLANTS ON PERFORMANCE OF INTACT MALES

<table>
<thead>
<tr>
<th>Trait</th>
<th>Control</th>
<th>Zeranol</th>
<th>Estradiol</th>
<th>SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Weaning wt, kg</td>
<td>164</td>
<td>168</td>
<td>171</td>
<td>4.4</td>
</tr>
<tr>
<td>Finished wt, kg</td>
<td>472</td>
<td>490</td>
<td>488</td>
<td>8.9</td>
</tr>
<tr>
<td>Growing ADG, kg/d</td>
<td>1.43b</td>
<td>1.54c</td>
<td>1.48bc</td>
<td>.07</td>
</tr>
<tr>
<td>Finishing ADG, kg/d</td>
<td>4.23b</td>
<td>4.19b</td>
<td>4.46c</td>
<td>.06</td>
</tr>
<tr>
<td>Growing F/Ga</td>
<td>1.24</td>
<td>1.39</td>
<td>1.26</td>
<td>.08</td>
</tr>
<tr>
<td>Finishing F/Ga</td>
<td>5.95</td>
<td>6.25</td>
<td>6.16</td>
<td>.21</td>
</tr>
<tr>
<td>Combined ADG, kg/d</td>
<td>1.33</td>
<td>1.39</td>
<td>1.37</td>
<td>.06</td>
</tr>
<tr>
<td>Combined F/Ga</td>
<td>1.60</td>
<td>1.78</td>
<td>1.54</td>
<td>.08</td>
</tr>
<tr>
<td>Testicle wt, g</td>
<td>452b</td>
<td>186c</td>
<td>235c</td>
<td>20.3</td>
</tr>
<tr>
<td>Testosterone, ng/ml</td>
<td>10.5b</td>
<td>4.9d</td>
<td>6.5c</td>
<td>.54</td>
</tr>
</tbody>
</table>

\(a\) F/G is kg of dry matter intake per kg of gain. 
\(b, c, d\) Means in the same row that do not have a common superscript are different (P<.05).

Steers were not evaluated. Weaning and finishing weights did not differ among treatments. Cattle implanted with zeranol gained faster (P<.05) during the growing period than the non-implanted intact males but were not more efficient. Cattle implanted with zeranol did not gain faster (P>.05) than those implanted with estradiol but were more efficient during the growing period. There were no differences in gains or efficiencies between intact males receiving the different implant treatments during the finishing period, or for the growing and finishing periods combined. Vanderwert et al. (1984) reported similar results, but the cattle in their study were not implanted until the post-weaning growth period was initiated. Using estradiol implants post-weaning, Newland et al. (1984) reported no early advantage in post-weaning ADG; when the overall growing and finishing periods were combined, however, ADG was increased for implanted intact males. Estradiol implants at 9 wk of age did not affect performance of the cattle in our study.

Regardless of implant treatment, young intact males in our study exhibited better feed efficiency and more rapid gains during the growing period than during the finishing period (table 1). Growth of the calves might have been influenced by parity of the dams, i.e., some post-weaning compensatory growth may have occurred as a result of insufficient milk production from primiparous mothers. Also, differences in rate and efficiency of gain may be explained, in part, by changes in composition of gain during finishing.

Testicular weights and testosterone levels indicate that the implants were retained and were effective in altering normal testosterone secretion of the intact males (table 1). These results are consistent with previous research where scrotal circumference or testicular weight was studied (Lamm et al., 1980; Brethour, 1982; Greathouse et al., 1983; Unruh et al., 1984). Vanderwert et al. (1984), however, did not observe changes in scrotal circumference for intact males that were implanted with zeranol during the post-weaning period.

It has been suggested that the action of the estradiol implant is mediated through changes in luteinizing hormone secretion (D'Occhio et al., 1984), or by blocking gonadotropin release at the level of the hypothalamus (Schanbacher, 1984). The net effect of estradiol implantation is an increase in lipogenic activity, while lipolytic activity seems to remain unchanged (Prior et al., 1984).

Carcass Traits. Factors influenced by sex (main effect) are presented in table 2. For these traits, significant interactions were not observed. Intact male calves had heavier (P<.05) carcass weights and less (P<.01) subcutaneous fat, which resulted in lower (P<.01) numerical yield grades. No differences (P>.05) were evident in skeletal maturity, whereas steer carcasses possessed slightly higher quality grades (P<.01). These results are consistent with numerous reports in the literature concerning the carcass characteristics of intact males and steers (Seideman et al., 1982). Gray et al. (1983) reported that intact males implanted
with zeranol from birth to slaughter were less masculine than non-implanted intact males.

Muscle color, firmness and texture were not influenced by sex class and/or implant treatment (data not shown). Similar results for lean color and texture were reported by Gray et al. (1983) when zeranol-implanted intact males (implanted from birth to slaughter) were compared with non-implanted intact males.

The effects of implants on carcass traits of intact males and steers are shown in Table 3. Non-implanted cattle had lighter (P<.05) carcasses, less (P<.05) subcutaneous fat and lower (P<.05) numerical yield grades than either implant treatment. This concurs with the data of Brethour (1982) and Newland et al. (1984).

Zeranol-implanted cattle had more (P< .05) external fat and had higher (P<.05) USDA quality grades than did the non-implanted and the estradiol-implanted cattle. Brethour (1982), Gray et al. (1984) and Vanderwert et al. (1984) have reported that implanting intact males with zeranol increases carcass fatness.

A study similar in design to the present study was reported by Johnson et al. (1984). Very few implant effects on carcass fatness were detected in their study, and it was concluded that producers should expect no improvement in USDA quality grade of implanted intact males. However, the cattle used in the study by Johnson et al. (1984) were implanted at initiation of the finishing period, and were

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**TABLE 3. EFFECTS OF IMPLANTS ACROSS SEX CLASSES ON CARCASS CHARACTERISTICS**

<table>
<thead>
<tr>
<th>Trait</th>
<th>Control</th>
<th>Zeranol</th>
<th>Estradiol</th>
<th>SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yield grade</td>
<td>2.85d</td>
<td>3.57c</td>
<td>3.43c</td>
<td>.07</td>
</tr>
<tr>
<td>Carcass wt, kg</td>
<td>283d</td>
<td>303c</td>
<td>301c</td>
<td>2.93</td>
</tr>
<tr>
<td>Adjusted fat thickness, cm</td>
<td>1.21e</td>
<td>1.70c</td>
<td>1.49d</td>
<td>.05</td>
</tr>
<tr>
<td>Quality gradea</td>
<td>593d</td>
<td>607c</td>
<td>598d</td>
<td>5.2</td>
</tr>
<tr>
<td>Maturityb</td>
<td>146c</td>
<td>151d</td>
<td>155d</td>
<td>1.0</td>
</tr>
</tbody>
</table>

---

**TABLE 2. EFFECTS OF SEX CLASS ACROSS IMPLANT TREATMENTS ON CARCASS CHARACTERISTICS**

<table>
<thead>
<tr>
<th>Trait</th>
<th>Mean</th>
<th>SEa</th>
<th>Mean</th>
<th>SEa</th>
</tr>
</thead>
<tbody>
<tr>
<td>% Fat, carcassb</td>
<td>31.918</td>
<td>.43</td>
<td>38.84h</td>
<td>.57</td>
</tr>
<tr>
<td>Yield grade</td>
<td>3.08f</td>
<td>.08</td>
<td>3.77h</td>
<td>.11</td>
</tr>
<tr>
<td>Carcass wt, kg</td>
<td>299e</td>
<td>3.36</td>
<td>283f</td>
<td>5.39</td>
</tr>
<tr>
<td>Adjusted fat thickness, cm</td>
<td>1.34g</td>
<td>.02</td>
<td>1.75h</td>
<td>.03</td>
</tr>
<tr>
<td>Quality gradec</td>
<td>5968f</td>
<td>6.1</td>
<td>609h</td>
<td>8.1</td>
</tr>
<tr>
<td>Maturityd</td>
<td>151</td>
<td>1.1</td>
<td>148</td>
<td>2.6</td>
</tr>
</tbody>
</table>

---

*SE = standard error of the mean.

bPredicted as percentage of ether-extractable fat of the edible portion of the dressed carcass by percentage of ether-extractable fat in the ribeye.

c500 to 599 is Gd9 to Gd99 and 600 to 699 is Ch9 to Ch99, respectively.

d100 to 199 is A9 to A99, respectively.

e,fMeans in the same row that do not have a common superscript are different (P<.05).

The effects of implants on carcass traits of intact males and steers are shown in Table 3. Non-implanted cattle had lighter (P<.05) carcasses, less (P<.05) subcutaneous fat and lower (P<.05) numerical yield grades than either implant treatment. This concurs with the data of Brethour (1982) and Newland et al. (1984).
TABLE 4. INTERACTION MEANS FOR THE EFFECTS OF SEX CLASS AND IMPLANT TREATMENT ON CARCASS CHARACTERISTICS

<table>
<thead>
<tr>
<th>Trait</th>
<th>Control</th>
<th>Implant treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Intact males</td>
<td>Steers</td>
</tr>
<tr>
<td>% Fat, rib section</td>
<td>34.86\textsuperscript{de}</td>
<td>43.48</td>
</tr>
<tr>
<td>% Fat, carcass\textsuperscript{a}</td>
<td>29.29\textsuperscript{de}</td>
<td>35.66</td>
</tr>
<tr>
<td>Juiciness\textsuperscript{b}</td>
<td>8.29\textsuperscript{ce}</td>
<td>9.95</td>
</tr>
<tr>
<td>Muscle fiber tenderness\textsuperscript{b}</td>
<td>9.35\textsuperscript{c}</td>
<td>10.45\textsuperscript{x}</td>
</tr>
<tr>
<td>Warner-Bratzler shear force, kg</td>
<td>2.31</td>
<td>2.16</td>
</tr>
<tr>
<td>Connective tissue amount\textsuperscript{b}</td>
<td>11.28</td>
<td>11.88</td>
</tr>
<tr>
<td>Total collagen, mg/g of tissue</td>
<td>9.56</td>
<td>8.12</td>
</tr>
<tr>
<td>Collagen, % soluble</td>
<td>41.89</td>
<td>37.84</td>
</tr>
</tbody>
</table>

\textsuperscript{a}Predicted as percentage of ether-extractable fat of the edible portion of the dressed carcass by percentage ether-extractable fat in the 9-10-11th rib section.

\textsuperscript{b}Taste panel rating (1 = tough, abundant connective tissue, or dry; 15 = tender, no connective tissue, or juicy).

\textsuperscript{c,d}Means for intact males within the same row that do not have a common superscript are different (P<.05).

\textsuperscript{e}Intact males vs steers within implant treatment (P<.05).

\textsuperscript{x,y}Means for steers within the same row that do not have a common superscript are different (P<.05).
not subjected to lifelong implantation. The age of the animal at the time of implanting and the length of time that the body is exposed to the exogenous hormone seem to play a critical role in determining the extent to which implant effects will be manifested.

Palatability and Connective Tissue. The sex-implant interactions are presented in table 4. Measurements of fatness, either of the 9-10th rib-section, or predicted for the edible carcass portion, indicate that zeranol-implanted intact males had greater lipid (P<.05) deposition than estradiol-implanted or control intact males. There were no differences among steer groups for measures of fatness. Zeranol-implanted intact males were equal in fatness to zeranol-implanted steers. Estradiol-implanted or non-implanted intact males were leaner (P<.05) than steers.

Juiciness tended to reflect differences in fatness (table 4). Zeranol-implanted intact males provided steaks that were juicier (P<.05) than the leaner steaks from non-implanted or estradiol-implanted intact males. As with fatness, no differences (P>.05) in juiciness were observed among steer groups.

Implantation treatment had very little effect on muscle fiber tenderness ratings (table 4). Steaks from zeranol-implanted steers were found to be less tender (P<.05) than steaks from control steers. Direct comparisons between intact males and steers were not different (P>.05). When tenderness was assessed using the Warner-Bratzler shear, no differences were observed for any of the comparisons. These data indicate that the accelerated production system used in our study was sufficient to ensure adequate tenderness.

Connective tissue was evaluated by the taste panel (table 4) and no differences (P>.05) were found. In addition, laboratory determination of total and percentage of soluble collagen also failed to detect differences among treatments.

Conclusions

Lifelong implantation of intact males with zeranol or estradiol did not alter gain or feed efficiency when the growing and finishing phases were combined. Yield grades of intact males and steers were numerically increased by implantation with zeranol or estradiol. However, only zeranol implantation significantly increased fatness of the rib-section (or estimated fatness of the edible carcass portion) for intact males to the level of zeranol-implanted steers.

Juiciness ratings tended to reflect differences in fatness. No detrimental effects on tenderness or connective tissue were observed.

These results and those of Greathouse et al. (1983) and Gray et al. (1983) show that zeranol implantation of young intact males improves traits often cited by meat packers and processors as reasons for discriminating against young intact males. Masculinity was decreased (Gray et al., 1983) and quality grade, fat thickness and longissimus sensory-panel juiciness were increased. From a production standpoint, performance was at least equal, and carcass weight was increased, in these medium-framed intact males. Although zeranol- and estradiol-17β-implanted intact males had more fat than non-implanted intact males, this disadvantage could be alleviated by slaughtering at lighter weights.

Literature Cited


355.


