Distribution of *Abacion texense* (Loomis, 1937), the only millipede species traversing the Rio Grande, Mississippi, and Pecos rivers
(Callipodida: Abacionidae)

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**Abstract.** Localities are documented for the milliped *Abacion texense* (Loomis, 1837) (Callipodida: Abacionidae) whose distribution forms both the northern and southern ordinal limits in the Western Hemisphere. The westernmost component of *Abacion* Rafinesque, 1820, *A. texense* is the only milliped species whose range spans the Mississippi and Pecos rivers and the Rio Grande. Distribution extremes are in Hennepin County (Co.), Minnesota, in the north; Terrell and Potter cos., Texas, in the west; Alcorn Co., Mississippi, in the east; and southwestern Tamaulipas, Mexico, in the south. Occurrences are projected for southeastern South Dakota, northwestern Alabama, and the southwestern periphery of Tennessee. The type series of *A. texense* consists solely of the male holotype, so a neotype will be needed if this individual is ever lost, because no paratypes were officially designated.

**Introduction**

Seventeen Atlantic Coastal millipede genera in the United States (US) range subcontinuously westward across the Mississippi River. Eight continue into prairie ecosystems of the Central Plains, where forested habitats become sparse to nonexistent. Distributions tend to be broader in the south, so western boundaries angle southwestward from north to south as evidenced by *Narceus* Rafinesque, 1820 (Spirobolida: Spirobolidae), *Virgoiulus* Enghoff, 1984 (Julida: Blaniulidae), and *Eurymerodesmus* Brölemann, 1900 (Polydesmida: Eurymerodesmidae) (Shelley 1990, McAllister et al. 2005, Shelley et al. 2006). Only three genera, however, range far enough westward to also traverse the Pecos River, whose confluence with the Rio Grande, in west Texas, is some 1,056 km (660 mi) west of the Mississippi: *Eurymerodesmus*, *Cleidogona* Cook, 1895 (Chordeumatida: Cleidogonidae), and *Abacion* Rafinesque, 1820 (Callipodida: Abacionidae). The last two join *Rhysodesmus* Cook, 1895 (Polydesmida: Xystodesmidae), as the only genera also occurring east of the Mississippi and south of the Rio Grande (Shear 1972; Hoffman 1998, 1999; Shelley 1999), and if *Abacion* is ever discovered in Essex County, Ontario, across the Detroit River from Michigan, it will join *Cleidogona* as the only milliped genus occurring in Canada, the US, and Mexico (Shelley 1988).

While *Abacion* and *Cleidogona* occur east of the Mississippi, west of the Pecos, and south of the Rio Grande, the only species with this distribution is *A. texense* (Loomis, 1937). It forms the northern ordinal range limit in both North America and the Western Hemisphere and also forms the southern along with *Aspidiophon divisum* Shelley, 2000 (Schizopetalidae) in Sinaloa (Shelley 2000a, Stoev and Shelley 2009). The lone familial representative in Mexico, *A. texense* was originally assigned to *Spirostrephon* Brandt, 1841, but Chamberlin and Hoffman (1958) transferred it into *Abacion*, a senior synonym. Published and unpublished localities coupled with known ranges indicate that early records of “*Lysioptetalum lactarium*” (Say, 1821) from Kansas, Minnesota, Nebraska, and Oklahoma (Bollman 1893; Kenyon 1893a, b; Gunthorp 1913, 1921; Chamberlin 1931) actually refer to *A. texense*. Shelley (1984) reviewed *Abacion* and mapped distributions of its then four component species, but for brevity did not provide counties or localities. Shelley et al. (2003) described a fifth species that is endemic to Rich Mountain, Polk Co., Arkansas, and regional works have provided localities for others (Williams and Hefner 1928; Johnson 1954; Shelley
distribution, we address A. texense specifically, providing a diagnostic account and full synonymy as
Shelley (1984) missed a few references. Locality details are provided for states in which this millipede
is known from five or fewer counties, and the latter alone are given for states with six or more occupied
counties. We project ranges in all states of occurrence and cite extremes for ones on borders; missing data
were not provided on vial labels. Anatomical details and diagnostic illustrations are available in Shelley
(1984: 986-987, figs. 10-11). Repository codens, all in the US, are as follows:

FMNH Field Museum of Natural History, Chicago, Illinois.
FSCA Florida State Collection of Arthropods, Gainesville, Florida.
INHS Illinois Natural History Survey, Champaign, Illinois.
MCZN Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts.
MLBM Monte L. Bean Life Science Museum, Brigham Young University, Provo, Utah.
NCSM North Carolina State Museum of Natural Sciences, Raleigh, North Carolina.
NMNH National Museum of Natural History, Smithsonian Institution, Washington, DC.
OSEC Emerson Entomology Museum, Oklahoma State University, Stillwater, Oklahoma.
TMMC Texas Memorial Museum, Austin, Texas.
UCMC University of Colorado Museum of Natural History, Boulder, Colorado.
UGA University of Georgia Museum of Natural History, Athens, Georgia.
UMSP - Entomology Department, University of Minnesota, St. Paul, Minnesota.
VMNH Virginia Museum of Natural History, Martinsville, Virginia.
WASC Private collection of W. A. Shear, Hampden-Sydney, Virginia.
WTAM Department of Life, Earth, and Environmental Sciences, West Texas A&M University, Canyon,
Texas.

Abacion texense (Loomis, 1937)

Spirostrephon texense: Chamberlin 1942a:16, fig. 28.
Spirostrephon jonesi Chamberlin 1942b:17.
Tynomma messicanum Chamberlin 1943a:33. Loomis 1968a:70.


Type specimen. Male holotype (NMNH) collected by O. F. Cook in December 1905 at Pierce, Wharton
Co., Texas; no paratypes were officially designated.

Diagnosis (adapted from Shelley, 1984). Gonopodal postfemur short, divided proximad; branch “A”
subquadrate, much shorter than branch “B”; solenomere subspiniform, shorter than and poorly demar-
cated from branch “B”; tibiotarsus with sides narrowing to subacuminate tip, reflexed apically.

Habitats. Habitats include beneath limestone rocks, decaying pine logs and bark, rotting oak logs,
railroad ties, and in mixed deciduous litter; elevations range from sea level to 1,620 m (5,400 ft). In Texas,
A. texense is commonly encountered in cave, sinkhole, and spring habitats; the Terrell Co. individual
was under a decaying Ashe juniper (Juniperus ashei J. Buchholz) log on desert soil and limestone. In addition
to juniper, dominant vegetation included prickly-pear cactus (Opuntia sp.), Vasey oak (Quercus pungens
Liebm.), Torrey yucca (Yucca torreyi Shafer), and mesquite (Prosopis glandulosa Torrey).

Distribution (Fig. 1). We modify the latest range statement (Stoev and Shelley 2009) as follows:
“Occurring in every US state from southern Minnesota and eastern Nebraska southward to Louisiana.
and Texas, continuing southward across the Rio Grande into northeastern Coahuila, northern Nuevo León, and most of Tamaulipas, Mexico. The distribution extends across the Mississippi River into Mississippi and southeastern Louisiana, spreads westward in Texas to the panhandle, and crosses the Pecos River near its confluence with the Rio Grande. *Abacion texense* forms the northern, western, and southern boundaries for both the genus and family, so in addition to Shelley’s (1984) spot map, borders of its range are depicted in ordinal and familial maps (Shelley 1989:382, fig. 1; Shear et al. 2003:3, fig. 1; Stoev et al. 2008:3, fig. 2). Stoev and Shelley (2009:164, fig. 4) show that *A. texense* is the lone ordinal component in Coahuila, Nuevo León, and Tamaulipas.

Even larger ranges have been projected. Shelley (1984) suggested occurrence in central Nebraska, southeastern South Dakota, and the Black Hills of southwestern South Dakota, but we have sampled in the last without encountering callipodidans and believe that *A. texense* does not occur there. From 2006 to 2008 CTM sampled in northwestern, northcentral, and northeastern Nebraska and southeastern South Dakota without finding this millipede, which we now believe does not inhabit the first two regions. Occupation of the last two is plausible because of their moister, verdant, riparian habitats and locations between the Minnesota and Nebraska sites. Shelley (1984) also suggested occurrence farther west in Texas, in the Davis Mountains, Jeff Davis Co., but we have also investigated this inselberg and worked westward to El Paso without finding a callipodid. Consequently, we believe that the Terrell Co. locality is at or very near the true western boundary of the species, genus, and family. To the east, Hoffman (1999) and Stoev et al. (2008) predicted discovery in Alabama and Tennessee, and although no-one has yet searched these areas, the northeastern Mississippi locality, in Alcorn Co., implies occurrence in the northwestern corner of Alabama and the southwestern periphery of Tennessee.

**Published range statements.** “Great Plains from Ames, Iowa, and Lincoln, Nebraska, through Kansas, Oklahoma, and western Missouri, south as far as Kerr, Bandera, and Wharton Counties, Texas, east through Arkansas and Louisiana to Rankin County, Mississippi” (Chamberlin and Hoffman 1958). On the advice of N.B. Causey, Loomis (1968a) expanded the area to include Monterrey and Bustamente, Nuevo León, Mexico, and Shelley (1984) added southern Minnesota and Coahuila and Tamaulipas. “Great Plains from Minnesota and eastern Nebraska south to northern Nuevo León, eastward as far as southern Mississippi, doubtlessly also in Alabama and Tennessee” (Hoffman 1999, Stoev et al. 2008).


MINNESOTA: Minnesota in general (Stoev et al. 2008, Stoev and Shelley 2009).


MISSOURI: Western Missouri (Chamberlin and Hoffman 1958, Loomis 1968a).


**MEXICO:** COAHUILA: northeastern Coahuila (Stoev and Shelley 2009). El Nacimiento, and 2 km (1.6 mi) S and 10 km (6.3 mi) SE Musquiz (Stoev and Shelley 2009).


**TAMAULIPAS:** Municipio de Ocampo, 18 km (11.3 mi) NW Chamal (Stoev and Shelley 2009).


**LOUISIANA:** Statewide. Acadia, Allen, Ascension, Beauregard, Bienville, Bossier, Caddo, Caldwell, Catahoula, Concordia, Evangeline, Franklin, Jefferson, Iberia, Landry, LaSalle, Lincoln, Madison, Natchitoches, Orleans, Ouachita, Rapides, St. Helena, St. Landry, St. Tammany, Tensas, Vernon, West Feliciana, and Winn pars. (FSCA, MCZN, NCSM, NMNH, VMNH).


Canyon, 24.1 km (15.1 mi) E Old Canyon, F, 28 April 1962, R.O. Albert (FSCA) *First record for the order, family, genus, and species from the Texas Panhandle*. Terrell Co., Independence Creek Preserve, 37 km (23.1 mi) SE Sheffield, M, 7 October 2005, C.T. McAllister (NCSM) *Westernmost record and first from west of the Pecos River for the family, genus, and species*.

**MEXICO:** COAHUILA: northern periphery and northeast corner; no new records.

NUEVO LEÓN: northern half and possibly farther south in the eastern periphery; no new records.

TAMAULIPAS: northern 3/4th; no new records. The published locality is the southernmost for the species, genus, and family, and one of the two southernmost for the order in the Western Hemisphere.

**Remarks.** In the original description, Loomis (1937) indicated, but did not actually state, that the male from Pierce, Wharton Co., Texas, is the holotype; no paratypes were mentioned, suggesting that the type series consisted of only this one individual. In subsequent remarks Loomis stated, “…many other specimens were collected the same year (1905) at Wharton, Texas, on the bank of the Colorado River,” and he also mentioned samples collected in 1927 in Dallas, Smith, and Tarrant cos. All this material was apparently available when Loomis described the species, but we do not know the number of individuals. “Many other specimens” is vague, and numbers were not provided for the 1927 samples. The FSCA houses two males and two females from the Colorado River bank sample that are labeled, “paratypes”; presumably, they constitute only part of this sample because “many specimens” implies more to us. As these specimens were not specifically designated as paratypes in the original account, they are not such despite the sample label (Article 72.4.6 of the Code, ICZN 1999); if they were legitimate paratypes, the 1927 samples from Dallas, Smith, and Tarrant cos. would also hold this status. This technicality is significant because if the holotype is ever lost, a neotype will have to be designated among new or existing material because no official paratypes exist.

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Literature Cited


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