June 2007

Caffeine Formulation for Avian Repellency

Scott J. Werner
USDA-APHIS-Wildlife Services, Scott.J.Werner@aphis.usda.gov

John L. Cummings
United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center

Shelagh K. Tupper
United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center

Jerome C. Hurley
United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center

Randal S. Stahl
USDA-APHIS-Wildlife Services, randal.s.stahl@aphis.usda.gov

See next page for additional authors

Follow this and additional works at: https://digitalcommons.unl.edu/icwdm_usdanwrc

Part of the Environmental Sciences Commons

https://digitalcommons.unl.edu/icwdm_usdanwrc/727

This Article is brought to you for free and open access by the U.S. Department of Agriculture: Animal and Plant Health Inspection Service at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in USDA National Wildlife Research Center - Staff Publications by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.
Caffeine Formulation for Avian Repellency

SCOTT J. WERNER,1 United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA
JOHN L. CUMMINGS, United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA
SHELAGH K. TUPPER, United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA
JEROME C. HURLEY, United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA
RANDAL S. STAHL, United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA
THOMAS M. PRIMUS, United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, 4101 LaPorte Avenue, Fort Collins, CO 80521-2154, USA

ABSTRACT Nonlethal management alternatives are needed to reduce avian depredation of agricultural crops. Caffeine has promise as an effective, economical, and environmentally safe avian repellent, yet formulation improvements are needed for field applications. We included sodium benzoate in subsequent formulations to enhance the solubility of caffeine. Red-winged blackbirds (Agelaius phoeniceus) discriminated between untreated rice and rice treated with 250 ppm or 10,000 ppm caffeine and sodium benzoate in captivity. We observed a positive concentration–response relationship among birds offered rice treated with 250 ppm, 1,000 ppm, 2,500 ppm, 5,000 ppm, 10,000 ppm, or 20,000 ppm caffeine and sodium benzoate. Relative to the pretreatment of the concentration–response test, repellency was >85% for rice treated with our 2,500- to 20,000-ppm treatments. We conducted seed germination experiments and associated residue analyses to evaluate the potential phytotoxicity of various caffeine formulations under controlled environmental conditions. The optimized formulation enhanced the solubility of tank mixtures and ameliorated the negative impacts of seed treatments to germination. Our caffeine formulation provides an applicable, nonlethal alternative for managing blackbird impacts to agricultural production. (JOURNAL OF WILDLIFE MANAGEMENT 71(5):1676–1681; 2007)

DOI: 10.2193/2006-451

KEY WORDS Agelaius phoeniceus, chemical repellent, depredation, foraging behavior, red-winged blackbird, sodium benzoate, wildlife damage management.

Several bird species cause monetary losses to agricultural production throughout the United States. For example, red-winged blackbirds (Agelaius phoeniceus), common grackles (Quiscalus quiscula), and brown-headed cowbirds (Molothrus ater) caused approximately US$13.4 million of damage to United States rice production in 2001 (Cummings et al. 2005). Efforts to manage such damages include the use of repellents and other nonlethal management alternatives (Werner et al. 2005). Chemical repellents can be used as seed treatments to reduce blackbird impacts to newly planted rice or as aerial applications to reduce blackbird consumption of ripening rice.

Schafer et al. (1983) identified caffeine as a potential avian repellent with relatively low toxicity (LD₅₀ = 316 mg/kg for European starlings [Sturnus vulgaris]). Avery and Cummings (2003) found that 2,500 ppm caffeine reduced rice consumption by 76% among male red-winged blackbirds in captivity. A rice seed treatment of 2,500 ppm caffeine would cost approximately $3/ha (at $8.10/kg; Flavine North America, Inc., Closter, NJ). Blackbirds consumed <10% of rice seeds treated with 10,000 ppm caffeine and >80% of untreated rice seeds under field conditions in southwestern Louisiana, USA (Avery et al. 2005). Thus, caffeine appeared to be an effective, economical, and environmentally safe avian repellent (Avery et al. 2005). However, formulation improvements were needed to make caffeine practical for agricultural applications. For example, the low solubility of caffeine in water impeded its mixture during our previous field evaluation (Avery et al. 2005).

We conducted 3 feeding experiments and 2 rice germination trials to develop an optimized formulation of caffeine as an avian repellent for agricultural applications. We included sodium benzoate in our seed treatments to obtain the requisite solubility of caffeine for subsequent field applications. The solubility of caffeine, and caffeine and sodium benzoate combined is approximately 1.5 g and 83 g per 100 mL water, respectively (Gemmill 1958). Our objectives were to 1) improve the solubility of caffeine for subsequent field applications, 2) evaluate avian repellency of caffeine and sodium benzoate in captivity, 3) determine the mode of action of caffeine and sodium benzoate as an avian repellent, and 4) optimize caffeine residues and germination of our seed treatments.

STUDY AREA

We conducted blackbird feeding experiments and rice germination trials at the United States Department of Agriculture’s National Wildlife Research Center (NWRC) in Fort Collins, Colorado, USA. We conducted preference and concentration–response tests within an open-sided building at the NWRC outdoor animal research facility.

1 E-mail: Scott.J.Werner@aphis.usda.gov
The NWRC Analytical Chemistry unit performed caffeine residue analyses associated with rice germination trials.

**METHODS**

**Preference Testing in Captivity**

We conducted 2 preference tests to determine if red-winged blackbirds could discriminate between untreated rice and rice treated with 250 ppm (test 1) or 10,000 ppm (test 2) caffeine and sodium benzoate. We used a cannon-net (Dill and Thornberry 1950) to capture 26 adult red-winged blackbirds (M) in November 2004 near Fort Collins, Colorado. We transported all birds to NWRC within 1 hour of capture. We transferred birds to individual cages following group quarantine and holding, and offered them untreated seed rice (ad libitum) in each of 2 food bowls for 3–5 days of acclimation. We provided water ad libitum to all birds throughout preference testing (i.e., acclimation, pretreatment, test).

We offered each bird 2 bowls of untreated rice (30 g each) during each day of the 4-day pretreatment. We collected unconsumed rice (remaining in food bowls) and rice spillage (remaining in trays beneath each bowl) at 0800–0930 hours daily and determined their mass (±0.1 g). We measured pretreatment and test consumption independently for food bowls located on the north and south sides of each cage. We accounted for changes in the mass of rice independent of rice consumption (e.g., desiccation) by weighing rice offered within a vacant cage throughout testing. We offered each bird 2 bowls of untreated rice (ad libitum) for 3 days between the pretreatment and test.

We offered each bird one bowl of untreated rice and one bowl of rice treated with 250 ppm (n = 6 birds) or 10,000 ppm (n = 20 birds) caffeine and sodium benzoate (1:1) during each day of the 4-day test. We applied treatment solutions (60 mL/kg rice) to 10 kg certified seed rice (Louisiana State University Rice Research Station, Crowley) using a rotating mixer and household spray equipment. We included 6 mL Transfilm® (PBI/Gordon, Kansas City, MO) to each treatment solution. We randomized the north–south positioning of treatments within individual cages on the first day and alternated positioning on subsequent days of the test. We offered untreated rice (ad libitum) in 2 bowls for 3 days following the test.

We measured posttest rice consumption during our evaluation of 10,000 ppm caffeine and sodium benzoate to investigate the mode of action of our caffeine formulation. Birds avoid and discontinue sampling food previously associated with secondary repellents (Werner and Clark 2003). If caffeine plus sodium benzoate acted as a postigestive toxin, or secondary repellent, we predicted that posttest consumption of untreated rice would be less than overall pretreatment (i.e., baseline) consumption of untreated rice. We ranked blackbirds based upon average pretreatment consumption and assigned them to 1 of 6 treatment groups. We randomly assigned posttest treatments to both groups. We provided 2 bowls of rice treated with 10,000 ppm caffeine and sodium benzoate to birds in group A (n = 10 birds) and 2 bowls of untreated rice to birds in group B (n = 10) during each day of the 4-day posttest.

The dependent measure for preference testing was average rice consumption during the pretreatment, test, and posttest periods. We used the general linear model procedure (SAS Institute, Cary, NC) to examine the main effect of testing period (error = period × subject), and the period–treatment (error = period × treatment × subject) and period–day–treatment (residual error) interactions during the pretreatment and test. We separated means of analysis of variance effects (α = 0.05) using Tukey’s tests. We used descriptive statistics (x ± SE) to summarize rice consumption throughout preference testing. The NWRC Institutional Animal Care and Use Committee approved the capture, care, and use of animals associated with our feeding experiments (Study Protocol QA1127).

**Concentration–Response Testing in Captivity**

We conducted a concentration–response test to evaluate repellency associated with varied concentrations of caffeine and sodium benzoate. We captured 48 adult red-winged blackbirds (M) in November 2004 near Fort Collins, Colorado, and transported them to NWRC. We repeated all quarantine, holding, acclimation, and pretreatment procedures (using one food bowl) used during preference testing. We measured daily consumption of untreated rice throughout the 4-day pretreatment.

We ranked blackbirds based upon average pretreatment consumption and assigned them to 1 of 6 treatment groups (n = 8 birds/group). We randomly assigned treatments among groups. Treatment groups were used to evaluate repellency associated with 250 ppm, 1,000 ppm, 2,500 ppm, 5,000 ppm, 10,000 ppm, and 20,000 ppm caffeine and sodium benzoate (1:1). We applied treatment solutions (60 mL/kg rice), including 6 mL Transfilm, to 10 kg certified seed rice. We measured consumption of treated rice offered in one bowl during each day of the 4-day test.

We hypothesized that repellency would be directly related to the concentration of our caffeine formulation. We predicted that test consumption associated with efficacious treatments would be <75% (Schneider 1982) of pretreatment consumption. We analyzed percent repellency (i.e., test day 1 relative to average pretreatment consumption) as a function of treatment concentration using linear regression. We used descriptive statistics (x ± SE) to summarize repellency among treatment groups.

**Germination Testing of Rice Seed Treatments**

We conducted 2 controlled trials to determine the effects of caffeine, sodium benzoate, and Transfilm on seed germination. We sealed 450 g of certified Cocodrie seed rice within each of 6 perforated Ziploc® (S.C. Johnson & Son, Inc., Racine, WI) bags to initiate trial 1. We submerged all bags in water for 36 hours. We placed soaked seeds in a cotton bag for 5 hours. We subsequently applied treatment formulations (Table 1) and replaced all seeds within a treatment group in a cotton bag for 24 hours. We then placed 20 treated seeds per treatment in each of 2 petri
Table 1. Caffeine formulations and associated germination of rice seed treatments that were tested for effectiveness as an avian repellent at the National Wildlife Research Center in Fort Collins, Colorado, USA, April 2005–July 2006.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Caffeine (g)</th>
<th>Sodium benzoate (g)</th>
<th>Transfilm (mL)</th>
<th>Water (mL)</th>
<th>Germination (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trial 1 (450 g rice)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.0</td>
<td>0.0</td>
<td>0</td>
<td>0</td>
<td>15</td>
<td>90</td>
</tr>
<tr>
<td>4.5</td>
<td>0.0</td>
<td>0</td>
<td>0</td>
<td>15</td>
<td>0</td>
</tr>
<tr>
<td>0.0</td>
<td>4.5</td>
<td>0</td>
<td>1</td>
<td>15</td>
<td>88</td>
</tr>
<tr>
<td>4.5</td>
<td>4.5</td>
<td>1</td>
<td>1</td>
<td>15</td>
<td>0</td>
</tr>
<tr>
<td>4.5</td>
<td>4.5</td>
<td>1</td>
<td>2</td>
<td>20</td>
<td>43</td>
</tr>
<tr>
<td>Trial 2 (3 kg rice)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.0</td>
<td>0.0</td>
<td>0</td>
<td>0</td>
<td>200</td>
<td>91</td>
</tr>
<tr>
<td>5.6</td>
<td>4.2</td>
<td>2</td>
<td>2</td>
<td>200</td>
<td>94</td>
</tr>
<tr>
<td>6.7</td>
<td>3.3</td>
<td>2</td>
<td>2</td>
<td>200</td>
<td>91</td>
</tr>
<tr>
<td>7.5</td>
<td>2.8</td>
<td>2</td>
<td>2</td>
<td>200</td>
<td>91</td>
</tr>
<tr>
<td>8.0</td>
<td>2.4</td>
<td>2</td>
<td>2</td>
<td>200</td>
<td>91</td>
</tr>
<tr>
<td>8.5</td>
<td>4.8</td>
<td>2</td>
<td>2</td>
<td>233</td>
<td>96</td>
</tr>
<tr>
<td>7.8</td>
<td>3.9</td>
<td>2</td>
<td>2</td>
<td>233</td>
<td>96</td>
</tr>
<tr>
<td>8.7</td>
<td>3.2</td>
<td>2</td>
<td>2</td>
<td>233</td>
<td>96</td>
</tr>
<tr>
<td>9.3</td>
<td>2.8</td>
<td>2</td>
<td>2</td>
<td>233</td>
<td>96</td>
</tr>
<tr>
<td>9.0</td>
<td>4.4</td>
<td>3</td>
<td>3</td>
<td>267</td>
<td>94</td>
</tr>
<tr>
<td>9.0</td>
<td>3.7</td>
<td>3</td>
<td>3</td>
<td>267</td>
<td>95</td>
</tr>
<tr>
<td>10.0</td>
<td>3.2</td>
<td>3</td>
<td>3</td>
<td>267</td>
<td>90</td>
</tr>
</tbody>
</table>

dishes (9.5-cm diam) lined with a hydrated (3.5 mL deionized water) Kimwipe® (Kimberly-Clark Global Sales, Inc., Roswell, GA). We sealed all dishes with Parafilm® (Pechiney Plastic Packaging, Chicago, IL) and placed them in indirect sunlight for 1 hour. We subsequently placed all dishes in an enclosed dark space (range = 21–23°C) and monitored seed germination each 24 hours. We tallied sprouts ≥5 mm in each dish at the conclusion of trial 1.

We conducted a second trial to optimize caffeine residues and germination of our seed treatments. We placed 3 kg of certified Cocodrie seed rice within each of 14 cotton bags. We submerged all bags in water for 36 hours, then drained seeds within bags for 5 hours. We subsequently applied treatment formulations using household spray equipment and a Hobart® mixer (Hobart Manufacturing Company, Troy, OH).

We varied the water content and the molar ratio of caffeine and sodium benzoate in trial 2 (Table 1). We varied the water contribution by 67 mL, 78 mL, and 89 mL of deionized water/kg rice. We included 1% Transfilm in each formulation. We placed 20 seeds per treatment in each of 4 petri dishes (9.5-cm diam) lined with a hydrated (3.5 mL deionized water) Kimwipe. We subsequently sealed all 4 petri dishes (9.5-cm diam) lined with a hydrated (3.5 mL deionized water) Kimwipe. We tallied sprouts ≥5 mm in each dish at the conclusion of trial 2 (USEPA 1996).

We also evaluated the purity of caffeine and sodium benzoate used for formulation treatments using HPLC. We used descriptive statistics to summarize rice germination (x ± SE; trials 1 and 2) and residue data (x ppm [dry]; trial 2).

RESULTS

Preference Testing in Captivity

Red-winged blackbirds discriminated between untreated rice and rice treated with 250 ppm caffeine and sodium benzoate during the preference test (Fig. 1A). Although we observed no difference in rice consumption between bowls during the pretreatment (Tukey P > 0.05), all birds consumed less rice treated with 250 ppm caffeine and sodium benzoate than untreated rice (F2,10 = 39.78, P < 0.001). On average, birds consumed 10.2 (±0.5) g/bird/day of untreated rice and 1.3 (±0.5) g/bird/day of treated rice during the test (Tukey P < 0.05). We observed no difference in overall rice consumption (i.e., in both bowls) between the pretreatment and test (F1,10 = 0.00, P = 0.984). We also observed no period–day–treatment interaction (F12,60 = 0.19, P = 0.998). Thus, blackbirds strongly preferred untreated rice when offered a choice during the 250-ppm caffeine and sodium benzoate test.

Blackbirds also discriminated between untreated rice and rice treated with 10,000 ppm caffeine and sodium benzoate (Fig. 1B). Although we observed no difference in rice consumption between bowls during the pretreatment (Tukey P > 0.05), blackbirds consumed less treated rice than untreated rice during the test (F2,36 = 465.61, P < 0.001). On average, birds consumed 11.2 (±0.5) g/bird/day of untreated rice and 0.0 (±0.0) g/bird/day of treated rice during the test (Tukey P < 0.05). We observed no difference in overall rice consumption (i.e., in both bowls) between the pretreatment and test periods (F1,36 = 0.19, P =
We also observed no period–day–treatment interaction ($F_{12,216} = 0.29$, $P = 0.990$).

All blackbirds continued to avoid rice treated with 10,000 ppm caffeine and sodium benzoate during the posttest (Fig. 1B). However, blackbirds did not avoid untreated rice subsequent to test exposure. On average, blackbirds consumed $0.0 \ (\pm 0.1)$ g/bird/day of treated rice and $11.1 \ (\pm 0.3)$ g/bird/day of untreated rice during the posttest.

**Concentration–Response Testing in Captivity**

Relative to pretreatment rice consumption, red-winged blackbirds consumed 52% and 88% less rice treated with 1,000 ppm and 2,500 ppm caffeine and sodium benzoate, respectively (Fig. 2). Blackbirds consumed $<1$ g of rice treated with 5,000 ppm, 10,000 ppm, or 20,000 ppm caffeine and sodium benzoate. Relative to pretreatment consumption, blackbirds exhibited 100% repellency for rice treated with $\geq 5,000$ ppm caffeine and sodium benzoate. We observed a direct concentration–response relationship among tested concentrations of our caffeine formulation ($r^2 = 0.374$, $P < 0.001$). Thus, rice consumption was inversely related to caffeine and sodium benzoate concentration subsequent to the pretreatment.

**Germination Testing of Rice Seed Treatments**

We observed 98.5% ($\pm 0.4\%$) and 97.9% ($\pm 0.5\%$) purity of caffeine and sodium benzoate used for treatment formulations, respectively. Approximately 90% of rice seeds germinated when treated only with water (control seed treatment), and with Transfilm and water (Table 1). We observed no germination among seed treatments of caffeine, sodium benzoate, and $<20$ mL water (Table 1). We thus varied the water contribution and molar ratio of caffeine and sodium benzoate among treatments in trial 2. The Louisiana Seed Commission (Louisiana Department of Agriculture and Forestry, Baton Rouge) certified that 80% germination was expected for the rice seed associated with trial 2. We observed $>85\%$ germination among all seed treatments in trial 2 (Table 1).

We observed $<33$ ppm caffeine (HPLC method limit of detection) among our control seed treatments (200 mL and 267 mL deionized water only) in trial 2. We recovered 1,310–2,115 ppm caffeine (1:1 and 2.5:1, respectively) among seeds in the 67-mL/kg treatment groups (Fig. 3). We observed 1,710–2,305 ppm caffeine (1:1 and 2.5:1) among seeds in the 78-mL/kg treatments and 1,585–2,535 ppm caffeine (1:1 and 2:1) among seeds in the 89-mL/kg treatments (Fig. 3). We observed maximum caffeine residues and rice germination with our 89-mL/kg and 2:1 caffeine to sodium benzoate formulation (Fig. 3).

**DISCUSSION**

Our formulation of caffeine, sodium benzoate, and Transfilm enhanced the solubility and limited the phytotoxicity of seed treatments. In contrast to other formulations that yielded $>2,000$ ppm caffeine residues (Fig. 3), we observed 95% germination in each dish associated with our 89-mL water/kg and 2:1 caffeine to sodium benzoate formulation.
repellent applications of our caffeine formulation. The data requirements for registration include field residue analyses, environmental fate, and acute and chronic toxicology associated with field applications (e.g., potential impacts to crawfish [Procambarus spp.] production). We plan to request an experimental use permit to evaluate our caffeine formulation under extended field conditions.

ACKNOWLEDGMENTS
We thank J. D. Eisemann, B. A. Kimball, G. M. Linz, and M. E. Tobin for their comments on an earlier version of this manuscript. We also thank the United States Department of Agriculture, Wildlife Services personnel in Louisiana (D. J. LeBlanc, E. A. Wilson) and Missouri, USA (R. W. Byrd) for dedicated assistance with this and ongoing repellents research. We appreciate the National Wildlife Research Center animal care staff who provided daily care throughout our feeding experiments. The Louisiana Rice Research Board, Louisiana State University Rice Research Station, Flavine North America, Inc., and USA Rice Foundation provided financial or logistical support for this study. Corporate collaborations do not imply endorsement by the United States Department of Agriculture.

LITERATURE CITED
Schaefer, E. W., W. A. Bowles, Jr., and J. Hurbut. 1983. The acute oral toxicity, repellency, and hazard potential of 998 chemicals to one or more species of wild and domestic birds. Archives of Environmental Contamination and Toxicology 12:355–382.


Associate Editor: Mason.