

2011

# Genetic modulation of horizontal cell number in the mouse retina

Irene Whitney  
*University of California, Santa Barbara,*

Mary Raven  
*University of California, Santa Barbara,*

Daniel Ciobanu  
*University of Nebraska, Lincoln, NE*

Ross Poché  
*Baylor College of Medicine, Houston, TX*

Qian Ding  
*University of Rochester, Rochester, NY*

*See next page for additional authors*

Follow this and additional works at: <http://digitalcommons.unl.edu/animalscifacpub>

 Part of the [Animal Sciences Commons](#)

---

Whitney, Irene; Raven, Mary; Ciobanu, Daniel; Poché, Ross; Ding, Qian; Elshatory, Yasser; Gan, Lin; Williams, Robert; and Reese, Benjamin, "Genetic modulation of horizontal cell number in the mouse retina" (2011). *Faculty Papers and Publications in Animal Science*. 757.

<http://digitalcommons.unl.edu/animalscifacpub/757>

This Article is brought to you for free and open access by the Animal Science Department at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Faculty Papers and Publications in Animal Science by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

---

**Authors**

Irene Whitney, Mary Raven, Daniel Ciobanu, Ross Poché, Qian Ding, Yasser Elshatory, Lin Gan, Robert Williams, and Benjamin Reese

# Genetic modulation of horizontal cell number in the mouse retina

Irene E. Whitney<sup>a,1</sup>, Mary A. Raven<sup>a,1</sup>, Daniel C. Ciobanu<sup>b</sup>, Ross A. Poché<sup>c</sup>, Qian Ding<sup>d</sup>, Yasser Elshatory<sup>d</sup>, Lin Gan<sup>d</sup>, Robert W. Williams<sup>e</sup>, and Benjamin E. Reese<sup>a,2</sup>

<sup>a</sup>Neuroscience Research Institute and Departments of Psychological and Brain Sciences and Molecular, Cellular and Developmental Biology, University of California, Santa Barbara, CA 93106; <sup>b</sup>Animal Science Department, University of Nebraska, Lincoln, NE 68583; <sup>c</sup>Department of Molecular Physiology and Biophysics, Baylor College of Medicine, Houston, TX 77030; <sup>d</sup>Department of Ophthalmology, University of Rochester, Rochester, NY 14642; and <sup>e</sup>Center of Genomics and Bioinformatics and Department of Anatomy and Neurobiology, University of Tennessee Health Science Center, Memphis TN 38120

Edited\* by John E. Dowling, Harvard University, Cambridge, MA, and approved April 26, 2011 (received for review February 28, 2011)

**Neuronal populations display conspicuous variability in their size among individuals, but the genetic sources of this variation are largely undefined. We demonstrate a large and highly heritable variation in neuron number within the mouse retina, affecting a critical population of interneurons, the horizontal cells. Variation in the size of this population maps to the distal end of chromosome (Chr) 13, a region homologous to human Chr 5q11.1–11.2. This region contains two genes known to modulate retinal cell number. Using conditional knock-out mice, we demonstrate that one of these genes, the LIM homeodomain gene *Islet-1* (*Isl1*), plays a role in regulating horizontal cell number. Genetic differences in *Isl1* expression are high during the period of horizontal cell production, and *cis*-regulation of *Isl1* expression within the retina is demonstrated directly. We identify a single nucleotide polymorphism in the 5' UTR of *Isl1* that creates an E-box sequence as a candidate causal variant contributing to this variation in horizontal cell number.**

allele-specific expression | follistatin | quantitative trait locus | recombinant inbred strain

The cellular architecture and connectivity of the vertebrate retina is largely conserved across species. What distinguishes these retinas most is their variation in size, in the numbers of each of the different cell types, and in their relative proportions. For instance, old world primates display a two- to threefold variation in the number of cone photoreceptors in the fovea (1, 2), and a twofold variation in the total number of retinal ganglion cells (3). Different strains of inbred mice have also been shown to differ in the size of their cone photoreceptor, amacrine, and ganglion cell populations (4–7). There are multiple potential causes of such variation in retinal nerve cell number, including degenerative changes associated with the accumulated effects of light damage or other processes that affect select types of neurons. In other cases, however, such variation is detected early during development (8), implicating a modulation of the very processes that determine nerve cell number from the outset.

The developmental basis for such variation is not well understood, although polymorphic genes that modulate cellular production and naturally occurring cell death are likely to be major factors. We have previously demonstrated in the mouse retina a nearly twofold variation in the total number of horizontal cells, being a type of retinal interneuron providing feedback inhibition to photoreceptors (9, 10). This large variation across mouse strains is present from the outset of retinal development (11) and is independent of variation in retinal area (9). Furthermore, these horizontal cells modulate their dendritic growth in relation to this variation in cellular density (12), indicating that the determinants of retinal nerve cell number also indirectly impact differentiation.

The present investigation has adopted a forward genetic approach to identify key determinants of this variation in retinal horizontal cell number. We demonstrate a strong heritable dif-

ference in horizontal cell number among 26 recombinant inbred (RI) strains of mice derived from reciprocal crosses between two inbred parental strains, A/J and C57BL/6J (hereafter B6/J). We mapped a quantitative trait locus (QTL) that modulates this neuronal population to the distal end of chromosome (Chr) 13. The QTL overlaps two candidate genes that have been shown to modulate numbers of retinal neurons. Using a conditional gene knock-out strategy, we demonstrate an important role for one of these genes, the LIM homeodomain transcription factor *Islet-1* (*Isl1*), in the suppression of this phenotype. *Isl1* expression is shown to differ between the parental strains during the period of horizontal cell neurogenesis, due to the presence of a *cis*-acting polymorphism. We identified such a potential variant, a single nucleotide polymorphism (SNP) creating a binding site for basic helix–loop–helix (bHLH) transcription factors in the B6/J strain.

## Results

**Mouse Retina Displays a Large and Highly Heritable Variation in Horizontal Cell Number.** Mice of the A/J strain contain  $9,884 \pm 168$  horizontal cells, whereas those of the B6/J strain contain  $18,471 \pm 288$  cells (mean and SE; Fig. 1, white and black bars). F<sub>1</sub> hybrids (B6AF1) have a value intermediate to the two parental strains,  $14,417 \pm 263$  cells (Fig. 1, light gray bar). Members of the AXB/BXA strain family have genomes that are a unique mix of A and B haplotypes across chromosomes. These progeny strains have numbers of horizontal cells that are distributed widely across the range defined by the two parental strains (Fig. 1, dark gray bars). Note, however, that variance within any strain is low, relative to that observed across strains, the coefficient of variation for each strain being less than 4%. Resampling the parental strains with additional mice at the end of the experiment confirmed no procedural or criterion drift contaminating our data collection procedures (Fig. 1, second white and black bars). These data show that horizontal cell number is a highly heritable trait, with a narrow-sense additive heritability ( $h^2$ ) of 0.89 (13). Such high heritability makes horizontal cell number a particularly attractive trait for genetic dissection.

**A QTL for Horizontal Cell Number Resides on Distal Chromosome 13.** We mapped the variation in this trait to a narrow locus at the distal tip of Chr 13, peaking at or beyond reference SNP (rs) 4230072 (113.52 Mb) (Fig. 2). This QTL, named “Horizontal cell

Author contributions: L.G., R.W.W., and B.E.R. designed research; I.E.W., M.A.R., D.C.C., and Q.D. performed research; D.C.C., R.A.P., Y.E., and L.G. contributed analytic tools/conditional knockout mice; I.E.W., M.A.R., and D.C.C. analyzed data; and B.E.R. wrote the paper.

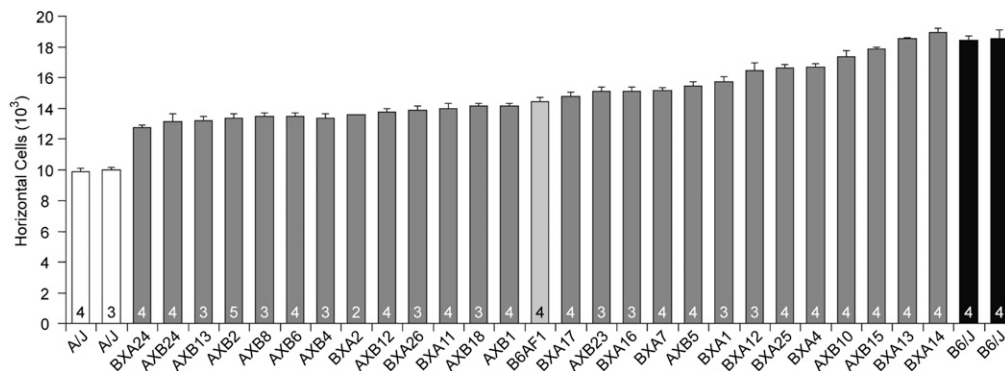
The authors declare no conflict of interest.

\*This Direct Submission article had a prearranged editor.

<sup>1</sup>I.E.W. and M.A.R. contributed equally to this work.

<sup>2</sup>To whom correspondence should be addressed. E-mail: breese@psych.ucsb.edu.

This article contains supporting information online at [www.pnas.org/lookup/suppl/doi:10.1073/pnas.1103253108/-DCSupplemental](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1103253108/-DCSupplemental).



**Fig. 1.** Estimated total number (mean and SEM) of horizontal cells in the retina of *AJ* (white bars) and *B6J* (black bars) strain mice, in the *B6AF1* progeny (light gray bar), and in 26 RI strains of mice derived from the parental strains (the *AXB/BXA* strain set; dark gray bars).  $n$  = number of mice sampled per strain.

number control, Chr 13” (*Hcnc13*) is associated with a significant likelihood ratio statistic (LRS) score of 30.0 ( $P < 0.05$ ), with *B* alleles increasing trait values. The additive effect of two *B* alleles at this locus accounts for ~3,250 cells, being roughly 40% of the difference between the parental strains. Polymorphisms within this QTL must therefore modulate horizontal cell number.

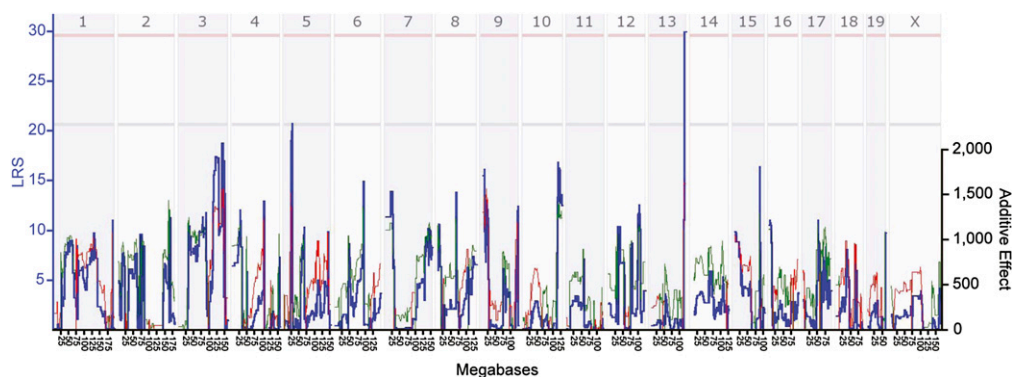
**Two Candidate Genes That Modulate Retinal Nerve Cell Number Are Positioned at the QTL.** One candidate gene within this QTL is follistatin (*Fst*), positioned at 115.24 Mb. Administration of follistatin during the neurogenetic period has been shown to increase the number of retinal horizontal cells in chicken embryos (14), and so we examined the effect of disrupting *Fst* gene expression by excising a floxed allele of *Fst* in cells expressing the *Cre* gene within the developing mouse retina (15, 16). Effective cre-mediated recombination of *Fst* within the retina was confirmed by qPCR, yet Fig. 3*A* shows, for the opposite retinas of these *Fst*-conditional knock-out (CKO) mice, no significant change in horizontal cell number compared with control littermates, in either the center or the periphery of the retina.

Another candidate gene within this QTL is *Isl1*, positioned at 117.08 Mb. Recent studies have shown its conditional deletion in the mouse retina to lower the number of cholinergic amacrine cells, bipolar cells, and retinal ganglion cells (17, 18). We consequently examined horizontal cell number in the *Isl1*-CKO retina and in littermate controls. Horizontal cell density was significantly elevated in the central parts of the *Isl1*-CKO retina (Fig. 3*B* and *C*, *Left*), being increased by 41%. Densities in the

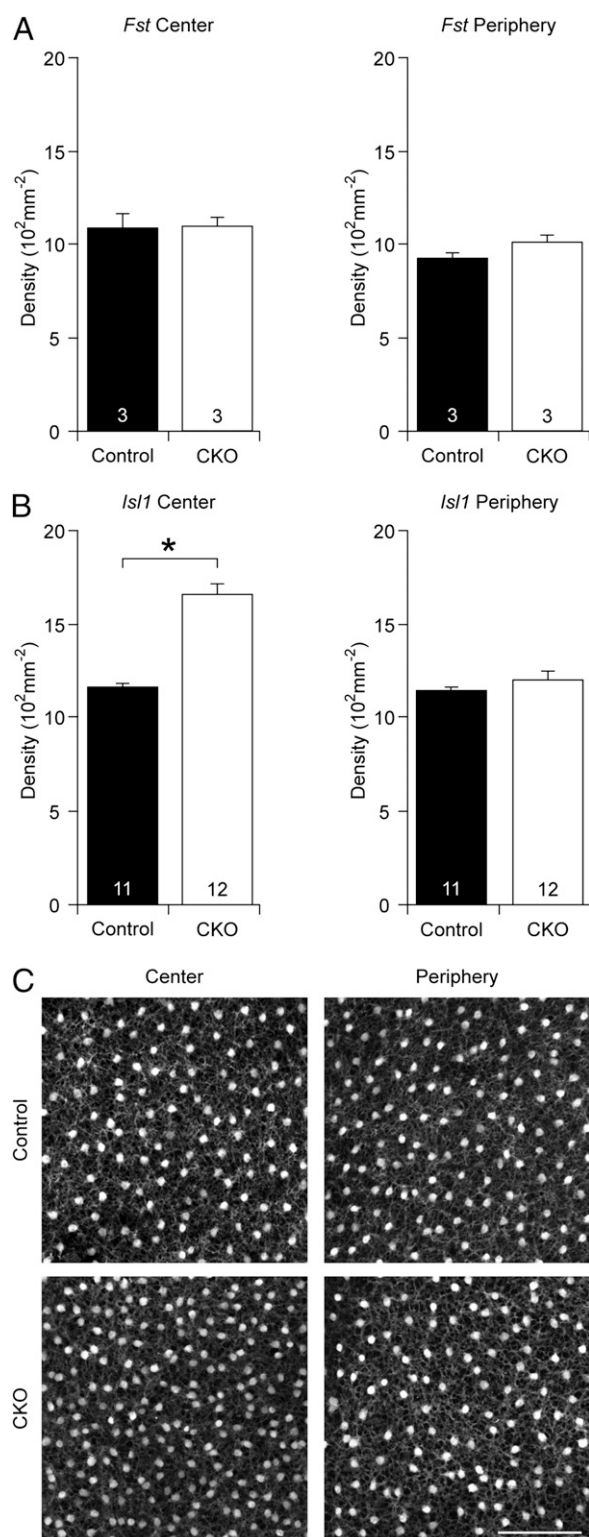
periphery were only marginally increased, by 8% (Fig. 3*B* and *C*, *Right*), likely due to the mosaicism in cre-recombinase activity: *Six3-cre* drives *LacZ* reporter expression in the *Rosa26* reporter line more thoroughly in the central than in the peripheral parts of the mouse retina (e.g., figure 1e in ref. 19). Conditional deletion of *Isl1* therefore increases the number of horizontal cells.

***Isl1* Is Not Expressed in Horizontal Cell Precursors.** The transcription factor genes *Foxn4*, *Ptf1A*, and *Prox1*, each of which is thought to play a hierarchical role in the specification of retinal cell fate, have all been shown to produce profound deficits in horizontal cell number when knocked out (20–22). How *Isl1* interacts with this hierarchy is unknown. *Isl1* is not normally expressed by horizontal cells in the mature mouse retina (23, 24), and we have confirmed that it is not expressed in *Prox1*-expressing precursors, a subset of which will become horizontal cells (20) (Fig. 4): Crossing mice with tamoxifen-dependent cre-recombinase driven by the *Isl1* locus (25) with the *Rosa26* reporter line expressing *LacZ* (26), and administering tamoxifen on embryonic (E) days E12.5–E16.5, yielded no *Prox1*<sup>+</sup> cells (magenta) on E18.5 that also expressed the *LacZ* gene product,  $\beta$ -galactosidase (green).

*Isl1* is, however, expressed by one type of horizontal cell in the chick retina, the axonless horizontal cell, whereas another LIM homeodomain transcription factor gene, *Lim1*, is expressed exclusively by the axon-bearing horizontal cells (14, 27), the only type present in the mouse retina (28). Overexpression of *Isl1* in postmitotic horizontal cells in the chick retina results in a switching to the axonless fate, presumably due to a repression of



**Fig. 2.** Genomewide QTL mapping of horizontal cell number. Each chromosome, from centromere to telomere, is represented left to right along the x axis. The *Right* y axis plots the additive effect, being the extent to which the presence of an *A* (green line) or *B* (red line) allele contributes to an increase in horizontal cell number. The *Left* y axis plots the strength of the linkage of horizontal cell number to different intervals along the genome (blue line). Pink and gray horizontal lines indicate genomewide likelihood ratio statistics (LRS) for significant ( $<0.05$ ) and suggestive ( $<0.67$ ) QTL, respectively. A single significant QTL, *Hcnc10*, at the distal end of chromosome 13 is present.



**Fig. 3.** (A) Conditional deletion of *Fst* does not alter horizontal cell density, in either the central or peripheral parts of the retina. (B) Conditional deletion of *Isl1* yields an increase in horizontal cell density, relative to littermate control mice, which is most pronounced in the central parts of the retina relative to the periphery. *n* = number of mice sampled. (C) Retinal fields from the center and periphery of *Isl1*-CKO mice and control littermates immunolabeled for calbindin. (Scale bar, 100 μm.)

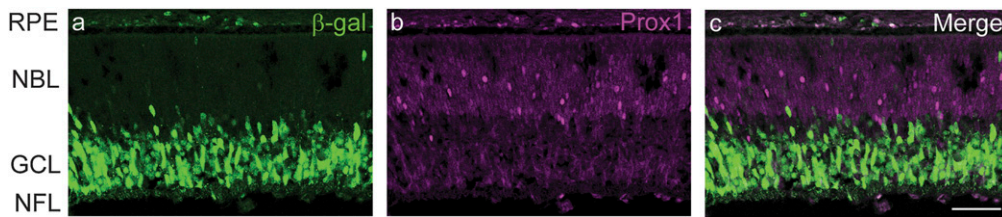
endogenous *Lim1* by *Isl1* (27). Conversely, the increase in horizontal cells, shown here in the *Isl1* conditional knockout, suggests a role for *Isl1* that is incompatible with *Lim1* expression and horizontal cell differentiation. These two LIM homeodomain transcription factor genes, *Isl1* and *Lim1*, are therefore likely to be mutually inhibitory within the mouse retina, as they are in other systems (29, 30), participating in the production of horizontal cells (27). The phenotypic difference in horizontal cell number between the parental strains, therefore, may arise from allelic variation within a regulatory sequence for *Isl1*.

#### **Cis-Acting Regulatory Variants Control Parental Differences in *Isl1* Expression.**

We sequenced the *Isl1* gene and confirmed the presence of four previously documented synonymous sequence differences between A/J and B6/J, identifying no additional SNPs within the coding region. We next examined potential regulatory regions of this gene, including the putative promoter (~1 kb upstream of the transcriptional start site), 5' UTR, the first two introns, and the 3' UTR, using the more complete sequencing of A/J as part of the Mouse Genome Project by the Wellcome Trust Sanger Institute (<http://www.sanger.ac.uk/resources/mouse/genomes/>). Sequence analysis revealed numerous possible regulatory SNPs between these two strains, including previously undocumented SNPs (Table S1). To ascertain whether the parental strains differ in their expression of *Isl1*, we measured transcript levels using qPCR during the period of horizontal cell genesis as well as in maturity, finding significantly greater levels of expression in A/J, the strain with fewer horizontal cells (Fig. 5A). We also compared the relationship between *Isl1* expression and horizontal cell number across the two parental and 26 RI strains of the AXB/BXA strain set from our previous microarray analysis of adult whole eye mRNA (6), available on GeneNetwork for public access (accession no. GN210). We found a significant negative correlation between *Isl1* expression and horizontal cell number across these 28 strains ( $r = -0.525$ ,  $P$  value = 0.00357).

To test directly whether differences in *Isl1* expression between the parental strains are at least partially controlled by a *cis*-regulatory variant, we examined the allele-specific expression (ASE) of *Isl1* in the retinas of reciprocal F<sub>1</sub> offspring of the A/J and B6/J strains. Significant differences between the parental alleles in the F<sub>1</sub> animals would indicate *cis*-modulation, whereas *trans*-acting regulation of *Isl1* expression would be reflected by the lack of difference between parental allelic expression, because both *A* and *B* alleles of *Isl1* will be equally affected by *trans*-factors (31). The significance of the difference in allelic expression was evaluated by comparing the deviation of the ratio of allelic expression in the mRNA from retina taking into account the difference in amplification efficiency of the alleles using genomic DNA (gDNA) as a control. In the genome of F<sub>1</sub> hybrids, there is a copy of each of the alleles in every somatic cell; the ratio in allelic abundance following amplification of the gDNA is therefore expected to be 1. Deviation from this ratio, however, is frequent, being a result of various factors such as secondary DNA structure, presence of unaccounted DNA polymorphisms that can influence the efficiency of allelic amplification, and the ratio between alleles, which should affect gDNA as well as the mRNA/cDNA equally. Any difference in allelic expression due to *cis*-elements alone should be expressed as a deviation in the ratio of allelic abundance profiled in the mRNA of reciprocal F<sub>1</sub> hybrids relative to the gDNA. We found the expression of the *A* allele in *Isl1* to be significantly greater, in both reciprocal hybrids (Fig. 5B), validating the mechanism of *cis*-regulation of this gene.

**An SNP Creates an E-Box in the *Isl1* 5' UTR in B6/J Mice.** We used two predictive programs of transcription factor binding sites, TESS and MATCH, to identify potentially functional consequences of sequence differences between A/J and B6/J in the promoter re-



**Fig. 4.** (A) Cells of the *Isl1* lineage born between E12.5 and E16.5, positive for the *LacZ* reporter gene product  $\beta$ -galactosidase (green), are all positioned in the inner retina on E18.5. (B) Horizontal cell precursors, identified by their expression of *Prox1* (magenta), are known to migrate beyond the future horizontal cell stratum in the outer retina, raising the possibility that they too are derived from the *Isl1*-expressing cells in the inner retina at these early stages. (C) When both channels are viewed together, however, not a single *Prox1*<sup>+</sup> precursor is found to coexpress  $\beta$ -gal. Samples are from the central retina, but identical results were obtained at the far retinal periphery. (Scale bar, 50  $\mu$ m.)

gion and 5' UTR of *Isl1* (32, 33). One difference stood out, in the 5' UTR where multiple bHLH transcription factors were predicted by both programs to bind to the B6/J but not A/J sequence (rs545658554). This group of transcriptional regulators targets a palindromic hexonucleotide DNA sequence known as an E-box, CANNTG (34). The inner two nucleotides of the E-box may vary, but the outer bases are conserved (35). The E-box is not present in A/J due to the G nucleotide located at position 5 (CAGCT/GG). The A allele with the G nucleotide is more highly conserved (Fig. 5C), being shared with five other vertebrates, including dog (XM\_848628), horse (XM\_001916545), cow (NM\_001099130), chimpanzee (XM\_001150577), and human (NM\_002202). This is also true of wild-derived subspecies and species of *Mus*, including *Mus musculus musculus*, *Mus musculus castaneus*, and *Mus spretus*, which have also been sequenced as part of the Mouse Genome Project. In marked contrast, 14 other strains of mice have the less conserved B allele. This suggests that the E-box is a regulatory element now widely propagated in a subset of laboratory mice, including B6/J.

## Discussion

Horizontal cells display a conspicuous variation in their number within the mouse retina that is independent of variation in the overall size of the retina itself. This variation is highly heritable, with surprisingly low within-strain differences—a coefficient of variation of less than 4% for every RI strain examined. Although sets of identified neurons in invertebrates often display this level of tight genetic control (36), there is no precedent for such precision in the mammalian central nervous system. The minimal residual nongenetic variation is presumably due to a combination of intrinsic developmental noise and sampling error (7).

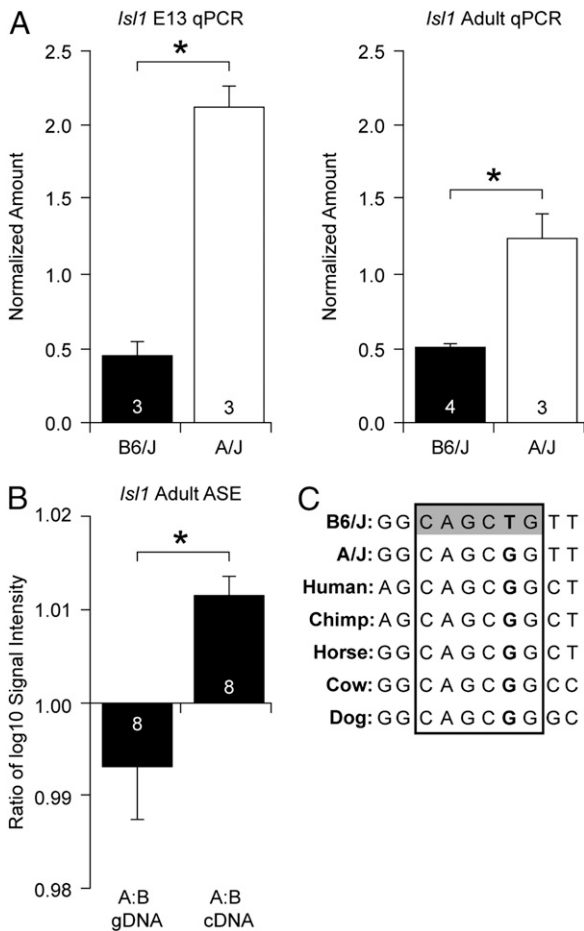
The heritable variation in horizontal cell number is independent of the variation in other types of retinal neurons in these same strains. Cone photoreceptors, for instance, are 70% more abundant in the B6/J strain relative to A/J (6), whereas dopaminergic amacrine cells exhibit a fourfold variation in total number across this same family of RI strains (4). The former trait mapped to a locus on Chr 10 (6), whereas the latter mapped to a locus on Chr 7, where at least three genes modulating dopaminergic amacrine cell number were shown to be present (4). The variation in horizontal cell number mapped to a locus on Chr 13, where two promising prospective candidates were identified, *Fst* and *Isl1*. Despite the documented role for *Fst* in controlling horizontal cell number in the chick retina (14, 37), no comparable effect was detected in *Fst*-CKO retinas. Conditional knock-out of *Isl1*, by contrast, produced a substantial increase in horizontal cells in a retina with drastically reduced numbers of ganglion cells, cholinergic amacrine cells, and bipolar cells (17, 18), indicating a suppressive role for *Isl1* upon horizontal cell number. Indeed, *Isl1* mRNA levels were greater in the A/J strain with lower numbers of horizontal cells during the period of horizontal cell neurogenesis (38). We found only synonymous

SNPs in the coding region of the *Isl1* gene, but detected an SNP in the 5' UTR, which creates a bHLH binding motif in the B6/J strain that may participate in the regulation of *Isl1* expression. Consistent with this, we have shown that *Isl1* transcript levels are controlled by a *cis*-acting variant in retinal tissue.

Exactly how the modulation of *Isl1* expression affects horizontal cell number remains to be determined. Previously, we had shown that all horizontal cells arise from the *Math5*<sup>+</sup> precursors that also generate the *Isl1*<sup>+</sup> amacrine and ganglion cells as well as certain photoreceptor cells (39, 40). Given that *Isl1*-lineage cells do not produce horizontal cells (Fig. 4), that *Lim1* protein is found only in horizontal cells (24), and the inhibitory relationship between these two genes (29, 30), a mutually exclusive expression pattern of *Isl1* and *Lim1* among common precursors could participate in determining horizontal cell number. The conditional knockout of *Isl1* may alleviate additional precursors of such repression of *Lim1*, allowing them to differentiate as horizontal cells. In the B6/J retina, the presence of the E-box may ensure further repression of *Isl1* to allow more cells to express *Lim1*, becoming horizontal cells. This E-box-mediated repression of *Isl1* expression is likely regulated by a bHLH transcriptional repressor specific to horizontal cell precursors. In other precursors that give rise to *Isl1*<sup>+</sup> cells in the B6/J retina (23), *Isl1* expression may not be suppressed, due to the absence of this bHLH transcriptional repressor and/or to the presence of other transcriptional activators.

bHLH transcription factors have been shown to be integral during embryonic development, playing roles in myogenesis, heart development, hematopoiesis, and neurogenesis (34). Of the various bHLH transcription factors predicted to bind the E-box, *E2A* and *NeuroD1* are expressed in developing vertebrate retina (41), and have been reported to play a role in retinal neurogenesis and differentiation (42, 43). In these instances, *E2A* and *NeuroD1* are considered to be proneural bHLH proteins, acting as transcriptional activators. However, they have been described in other tissues to function as transcriptional repressors during development (44, 45). In the present instance, their proposed action upon *Isl1* is predicted to be repressive, given the greater number of horizontal cells in B6/J and the greater *Isl1* mRNA expression observed in the A/J strain. The creation of a novel E-box due to an SNP, leading to altered gene expression, is not unprecedented (46), including an instance when this resulted in the suppression of gene expression (47). This polymorphism in the *Isl1* 5' UTR may therefore contribute to the variation in horizontal cell number documented herein.

Prominent variation in nerve cell number is predicted to have functional consequences for neural organization and circuitry (36). In the case of the horizontal cells, dendritic field area is modulated in these parental strains of mice to maintain an average dendritic overlap of around 6.0 (12). Because variation in the number of their afferents, the cone photoreceptors (6), is



**Fig. 5.** (A) Retinal expression of *Is11* during the period of horizontal cell neurogenesis, at E13, shows a nearly fivefold difference favoring *A/J* ( $P = 0.0007$ , Student's  $t$  test). A difference is also detected in maturity ( $P = 0.0028$ ). (B) ASE confirms *cis*-regulation of *Is11* in AXB/BXA, evidenced by testing the deviation of the allelic expression changes in the retina from the allelic ratio in the genomic DNA in reciprocal  $F_1$  hybrids. ( $P = 0.007$ ). (C) An SNP in the *Is11* 5' UTR creates an E-box sequence in the B6/J strain. The A SNP is the conserved allele. (Alignment generated with CLC Sequence Viewer).

independent of the variation in horizontal cell number across this large family of RI strains, this ratio must be unique for each strain of mouse, yet the horizontal cells modulate their connectivity with the population of afferents to ensure a uniform sampling of every pedicle by six neighboring horizontal cells (12). The genetic determinants of nerve cell number, therefore, by setting the grain of both the photoreceptor and horizontal cell mosaics independently, indirectly modulate the differentiation and connectivity of the horizontal cell, a critical postreceptoral interneuron shaping the visual signal within the vertebrate retina.

## Materials and Methods

**Mouse Strains.** B6/J and A/J mice, their  $F_1$  progeny (B6AF1), and RI mice from 26 strains in the AXB/BXA strain set were obtained from The Jackson Laboratory. A minimum of three mice per strain were examined, with one exception (BXA2,  $n = 2$ ), sampling a single eye from each mouse. The RI strains were originally generated by the continuous inbreeding of  $F_1$  progeny (either AB6F1 to make AXB strains or B6AF1 to make BXA strains) through successive generations to the two chromosomes that are homozygous but recombined versions of the yield parental haplotypes (for further details on the derivation of these strains, see [www.genenetwork.org/dbdoc/AXBXA-Publish.html](http://www.genenetwork.org/dbdoc/AXBXA-Publish.html)). *Fst*-CKO mice and their control littermates were produced by crossing *Chx10-GFPcre*-expressing mice [Tg(*Chx10*-EGFP/*cre*, -ALPP)2Clc] (15) with mice bearing a floxed allele of *Fst* (*Fst*<sup>tm2Zuk</sup>), and subsequently bred

with mice homozygous for floxed *Fst* allele (16). Three *Fst*-CKO mice and three littermate control mice were examined. *Is11*-CKO mice and their control littermates were produced as described previously. Briefly, *Six3-cre*-expressing mice [Tg(*Six3-cre*)69Frty] (48) were crossed with mice harboring either a floxed allele of *Is11* or an *Is11-lacZ* knock-in allele (*Is11*<sup>tm1Gan</sup>) and subsequently bred with mice homozygous for floxed *Is11* allele (18). Twelve *Is11*-CKO mice and 11 littermate control mice were examined. All mice used in the present study were between 3 and 12 wk of age.

**Retinal Immunofluorescence and Quantification of Horizontal Cell Number.** Standard procedures for perfusion, dissection, and immunofluorescence were used to prepare retinal wholemounts for quantification of horizontal cell number, as described previously by us (9). Mean horizontal cell density (derived from sampling eight fields totaling ~15% of the retinal surface area) was multiplied by retinal area to estimate total horizontal cell number. Further details are provided in *SI Materials and Methods*.

**QTL Mapping.** Each of the RI strains has a unique mix of A and B genotypes across all chromosomes (with the exceptions of the Y chromosome and the mitochondrial genome). QTL mapping takes advantage of this nearly random recombination of parental genotypes to estimate the genetic control of a given trait (in our case, horizontal cell number) and the presence of A or B alleles across the genome. Each of the RI strains has been genotyped with high-density microsatellite and SNP markers (49). QTL mapping of horizontal cell number was performed using standard interval mapping and the genotypes of 3,785 informative markers using computational and visualization tools available in GeneNetwork (GN) ([www.GeneNetwork.org](http://www.GeneNetwork.org)). GeneNetwork implements now standard methods of simple and composite interval mapping and estimates the genomewide  $P$  value of a type I error by permutation. The primary horizontal cell data have been permanently deposited in GeneNetwork as phenotype accession identifier GN no. 10132 (horizontal cell number per retina) in the mouse AXB/BXA Published Phenotypes database.

**Lineage Marking.** To study the lineage of *Is11*-expressing retinal cells, male tamoxifen-inducible *Is11-cre* mice (*Is11*<sup>tm1(cre/Esr1\*)Krc</sup>) (25) (kindly provided by Dr. Amy Kiernan, University of Rochester, Rochester, NY) were crossed to females that carried the Rosa reporter allele, *R26R-LacZ* [Gt(*ROSA*)26Sor<sup>tm1Sor</sup>] (26) (The Jackson Laboratory). Time-mated pregnant mice were injected intraperitoneally with tamoxifen at 4 mg/30 g body weight at E12.5, E13.5, E14.5, E15.5, and E16.5. Embryos were harvested at E18.5, fixed for 2 h in 4% paraformaldehyde in PBS, and saturated in 30% sucrose dissolved in PBS at 4 °C overnight. Samples were then embedded in OCT and sectioned at 20  $\mu$ m on a cryostat. The following antibodies were used: chicken anti- $\beta$ -galactosidase (1:1,000; Abcam) and rabbit anti-Prox1 (1:1,000; Covance). Fluorescence images were captured and analyzed by a Zeiss LSM 510 confocal microscope using a 20 $\times$  objective.

**Quantitative RT-PCR.** *Is11* transcript levels were quantified during the period of horizontal cell genesis and in maturity using qPCR, as described previously (4) with a few modifications. Further details are provided in *SI Materials and Methods*.

**Rapid Amplification of the cDNA Ends and cDNA Sequencing.** The *Is11* mouse sequence available in Ensembl (ENSMUST00000036060.4) and the Primer 3 program were used to design primers to amplify and sequence the entire transcript. Further details are provided in *SI Materials and Methods*.

**Allele-Specific Expression Assay.** An ASE was performed using male and female pooled samples from each reciprocal  $F_1$  cross of the A/J and B6/J strains, AB6F1 and B6AF1, as previously described (31). Further details are provided in *SI Materials and Methods*.

**Statistics.** Student's  $t$  test was used for all comparisons between A/J versus B6/J, and between conditional knock-out mice and their respective littermate controls. A one-tailed Student's  $t$  test with unequal variance examined the difference in allelic expression of *Is11* by testing the deviation in the ratio of allelic abundance (log<sub>10</sub>) profiled in the mRNA of reciprocal  $F_1$  hybrids relative to the gDNA, as previously described (31). A one-tailed test was chosen because the ASE sought to test the direction of the difference detected by qPCR, either confirming it (therefore being *cis*-regulated) or finding no difference (and therefore *trans*-regulated).

**ACKNOWLEDGMENTS.** We thank H. Nasser for assistance with tissue preparation, P. Keeley for assistance with harvesting fresh retinas for sequencing, K. Taylor and M. Radeke for assistance and advice with qPCR,

respectively, and T. Cunningham for assistance with the ASE assay. This research was supported by the NIH (EY-011087 and EY-019968 to B.E.R.).

1. Curcio CA, Sloan KRJ, Jr., Packer O, Hendrickson AE, Kalina RE (1987) Distribution of cones in human and monkey retina: Individual variability and radial asymmetry. *Science* 236:579–582.
2. Wikler KC, Williams RW, Rakic P (1990) Photoreceptor mosaic: Number and distribution of rods and cones in the rhesus monkey retina. *J Comp Neurol* 297:499–508.
3. Curcio CA, Allen KA (1990) Topography of ganglion cells in human retina. *J Comp Neurol* 300:5–25.
4. Whitney IE, Raven MA, Ciobanu DC, Williams RW, Reese BE (2009) Multiple genes on chromosome 7 regulate dopaminergic amacrine cell number in the mouse retina. *Invest Ophthalmol Vis Sci* 50:1996–2003.
5. Whitney IE, Keeley PW, Raven MA, Reese BE (2008) Spatial patterning of cholinergic amacrine cells in the mouse retina. *J Comp Neurol* 508:1–12.
6. Whitney IE, Raven MA, Lu L, Williams RW, Reese BE (2011) A QTL on chromosome 10 modulates cone photoreceptor number in the mouse retina. *Invest Ophthalmol Vis Sci* 52:3228–3236.
7. Williams RW, Strom RC, Goldowitz D (1998) Natural variation in neuron number in mice is linked to a major quantitative trait locus on Chr 11. *J Neurosci* 18:138–146.
8. Strom RC, Williams RW (1998) Cell production and cell death in the generation of variation in neuron number. *J Neurosci* 18:9948–9953.
9. Raven MA, Stagg SB, Reese BE (2005a) Regularity and packing of the horizontal cell mosaic in different strains of mice. *Vis Neurosci* 22:461–468.
10. Williams RW, Strom RC, Zhou G, Yan Z (1998) Genetic dissection of retinal development. *Semin Cell Dev Biol* 9:249–255.
11. Raven MA, Stagg SB, Nassar H, Reese BE (2005b) Developmental improvement in the regularity and packing of mouse horizontal cells: Implications for mechanisms underlying mosaic pattern formation. *Vis Neurosci* 22:569–573.
12. Reese BE, Raven MA, Stagg SB (2005) Afferents and homotypic neighbors regulate horizontal cell morphology, connectivity, and retinal coverage. *J Neurosci* 25:2167–2175.
13. Hegmann JP, Possidente B (1981) Estimating genetic correlations from inbred strains. *Behav Genet* 11:103–114.
14. Edqvist PH, Lek M, Boije H, Lindbäck SM, Hallböök F (2008) Axon-bearing and axonless horizontal cell subtypes are generated consecutively during chick retinal development from progenitors that are sensitive to follistatin. *BMC Dev Biol* 8:46.
15. Rowan S, Cepko CL (2004) Genetic analysis of the homeodomain transcription factor Chx10 in the retina using a novel multifunctional BAC transgenic mouse reporter. *Dev Biol* 271:388–402.
16. Jorgez CJ, Klysiak M, Jamin SP, Behringer RR, Matzuk MM (2004) Granulosa cell-specific inactivation of follistatin causes female fertility defects. *Mol Endocrinol* 18:953–967.
17. Pan L, Deng M, Xie X, Gan L (2008) ISL1 and BRN3B co-regulate the differentiation of murine retinal ganglion cells. *Development* 135:1981–1990.
18. Elshatory Y, et al. (2007b) Islet-1 controls the differentiation of retinal bipolar and cholinergic amacrine cells. *J Neurosci* 27:12707–12720.
19. Poché RA, et al. (2008) Somal positioning and dendritic growth of horizontal cells are regulated by interactions with homotypic neighbors. *Eur J Neurosci* 27:1607–1614.
20. Dyer MA, Livesey FJ, Cepko CL, Oliver G (2003) Prox1 function controls progenitor cell proliferation and horizontal cell genesis in the mammalian retina. *Nat Genet* 34:53–58.
21. Fujitani Y, et al. (2006) Ptf1a determines horizontal and amacrine cell fates during mouse retinal development. *Development* 133:4439–4450.
22. Li S, et al. (2004) *Foxn4* controls the genesis of amacrine and horizontal cells by retinal progenitors. *Neuron* 43:795–807.
23. Elshatory Y, Deng M, Xie X, Gan L (2007a) Expression of the LIM-homeodomain protein Isl1 in the developing and mature mouse retina. *J Comp Neurol* 503:182–197.
24. Poché RA, et al. (2007) Lim1 is essential for the correct laminar positioning of retinal horizontal cells. *J Neurosci* 27:14099–14107.
25. Laugwitz KL, et al. (2005) Postnatal Isl1+ cardioblasts enter fully differentiated cardiomyocyte lineages. *Nature* 433:647–653.
26. Soriano P (1999) Generalized lacZ expression with the ROSA26 Cre reporter strain. *Nat Genet* 21:70–71.
27. Suga A, Taira M, Nakagawa S (2009) LIM family transcription factors regulate the subtype-specific morphogenesis of retinal horizontal cells at post-migratory stages. *Dev Biol* 330:318–328.
28. Peichl L, González-Soriano J (1994) Morphological types of horizontal cell in rodent retinae: A comparison of rat, mouse, gerbil, and guinea pig. *Vis Neurosci* 11:501–517.
29. Sun Y, et al. (2008) A central role for Islet1 in sensory neuron development linking sensory and spinal gene regulatory programs. *Nat Neurosci* 11:1283–1293.
30. Kania A, Johnson RL, Jessell TM (2000) Coordinate roles for LIM homeobox genes in directing the dorsoventral trajectory of motor axons in the vertebrate limb. *Cell* 102:161–173.
31. Ciobanu DC, et al. (2010) Detection, validation, and downstream analysis of allelic variation in gene expression. *Genetics* 184:119–128.
32. Kel AE, et al. (2003) MATCH: A tool for searching transcription factor binding sites in DNA sequences. *Nucleic Acids Res* 31:3576–3579.
33. Schug J (2008) Using TESS to predict transcription factor binding sites in DNA sequence. *Curr Protoc Bioinformatics* 21:2.6.1–2.6.15.
34. Massari ME, Murre C (2000) Helix-loop-helix proteins: Regulators of transcription in eucaryotic organisms. *Mol Cell Biol* 20:429–440.
35. Beltran AC, Dawson PE, Gottesfeld JM (2005) Role of DNA sequence in the binding specificity of synthetic basic-helix-loop-helix domains. *ChemBioChem* 6:104–113.
36. Williams RW, Herrup K (1988) The control of neuron number. *Annu Rev Neurosci* 11:423–453.
37. Moreira EF, Adler R (2006) Effects of follistatin overexpression on cell differentiation in the chick embryo retina. *Dev Biol* 298:272–284.
38. Young RW (1985) Cell differentiation in the retina of the mouse. *Anat Rec* 212:199–205.
39. Feng L, et al. (2006) Requirement for Bhlhb5 in the specification of amacrine and cone bipolar subtypes in mouse retina. *Development* 133:4815–4825.
40. Yang Z, Ding K, Pan L, Deng M, Gan L (2003) Math5 determines the competence state of retinal ganglion cell progenitors. *Dev Biol* 264:240–254.
41. Blackshaw S, et al. (2004) Genomic analysis of mouse retinal development. *PLoS Biol* 2:E247.
42. Yang HJ, Silva AO, Koyano-Nakagawa N, McLoon SC (2009) Progenitor cell maturation in the developing vertebrate retina. *Dev Dyn* 238:2823–2836.
43. Liu H, et al. (2008) NeuroD1 regulates expression of thyroid hormone receptor 2 and cone opsins in the developing mouse retina. *J Neurosci* 28:749–756.
44. Perez-Moreno MA, et al. (2001) A new role for E12/E47 in the repression of E-cadherin expression and epithelial-mesenchymal transitions. *J Biol Chem* 276:27424–27431.
45. Itkin-Ansari P, et al. (2005) NeuroD1 in the endocrine pancreas: Localization and dual function as an activator and repressor. *Dev Dyn* 233:946–953.
46. Teng AC, Adamo K, Tesson F, Stewart AF (2009) Functional characterization of a promoter polymorphism that drives ACSL5 gene expression in skeletal muscle and associates with diet-induced weight loss. *FASEB J* 23:1705–1709.
47. Wang D, et al. (2008) Regulatory polymorphism in vitamin K epoxide reductase complex subunit 1 (VKORC1) affects gene expression and warfarin dose requirement. *Blood* 112:1013–1021.
48. Furuta Y, Lagutin O, Hogan BLM, Oliver GC (2000) Retina- and ventral forebrain-specific Cre recombinase activity in transgenic mice. *Genesis* 26:130–132.
49. Williams RW, Gu J, Qi S, Lu L (2001) The genetic structure of recombinant inbred mice: High-resolution consensus maps for complex trait analysis. *Genome Biol*, 2: research0046.0041–0046.0018.



# Supporting Information

Whitney et al. 10.1073/pnas.1103253108

## SI Materials and Methods

**Retinal Immunofluorescence.** Mice were given a lethal dose of sodium pentobarbital (120 mg/kg, i.p.) and, once heavily anesthetized, they were intracardially perfused with 0.9% saline followed by 4% paraformaldehyde in 0.1 M sodium phosphate buffer (pH 7.2 at 20 °C). (All procedures were conducted under authorization by the respective institutional animal care and use committees at the University of California at Santa Barbara, Baylor College of Medicine, and the University of Rochester and conform to the American Veterinary Medical Association's *Guidelines on Euthanasia*). Retinas were dissected from the eye, prepared as wholemounts, and then labeled using standard immunofluorescence procedures, as described elsewhere (1). The retinas were incubated with a mouse monoclonal antibody to calbindin D-28 (1:10,000; Sigma) and subsequently incubated with a rabbit antimouse IgG conjugated to Cy2 (1:200; Jackson Labs). Only entire, intact, whole retinas were used for subsequent quantification.

**Quantification of Horizontal Cell Number.** Images of the mosaic of horizontal cells in these wholemounts were obtained using a Nikon FXA fluorescence photomicroscope and an Olympus DP11 digital camera. Labeled horizontal cells were sampled near the optic nerve head (central samples) and near the retinal periphery (peripheral samples) in the four quadrants of each wholemounted retina, from which a mean density for each animal was determined. Each sampled field occupied 0.225 mm<sup>2</sup> of retinal area (aspect ratio of 1:1.25), so that ~15% of the entire retinal surface was analyzed. Retinal areas were also measured (Bioquant Nova Prime; R&M Biometrics), and multiplied by mean density to estimate the total number of horizontal cells per retina. (All procedures are identical to those we have used elsewhere to compare horizontal cell number across different laboratory strains of mice (1). Representative fields in Fig. 3 were collected with an Olympus FV500 scanning laser confocal microscope using a 40× oil immersion lens. Image stacks were collected at 1-μm intervals over a total depth of 20 μm, from which maximum projections were generated.

**Quantitative RT-PCR.** Time-mated pregnant mice were heavily sedated and embryos removed for the dissection of retinal tissue. The Theiler stages of embryonic mouse development were used to collect gestationally matched A/J and B6/J embryos on embryonic (E) day 13. Each embryonic sample consisted of pooled retinas from a minimum of five embryos per litter. Retinal tissue was also collected from adult A/J and B6/J mice ranging in age from 114 to 123 d old, averaging 121 d; here, each sample consisted of two pooled retinas from one animal. Following the extraction and

verification of RNA integrity, single-strand cDNA was synthesized for quantitative real time RT-PCR that was performed with the BioRad MyiQ Single Color Real-Time PCR Detection system. Primers 5' TGAGTGAAGTTCGCCTTGCAAAG 3' forward and 5' CATGCTGTTGGGTGTATCTGGG 3' reverse were used in the amplification of *Isl1* transcripts. Normalized amounts were generated using four housekeeping genes for both the embryonic and adult time points.

**Rapid Amplification of the cDNA Ends and cDNA Sequencing.** RNA was extracted from postnatal day 10 B6/J and A/J retinas, as described previously (2), and used for first strand cDNA synthesis (GE Healthcare), which was amplified (GoTaq Flexi DNA polymerase; Promega) using pairs of primers that covered the *Isl1* transcript. The 5' and 3' rapid amplification of cDNA ends (RACE) was performed using the FirstChoice RLM-RACE kit (Ambion), according to the manufacturer's protocols, followed by sequencing of the PCR products using dye terminators and capillary electrophoresis (ABI3130; Applied Biosystems). Sequencer software (Gene Codes, version 4.10.1) was used to assemble the sequences and to identify polymorphisms.

**Allele-Specific Expression Assay.** Samples from each reciprocal F<sub>1</sub> cross of the A/J and B6/J strains, AB6F1 and B6AF1, were used, each sample consisting of pooled retinas from two animals averaging 44 d of age. The RNA was extracted from retinas as described previously (2), and genomic DNA was extracted from the corresponding tail clips using the DNeasy Blood and Tissue kit (Qiagen). The DNA samples were used for normalization and assayed at the same time with the RNA pools. First strand cDNA was synthesized using the iScript cDNA synthesis kit (BioRad); cDNA and genomic DNA were amplified (GoTaq Flexi DNA polymerase; Promega) using primers located in the same exon (5' AAGCAGCCGGAGAAGACCAC 3' and 5' TCCTTCATGAGCGCATCTGG 3'). Each sample was amplified and analyzed in duplicate. Excess of dNTPs and primers from PCR products were degraded using ExoSap-IT (United States Biochemical). Single-base extension of the PCR products was performed using SNaPshot (Applied Biosystems) and an extension primer (5' TCCGCAAGGTGTGCAGCTGCTTCTC 3') located next to a synonymous SNP (accession ID 50976547). The extension products were treated with calf intestinal phosphatase (New England Biolabs) to degrade excess of labeled ddNTPs followed by capillary electrophoresis (ABI3130; Applied Biosystems). Quantification of allelic expression was analyzed using Peak Scanner (Applied Biosystems) using the peak height corresponding to each allele.

1. Raven MA, Stagg SB, Reese BE (2005) Regularity and packing of the horizontal cell mosaic in different strains of mice. *Vis Neurosci* 22:461–468.

2. Whitney IE, Raven MA, Ciobanu DC, Williams RW, Reese BE (2009) Multiple genes on chromosome 7 regulate dopaminergic amacrine cell number in the mouse retina. *Invest Ophthalmol Vis Sci* 50:1996–2003.

**Table S1. SNPs and short indels between B6/J and A/J within the coding and putative regulatory regions of *Isl1***

Location	Base pair (mm9)	Reference SNP accession ID	B/A allele
Promoter	117101122	50480348	T/G
Promoter	117101115	549680099	C/T
Promoter	117101086	51306857	T/C
Promoter	117101009	549678357	A/G
Promoter	117100884	51567098	C/G
Promoter	117100098	30049912	A/G
Promoter	117100063	46344459	C/T
5' UTR	117099762	545658554	T/G
5' UTR	117099749	NA	T/C
Intron 1	117099549	NA	G/-
Intron 1	117099447	NA	T/GorC
Intron 1	117099003	NA	A/-
Intron 2	117098742	30022127	C/G
Intron 2	117098293	29532860	G/A
Intron 2	117098279	29247586	A/T
Intron 2	117097993	NA	C/G
Intron 2	117097984	NA	T/G
Intron 2	117097982	NA	A/-
Intron 2	117097979	51985069	A/G
Intron 2	117097701	49007764	A/G
Intron 2	117097700	48694792	G/T
Intron 2	117097690	49652582	T/A
Intron 2	117097497	46219885	C/T
Intron 2	117097437	48981166	G/C
Intron 2	117097223	50326404	A/G
Intron 2	117097088	NA	-/T
Intron 2	117096950	NA	C/-
Intron 2	117096949	NA	T/-
Intron 2	117096744	29972543	G/T
Intron 2	117096717	29775327	G/A
Intron 2	117096568	29783182	G/C
Intron 2	117096351	29730164	C/A
Intron 2	117096127	29229242	T/C
Exon 3	117095622	13466915	T/C
Exon 4	117093450	29525872	C/T
Exon 4	117093287	50976547	G/A
Exon 6	117089739	2957061	G/T
3' UTR	117089521	29635194	T/G
3' UTR	117089225	46842279	C/T
3' UTR	117089182	49620560	A/G
3' UTR	117089161	NA	T/G
3' UTR	117088923-18	NA	-/CCAACCT
3' UTR	117088752	NA	-/T
3' UTR	117088661	NA	T/-

NA, not annotated. Hyphen indicates absence of base.