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Leptospirosis in Fox Squirrels (Sciurus niger) of Larimer County, Colorado, USA

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ABSTRACT: Leptospirosis is a zoonotic disease caused by the bacterium Leptospira interrogans. The organism is typically maintained within a geographic region by colonizing renal tubules of carrier animals and shed into the environment in urine. We assessed whether L. interrogans was present in fox squirrels (Sciurus niger) in Larimer County, Colorado, USA, and whether it is associated with disease. Twenty-two squirrels were trapped from 29 November 2011 to 15 December 2011 for use in an unrelated study. The squirrels were individually housed for 33–65 days and euthanized; no clinical disease was observed. On gross examination, significant renal lesions were observed in 6 of 22 animals (27%). Histologically, affected animals had severe neutrophilic tubulitis with interstitial nephritis. Immunohistochemistry was conducted on the kidneys of all animals and 10 of 22 (45%) were positive for L. interrogans, with varying severity of infection. The same 10 squirrels were serologically positive for antibodies specific to L. interrogans. These results suggest that L. interrogans is present in fox squirrels in Larimer County, Colorado, USA, and may be associated with varying degrees of renal disease. Further investigation into the role of wildlife in the ecology of leptospirosis within the region is warranted.

Key words: Colorado, Leptospira interrogans, leptospirosis, squirrel, wildlife.
approved all animal trapping, handling, and housing procedures.

As part of the original vaccine study, we observed fox squirrels daily and collected blood samples on day 0 of the study (approximately 22–37 days in captivity) and preceding euthanasia (4 or 28 days later). Fox squirrels were anesthetized using isoflurane before being humanely euthanized via intracardiac injection of an overdose of barbiturates (Beuthanasia®, Intervet Inc., Merck Animal Health, Summit, New Jersey, USA). Complete postmortem examinations were conducted and multiple tissues were collected for histologic examination. Tissues were fixed in 10% neutral-buffered formalin, paraffin embedded, cut at 5 µm, and examined by light microscopy. A Warthin-Starry silver stain and leptospiral immunohistochemistry (IHC) were performed on renal tissue from each animal. The IHC was conducted using a primary antibody composed of a cocktail of six antisera: Canicola–Hond Utrecht IV, Grippotyphosa-Andaman, Hardjo-Hardojoprajitino, Copenhageni-M20A, Pomona-Pomona, and Bratislava–Jez Bratislava (US Department of Agriculture, Animal and Plant Inspection Service, National Veterinary Services Laboratory [NVSL], Ames, Iowa, USA). Sera from all fox squirrels were also tested for antibodies specific to five L. interrogans serovars: Canicola, Grippotyphosa, Hardjo, Icterohaemorrhagiae, and Pomona, via a microagglutination test at the NVSL.

Gross examination revealed significant renal lesions in six of 22 animals, (27%: 5 F, 1 M). Bilaterally, the kidneys were mottled with multifocal to coalescing areas of pallor 1–4 mm diameter. On the cut surface, affected areas extended into the parenchyma. All other organs were grossly within normal limits. Histologically, renal lesions were observed in 10 animals (19%; 6 F, 4 M). In the six squirrels in which gross changes were identified, much of the renal parenchyma was effaced by inflammation (Fig. 1A). The interstitial space was expanded by large aggregates of lymphocytes and plasma cells; within affected regions, many tubules were dilated, and tubular lumens contained aggregates of viable and degenerate neutrophils (Fig. 1B) with varying amounts of tubular degeneration and necrosis. In four other animals (8%; 1 F, 3 M), there were only mild, patchy aggregates of interstitial lymphocytes and plasma cells, devoid of the tubulitis observed in the more severely affected animals. Additional tissues reviewed histologically included lung, liver, and multiple sections of intestinal tract. There was mild-to-moderate, bronchiole-associated hyperplasia of the lymphoid tissue in the lung of five animals; two animals had patchy areas of pulmonary inflammation. Very mild periportal, lymphoplasmacytic inflammation was observed in the liver of 14 squirrels, but the cellular infiltrate did not distort the hepatic parenchyma. Two animals had both enteritis and colitis, which was moderate and mononuclear in nature.

Warthin-Starry silver stain highlighted filamentous organisms on the luminal surface of tubular epithelial cells of four animals with severe tubulitis (Fig. 1C). In the six severely affected animals, there was strong immunostaining of the tubular epithelial cells, both on the luminal membrane, but also cytoplasmically, and patchy staining of leukocytes within the interstitium (Fig. 1D). Minimal immunostaining was observed in four additional animals; mild-to-moderate interstitial inflammation had been observed histologically in these four animals, but none had severe tubulitis.

The six severely affected animals had high titers of circulating antibodies to L. interrogans; highest titers were observed against serovars Grippotyphosa, Hardjo, and Icterohaemorrhagiae (Table 1). Sera from four fox squirrels with histologic evidence of a mild infection were also positive for antibodies specific to L. interrogans. Two of these fox squirrels were sampled 28 days apart with little or no increase in antibody titers. This mild infection paired with stable antibody titers
may be indicative of a “carrier” state or animals recovering from infection. Three individuals with minimal or no histologic evidence of renal disease showed low reactivity (titer ≤200) to *L. interrogans* serovars Hardjo or Icterohaemorrhagiae on the last blood collection only. These low and inconsistent titers may indicate a possible exposure while in captivity, recovery from infection with waning antibody titers, or false results.

For almost 25 yr, no case reports or prevalence studies were published, to our knowledge, regarding *L. interrogans* in animals of Larimer County, Colorado, USA. However, between 2005 and 2008, a growing number of domestic dogs in Colorado, USA, were diagnosed with leptospirosis; 61% of those were living in urban or suburban environments, including Larimer County, Colorado, USA, where there were 85 suspected and 15 confirmed cases; Grippotyphosa was the most commonly reported serovar (Veir 2009). Many peridomestic animals also live in these environments, including Norway rats (*Rattus norvegicus*), mice (*Mus musculus*), raccoons, and fox squirrels. Previous studies in Larimer County, Colorado, USA, have shown evidence of *L. interrogans* infection in Norway rats, house mice, and raccoons (Roberts 1972; Al Saadi and Post 1976; Duncan et al., 2012). Although fox squirrels in Colorado, USA, have not been investigated until this study, to our knowledge, a gray squirrel (*Sciurus carolinensis*) from Connecticut, USA, had antibodies to *L. interrogans*
Table 1. Antibody titers (microagglutination test) and renal immunohistochemistry (IHC) results for *Leptospira interrogans* in fox squirrels captured in Larimer County, Colorado, USA.

<table>
<thead>
<tr>
<th>ID (days)</th>
<th>Canicola</th>
<th>Grippio</th>
<th>Hardjo</th>
<th>Ictero</th>
<th>Pomona</th>
<th>Days between serology</th>
<th>Canicola</th>
<th>Grippio</th>
<th>Hardjo</th>
<th>Ictero</th>
<th>Pomona</th>
<th>Serology at euthanasia</th>
<th>Kidney IHC</th>
</tr>
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<tr>
<td>K15 (27)</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>4</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>Negative</td>
</tr>
<tr>
<td>K20 (22)</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>4</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>Negative</td>
</tr>
<tr>
<td>K14 (28)</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>100</td>
<td>—</td>
<td>4</td>
<td>—</td>
<td>—</td>
<td>200</td>
<td>400</td>
<td>—</td>
<td>Very mild, patchy infection</td>
<td></td>
</tr>
<tr>
<td>K16 (24)</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>4</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>200</td>
<td>—</td>
<td>Very mild, patchy infection</td>
<td></td>
</tr>
<tr>
<td>K05 (37)</td>
<td>—</td>
<td>51,200</td>
<td>25,600</td>
<td>51,200</td>
<td>6,400</td>
<td>4</td>
<td>102,400</td>
<td>102,400</td>
<td>102,400</td>
<td>12,800</td>
<td>—</td>
<td>Severe infection</td>
<td></td>
</tr>
<tr>
<td>K06 (37)</td>
<td>—</td>
<td>1,600</td>
<td>400</td>
<td>1,600</td>
<td>100</td>
<td>4</td>
<td>100</td>
<td>1,600</td>
<td>800</td>
<td>6,400</td>
<td>200</td>
<td>Severe infection</td>
<td></td>
</tr>
<tr>
<td>K07 (36)</td>
<td>—</td>
<td>51,200</td>
<td>12,800</td>
<td>25,600</td>
<td>1,600</td>
<td>4</td>
<td>51,200</td>
<td>12,800</td>
<td>51,200</td>
<td>1,600</td>
<td>—</td>
<td>Severe infection</td>
<td></td>
</tr>
<tr>
<td>K10 (29)</td>
<td>—</td>
<td>12,800</td>
<td>6,400</td>
<td>3,200</td>
<td>—</td>
<td>4</td>
<td>25,600</td>
<td>25,600</td>
<td>6,400</td>
<td>100</td>
<td>—</td>
<td>Severe infection</td>
<td></td>
</tr>
<tr>
<td>K17 (24)</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>100</td>
<td>200</td>
<td>—</td>
<td>Mild, patchy infection</td>
<td></td>
</tr>
<tr>
<td>K19 (24)</td>
<td>100</td>
<td>102,400</td>
<td>1,600</td>
<td>1,600</td>
<td>200</td>
<td>2S</td>
<td>100</td>
<td>12,800</td>
<td>1,600</td>
<td>3,200</td>
<td>100</td>
<td>Mild, patchy infection</td>
<td></td>
</tr>
<tr>
<td>K02 (37)</td>
<td>—</td>
<td>12,800</td>
<td>—</td>
<td>6,400</td>
<td>—</td>
<td>2S</td>
<td>—</td>
<td>51,200</td>
<td>200</td>
<td>25,600</td>
<td>—</td>
<td>Severe infection</td>
<td></td>
</tr>
<tr>
<td>K04 (37)</td>
<td>—</td>
<td>12,800</td>
<td>800</td>
<td>25,600</td>
<td>400</td>
<td>2S</td>
<td>102,400</td>
<td>3,200</td>
<td>51,200</td>
<td>800</td>
<td>—</td>
<td>Severe infection</td>
<td></td>
</tr>
</tbody>
</table>

*a* Days in captivity before initial blood sample was collected.  
*b* Initial serology was conducted after 22–37 days of captivity.  
*c* Dashes indicate no titer identified at time of sampling.
serovars Grippotyphosa and Canicola (Richardson and Gauthier, 2003). Similarly, *L. interrogans* serovar Grippotyphosa has been isolated from the kidneys of five southern flying squirrels (*Glaucomys volans*) and was subsequently identified in two human handlers, demonstrating the potential for zoonotic transmission (Masuzawa et al., 2006).

We provide evidence of *L. interrogans* infection, with both mild and severe renal disease, in a population of peridomestic fox squirrels in Larimer County, Colorado, USA. The high prevalence of disease (45%) and antibody-positive animals (55%) was unexpected and suggests that fox squirrels may be important in the epidemiology of leptospirosis in the region. Although the causative serovar cannot be unequivocally determined, the high antibody titers suggest probable infection with serovars Grippotyphosa, Hardjo, or Icterohaemorrhagiae. Many species of rodents are considered reservoirs of *L. interrogans*; therefore, squirrels may be important hosts in the maintenance and transmission of *L. interrogans* in some regions. Although the course of infection with *L. interrogans* is unknown, the squirrels in this study were captured in urban and suburban areas, and the epidemiology of leptospirosis in fox squirrels and increased incidence in dogs may be related. The possibility of squirrel-dog transmission warrants further investigation to better understand the spread of leptospirosis at the urban-wildlife interface.

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**LITERATURE CITED**


