Widespread detection of antibodies to *Leptospira* in feral swine in the United States

K. Pedersen  
*U.S. Department of Agriculture, Animal and Plant Health Inspection Service*, Kerri.Pedersen@aphis.usda.gov

K. L. Pabilonia  
*Colorado State University*, kristy.pabilonia@colostate.edu

T. D. Anderson  
*Colorado State University*

S. N. Bevins  
*U.S. Department of Agriculture, Animal and Plant Health Inspection Service*

C. R. Hicks  
*U.S. Department of Agriculture, Animal and Plant Health Inspection Service*

*See next page for additional authors*

Follow this and additional works at: [http://digitalcommons.unl.edu/icwdm_usdanwrc](http://digitalcommons.unl.edu/icwdm_usdanwrc)

Part of the [Large or Food Animal and Equine Medicine Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc), [Life Sciences Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc), [Other Veterinary Medicine Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc), [Veterinary Infectious Diseases Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc), [Veterinary Microbiology and Immunobiology Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc), and the [Veterinary Preventive Medicine, Epidemiology, and Public Health Commons](http://digitalcommons.unl.edu/icwdm_usdanwrc)

[http://digitalcommons.unl.edu/icwdm_usdanwrc/1562](http://digitalcommons.unl.edu/icwdm_usdanwrc/1562)
Authors
Widespread detection of antibodies to *Leptospira* in feral swine in the United States

K. PEDERSEN¹*, K. L. PABILONIA², T. D. ANDERSON², S. N. BEVINS³, C. R. HICKS⁴, J. M. KLOFT⁵ AND T. J. DELIBERTO³

¹ U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, Fort Collins, CO, USA
² Colorado State University, Department of Microbiology, Immunology and Pathology, College of Veterinary Medicine and Biomedical Sciences, Fort Collins, CO, USA
³ U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, National Wildlife Research Center, Fort Collins, CO, USA
⁴ U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, Groveport, OH, USA
⁵ U.S. Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services, Manhattan, KS, USA

Received 14 May 2014; Final revision 31 October 2014; Accepted 31 October 2014; first published online 18 December 2014

SUMMARY

As feral swine continue to expand their geographical range and distribution across the United States, their involvement in crop damage, livestock predation, and pathogen transmission is likely to increase. Despite the relatively recent discovery of feral swine involvement in the aetiology of a variety of pathogens, their propensity to transmit and carry a wide variety of pathogens is disconcerting. We examined sera from 2055 feral swine for antibody presence to six serovars of *Leptospira* that can also infect humans, livestock or domestic animals. About 13% of all samples tested positive for at least one serovar, suggesting that *Leptospira* infection is common in feral swine. Further studies to identify the proportion of actively infected animals are needed to more fully understand the risk they pose.

Key words: Disease, feral swine, *Leptospira*, leptospirosis, *Sus scrofa*.

INTRODUCTION

Leptospirosis is a disease of global importance and is one of the most widespread zoonoses worldwide [1], infecting most mammalian species. Leptospirosis is caused by bacterial spirochaetes and includes both saprophytic and pathogenic species belonging to the genus *Leptospira*. The bacteria are classified genetically into 19 species, [2], with at least 12 of these being pathogenic and including more than 250 pathogenic serovars. Most serovars are host adapted, but the leptospires are harboured in the kidney and can serve as a source of infection for other animals or humans [1].

Human infection typically occurs after direct exposure to contaminated animal urine, or indirectly through contaminated water, when it comes in contact with the skin, eyes, or mucous membranes [1]. Severity of the disease depends on the age and health of the person infected. Symptoms typically include fever, chills, and intense headache, although some infections
including jaundice, acute renal and hepatic failure, pulmonary distress and haemorrhage can result in death [2, 3]. In the continental United States, cases are usually linked to people who are infected through occupational exposure (i.e. slaughter workers, farmers, etc.) [4], and to residents of Hawaii, where *Leptospira* thrive due to the tropical and subtropical climate [5]. However, over the last 20 years, outdoor enthusiasts have more commonly been the victims of infection with the pathogen [4, 6, 7] due to recreational exposure to contaminated water. Leptospirosis is now classified as a re-emerging zoonotic infection because of this increase in the number of human outbreaks over the last decade [8] resulting in increased attention worldwide.

Although originally introduced into the United States in the 16th century, feral swine (*Sus scrofa*) have markedly increased their range and population size over the last 20 years [9]. This is due to a number of factors including natural range expansion, accidental escape or intentional release, translocation into new areas specifically for hunting purposes, and their adaptability to a wide range of habitats [10]. As this expansion continues across the country, and recreational hunting and wild pig meat consumption become more popular, the possibility of direct contact between feral swine and humans will increase, and could result in increased opportunities for disease transmission [11]. The rate of expansion is disconcerting since feral swine can serve as reservoirs for a number of diseases of agricultural and zoonotic concern [12, 13]. Leptospirosis, in particular, is a problem because it can infect both humans and livestock, and can survive in moist soil and freshwater environments for long periods [2]. In domestic swine *Leptospira* is an important cause of reproductive failure, most frequently with serovar Bratislava [14]. Whether or not this pathogen causes similar reproductive effects in swine is unknown, but feral swine populations have continued to increase in both population size and geographical range which suggests that if there is reproductive failure associated with *Leptospira* in feral swine, it is not having a population-level effect. Although *Leptospira* antibodies have been detected in feral swine in the United States previously, these studies were focused on specific geographical areas [15–17]. We are unaware of a nationwide comprehensive effort to determine the geographical distribution and apparent prevalence of this pathogen in the United States. In order to provide insight into this disease system, we tested feral swine sera from across the United States to characterize the antibody prevalence of feral swine to six serovars of *Leptospira* of agricultural or zoonotic concern (Bratislava, Canicola, Grippotyphosa, Hardjo, Icterohaemorrhagiae, Pomona).

**METHODS**

**Sample collection**

Samples are routinely collected for disease surveillance from feral swine removed for wildlife damage management purposes by the U.S. Department of Agriculture’s Wildlife Services (WS), across the United States. Although samples are tested for various diseases at the time of collection, additional serum samples are stored at −80 °C as part of a tissue archive operated by the WS’ National Wildlife Disease Program.

For this study, 2055 feral swine serum samples collected between February 2007 and June 2011 were selected from the archive. Counties with 10 or more samples available in the archive were prioritized, resulting in at least one county (107 counties total) per state for 28 states (Supplementary Table S1). The samples were comprised of 1091 females, 953 males, and 11 of unspecified sex (Table 1).

**Sample testing**

All samples were tested at Colorado State University with the microscopic agglutination test (MAT) [18] to detect antibodies against six *Leptospira* serovars: Hardjo, Icterohaemorrhagiae, Canicola, Grippotyphosa, Pomona, and Bratislava. A titre of ≥1:200 was considered positive. MAT results were reported as the endpoint dilution of serum where 50% agglutination of cells was observed (1:50, 1:100, 1:200, 1:400, 1:800, 1:1600).

### Table 1. Number of feral swine tested by sex and test result for exposure to *Leptospira* by microscopic agglutination testing (MAT)

<table>
<thead>
<tr>
<th>Sex†</th>
<th>No. tested*</th>
<th>No. positive</th>
<th>% positive (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Males</td>
<td>953</td>
<td>138</td>
<td>14.5% (12.3–16.9)</td>
</tr>
<tr>
<td>Females</td>
<td>1091</td>
<td>131</td>
<td>12.0% (10.1–14.1)</td>
</tr>
</tbody>
</table>

† Eleven additional pigs of unspecified gender were negative.
1:1600, 1:3200, 1:>6400). Titres >1:6400 were not measured to their endpoint since titres ≥1:800 were considered evidence of recent or current infection [19]. The serovar in which agglutination was detected at the highest dilution was considered the infective serogroup, which for some samples included multiple serovars.

**Statistical analysis**

Mean Leptospirosis exposure and associated 95% confidence intervals for the different sexes were calculated using a binomial distribution. Potential disease associated risk factors were analysed using a mixed model (Proc Glimmix) and run using SAS version 9.2 [20]. The analysis was conducted using a logistic link function and binary error. Leptospira exposure (positive or negative) was the dependent variable. The primary regression factor of interest (α = 0.10) was the sex of sample animals.

**RESULTS**

Of 2055 samples tested, 269 (13.1%) were positive for at least one serovar and 97 (36%) of these were infected with multiple serovars. Co-infections with Pomona and Bratislava were more common than any other serovar combination. Samples tested positive for at least one serovar in 20 of 28 states and 71% of 107 counties represented in this study (Supplementary Table S1).

Sex was significantly associated with exposure (F = 2.78, P = 0.095), with slightly more males having been exposed than females (OR 1.28, Table 1).

Pomona was the most common serovar identified followed by Bratislava, Grippotyphosa, Icterohaemorrhagiae, Hardjo and Canicola, respectively (Table 2). Current or recent infections (titres of ≥800) were identified most commonly as Pomona, followed by Bratislava and Grippotyphosa (Table 2).

**DISCUSSION**

We determined that exposure to Leptospira is common in feral swine, and is not limited to certain regions of the country since antibody-positive feral swine were identified in 70% of the states included in this study (Fig. 1). Although no definitive patterns of distribution were identified based on the samples available, it is clear that feral swine, regardless of location, are exposed to pathogenic Leptospira and thus may be involved in the epidemiology of the pathogen. Domestic pigs can harbour the spirochaete in the kidneys and intermittently shed the organism [11] suggesting that feral swine could do the same.

Feral swine typically use wet areas for wallowing and rooting which may result in contamination of the habitat [21]. Feral swine root in agricultural fields and utilize artificial water sources such as stock tanks or those created by irrigation [22] which may perpetuate transmission. The high rate of Leptospira exposure observed in feral swine relative to other pathogens [22], may be a result of their propensity to utilize habitats which may have been contaminated by other wildlife species such as raccoons (Procyon lotor), squirrels (family Sciuridae) or rats (genus Rattus) [19]. Although transmission most likely occurs through these indirect routes, direct transmission may also occur through direct contact with infected urine, placental fluids, or milk and through venereal or placental transmission. Swine may also serve as a reservoir and subsequent source of environmental contamination in formerly pathogen-free areas. As feral swine continue to expand their geographical range throughout the United States, it will be important to understand their aetiological role, if any, in leptospirosis outbreaks. This may be especially important in areas where freshwater-based activities, such as swimming, kayaking, or adventure races are popular, consequently increasing the risk for zoonotic transmission of this organism.

### Table 2. Antibody titres to six serovars of Leptospira detected in feral swine in the United States

<table>
<thead>
<tr>
<th>Serovar*</th>
<th>Positive (n)</th>
<th>1:200</th>
<th>1:400</th>
<th>1:800</th>
<th>1:1600</th>
<th>1:3200</th>
<th>1:&gt;6400</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bratislava</td>
<td>106</td>
<td>50</td>
<td>25</td>
<td>16</td>
<td>6</td>
<td>8</td>
<td>1</td>
</tr>
<tr>
<td>Canicola</td>
<td>12</td>
<td>10</td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Grippotyphosa</td>
<td>75</td>
<td>29</td>
<td>15</td>
<td>18</td>
<td>8</td>
<td>3</td>
<td>2</td>
</tr>
<tr>
<td>Hardjo</td>
<td>32</td>
<td>16</td>
<td>13</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Icterohaemorrhagiae</td>
<td>59</td>
<td>32</td>
<td>18</td>
<td>8</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pomona</td>
<td>124</td>
<td>39</td>
<td>26</td>
<td>22</td>
<td>12</td>
<td>10</td>
<td>15</td>
</tr>
</tbody>
</table>

* Some feral swine were positive for multiple serovars.
We detected a significant difference in antibody prevalence between males and females. Feral swine social groups, called sounders, are comprised of sows and their offspring. Adult males are solitary [23], after they disperse from the natal social group. As a result, males occupy larger home ranges than females [24]. This transient social behaviour may explain why males were more frequently exposed to *Leptospira*. This distinction between sexes (Table 1) was not replicated in other studies [25, 26], but may have been merely a function of sample size since we sampled at least five times the number of animals as these other studies.

Our data suggest that males may be important contributors in the transmission cycle of *Leptospira*; their propensity to occupy larger home ranges compared to females may lead to increased opportunities for both contraction and dispersal of the pathogen via indirect contact with contaminated environments, or directly through contact with infected animals. Surveillance efforts may be most cost effective by targeting males rather than females that may be less likely to be involved in transmission.

Pomona is the primary serovar that infects domestic swine [27]; therefore it is not surprising that it was also the most common serovar that we detected in feral swine. This finding is consistent with previous studies [25, 28]; however, these high rates of exposure to Pomona are alarming since this serovar is associated with infections in humans [27] and domestic swine [29]. Similarly, the high prevalence of Bratislava was consistent with other studies [19] that have identified the serovar frequently in domestic swine [14, 29].

Cross-reaction of serovars is a known limitation of serological testing [30] and cross-reactivity with Pomona in particular has been well documented [31]. For the purposes of this study, the serovar with the highest antibody titre was determined to be the infecting serovar per individual animal. This design was used to account for the potential for cross-reactivity. However, this does not take into account the potential for individual feral swine to have antibodies to more than one serovar, indicating the potential for multiple exposures to multiple *Leptospira* during the life of the feral swine. In addition, although identification of a certain serovar may suggest a
specific animal source, because of the strong associations of some serovars with particular species, cross-reactivity between serovars may prevent pinpointing a specific source and also may prevent a robust diagnosis [4]. Although cross-reactivity likely occurred in these data since 36% of the positives had multiple serovars with titres $\geq$ 200, it is clear that *Leptospira* exposure is widespread in feral swine and that they serve as reservoir hosts for Pomona and Bratislava similar to domestic swine. Infections with serovar Bratislava in domestic swine tend to occur at low titres (i.e. titres <200) [29], and if this same pattern holds true in feral swine it is possible that many of the feral swine we identified as Bratislava-negative were actually infected with this serovar, but below the cutoff value of 200 that we used in this study.

As feral swine populations continue to expand into urban areas and their ranges overlap with domestic swine and human activities, transmission of pathogens such as *Leptospira* will become of increasing concern. Since widespread exposure was documented during this study, we recommend that further studies be conducted to more fully understand the role of feral swine in shedding this pathogen.

SUPPLEMENTARY MATERIAL
For supplementary material accompanying this paper visit http://dx.doi.org/10.1017/S0950268814003148.

ACKNOWLEDGEMENTS
We thank all of the wildlife biologists and technicians who participated by collecting the samples included in this paper and spending many hours trapping feral swine and preparing samples for testing. We also thank Cindy Hirota and Kathryn Brown who provided technical support for this project. Mention of trade names or commercial products in this work is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture.

DECLARATION OF INTEREST
None.

REFERENCES