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Development of serum-free media in CHO-DG44 cells using a central composite statistical design

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Abstract A serum free medium was developed for the production of recombinant antibody against Botulinum A (BoNTA) using dihydrofolate reductase deficient Chinese Hamster Ovary Cells (CHO-DG44) in suspension culture. An initial control basal medium was prepared, which was similar in composition to HAM's F12: IMDM (1:1) supplemented with insulin, transferrin, selenium and a lipid mixture. The vitamin concentration of the basal medium was twice that of HAM's F12: IMDM (1:1). CHO-DG44 cells expressing S25 antibody grew from 2×10^5 cells to maximum cell density of 1.04×10^6 cells/ml after 5 days in this control medium. A central composite design was used to identify optimal levels and interaction among five groups of medium components. These five groups were glutamine, Essential Amino Acids (EAA), Non

Essential Amino Acids (NEAA), Insulin, Transferrin, Selenium (ITS), and lipids. Fifty experiments were carried out in four batches, with two controls in each batch. There was little effect of ITS and Lipid concentrations over the range studied, and glutamine concentration showed a strong interaction with EAA. The optimal concentrations of the variables studied were 2.5 mM Glutamine, 7.4 mM (2×) EAA, 1.4 mM (0.5×) NEAA, 1× ITS supplement, 0.7× Lipids supplement. The maximum viable cell density attained in the optimized medium was 1.4×10^6 cells/ml, a 35% improvement over the control culture, while the final antibody titer attained was 22 ± 3.4 µg/mL, a 50% improvement.

Keywords Response surface method · Media optimization

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Introduction

Botulinum neurotoxin, produced by the anaerobic *Clostridium botulinum*, is one of the most potent toxins known to humans (Franz et al. 1997; Gill 1982). *Clostridium botulinum* produces seven antigenically distinct neurotoxins (A–G) differentiated serologically by specific neutralization (Hatheway 1990). The active neurotoxin is synthesized as two polypeptide chains, a heavy chain (100 KDa) and a light chain (50 KDa), connected via disulfide linkage (DasGupta and Sathyamoorthy 1984; Syuto and

Kubo 1981). Recombinant monoclonal antibodies (mAb) can neutralize the effects of BoNT A without requiring human donors for plasmapheresis (Lang et al. 1993). Potent neutralizing monoclonal antibodies were identified recently, characterized, cloned and expressed in Chinese Hamster Ovary (CHO) cells to yield humanized mAbs (Nowakowski et al. 2002).

The large-scale, commercial production of therapeutically important proteins from rCHO cells typically involves a suspension culture-based manufacturing process (Sinacore et al. 1996). It is desirable to use serum-free medium in suspension culture because serum can cause problems in subsequent processes (Glassy et al. 1988; Keen and Rapson 1995). However, there is no universal serum-free medium applicable to all cell lines. There is need to develop specific medium suitable for each cell line (Hayter et al. 1991; Zang et al. 1995).

The medium used for animal cell culture is very complex. Further, as the impact of medium components on cell growth or product synthesis is rather difficult to fully elucidate, statistical methods are adopted to develop a medium for cell culture that offers optimal viable cell density and product formation. The traditional one-factor-at-a-time approach to optimization is time-consuming and incapable of reaching the true optimum especially because of interactions among the factors that influence the growth process. Moreover, this approach assumes that the various growth parameters do not interact and the process response is a direct function of a single varied parameter (Castro et al. 1992; Freshney 1994). In reality, the observed behavior of growth results from the interactive influences of the various variables (Chen et al. 2002). To be effective, optimization requires statistical methods that take these interactions into account.

Response surface methodology (RSM), an experimental strategy for seeking the optimum conditions for a multivariable system, is a much more efficient technique for optimization (Box et al. 1978). RSM comprises of mathematical and statistical procedures that can be used to study relationships between one or more responses and a number of independent variables. In addition to analyzing the effects of independent variables, this experimental methodology generates a mathematical model that accurately describes the overall process (Myers and Montgomery 1995; Senanayake and Shahidi 2002). RSM has

been employed to solve multivariate problems and optimize responses in many types of experiments (Maddox and Richert 1977; Giovanni 1983; Houg et al. 1989). In this approach, concentrations of medium components are the variables; each variable refers to some base value and varies in a certain pattern. This pattern is designed by using statistical methods to yield the most information by a minimum number of experiments.

In this study, we have adopted a RSM approach to locate the optimum levels of Glutamine, Essential Amino Acids (EAA), Non-Essential Amino Acids (NEAA), Insulin, Transferrin, Selenium (ITS), and Lipids. Our objective is to gain insight into the interactions among these factors that significantly impact the response and viable cell density. Glutamine acts as the primary source of nitrogen as well as an additional carbon and energy source. It contributes precursors to the formation of the major intracellular binding blocks: amino acids, proteins. Approximately 30–65% of the cell energy requirement is derived from glutamine metabolism (Zielke et al. 1984; Reitzer et al. 1979). Amino acids are primary sources of nitrogen that protect cells from nutrient deprivation (Franek and Sramkova 1996), elevated osmolarity (Øyaas et al. 1994) and elevated pCO₂ (de Zengotitia et al. 2002). Insulin serves as a growth and maintenance factor and is considered to be important for serum-free cultures (Schubert 1979). Insulin stimulates uridine and glucose uptake and synthesis of RNA, proteins and lipids; it also increases fatty acid and glycogen synthesis (Mather and Sato 1979). Transferrin is one of the most essential growth-promoting supplements in serum-free medium, and its omission causes severe inhibition of cell growth (Kovar and Franek 1984). Transferrin is an iron binding glycoprotein that interacts with surface receptors. It is closely related to the transport of iron across the plasma membrane (Bretscher 1985). Transferrin has additional in-vitro functions, e.g., chelation of deleterious trace materials, that are unlikely replaced by other components. Selenium is a trace element essential for mammalian cell cultures (Nielsen et al. 1981); its mechanism is poorly understood. There is evidence that selenium enhances growth rate in serum free-cultures (Darfler and Insel 1983). Lipids are required for proliferation, differentiation, and antibody secretion. They play a major role in the cell membrane which is composed of a

phospholipid bilayer, and help in the transmission of nutrients into the cell and excretion of proteins out the cell (Farrant et al. 1984). The major functions exhibited by the variables motivated us to choose the response surface methodology in order to observe the higher order interactions that would maximize the response. Results were analyzed statistically by SAS and optimum conditions were selected graphically. Interactions among these factors were also examined.

Materials and methods

Cell line and medium

CHO-DG44 cells, which are dhfr negative, were obtained as a gift from Dr. Larry Chasin (Columbia University). The parental cell line was obtained by weaning CHO-DG44 from the serum according to standard cell culture techniques. The base medium used during the weaning process was a commercial serum-free medium, CHO-S-SFMII, known to contain animal-derived proteins and hydrolysates. The process of weaning CHO-DG44 of its serum dependence lasted approximately 4–5 months. The resulting cell line was used as the starting point for all subsequent development efforts including recombinant cell line generation, medium development studies. The recombinant cell line was constructed by inserting the chimeric light and heavy chain IgG genes against BoNT serotype A, along with the gene

for dhfr into the plasmid pcDNA3.1(+) (Invitrogen, Carlsbad/CA) and the procedures are detailed elsewhere (Mowry et al. 2004). Recombinant cell lines were derived from our parent cell line using standard molecular biology techniques.

Medium

The basal medium was prepared similar to the HAM's F12:IMDM (1:1) medium excluding Hypoxanthine and Thymidine, by adding components separately. The concentration of the inorganic salts and other components such as linoleic acid, lipoic acid, phenol red, putrescine 2HCl, sodium pyruvate, and HEPES is the same as with HAM's F12:IMDM (1:1). The concentrations of the glucose and glutamine in the starting basal medium were 4 g/L and 4 mM, respectively. For amino acids the medium was supplemented with 1.75× of EAA and 1.75× of NEAA, which come from Gibco as 50× and 100× solutions respectively. The concentrations of individual amino acids in EAA (50×) and NEAA (100×) solutions are given in Table 1. The additional components added to the basal medium are vitamins, the concentrations of which were double those in HAM's F12:IMDM (1:1), 1× (ITS), and 0.7× Lipids Supplement, which come as 100× solutions from Gibco. The concentration of the individual components in ITS and Lipids Supplements are given in the Table 1. The composition of the complete basal medium is given in Table 2.

Table 1 Composition of the individual components in mg/L in the solutions EAA, NEAA, ITS, Lipids, concentrated solutions from Gibco

(50×) EAA [mg/L]	(100×) NEAA [mg/L]	(100×) ITS [mg/L]	(100×) Lipids [mg/L]
L-Arginine [6320]	L-Alanine [890]	Insulin [1000]	Arachidonic acid [2]
L-Cystine[1200]	L-Asparagine[1320]	Transferrin [0.67]	Cholesterol [220]
L-Histidine × HCl × H ₂ O [2100]	L-Aspartic acid [1330]	Selenium [0.55]	DL-alpha-tocopherol-acetate [70]
L-Isoleucine [2620]	L-Glutamic acid [1470]		Linoleic acid [10]
L-Leucine [2620]	Glycine [750]		Linolenic acid [10]
L-Lysine HCl [3625]	L-Proline [1150]		Myristic acid [10]
L-Methionine [755]	L-serine [1050]		Oleic acid [10]
L-Phenylalanine [1650]			Palmitic acid [10]
L-Threonine [2380]			Pluronic [0.1%]
L-Tryptophan [510]			Stearic acid [10]
L-Tyrosine [1800]			Tween 80 [2200]
L-Valine [2340]			

Table 2 Concentrations of the components in the control medium

Components	Composition [mg/mL]	Components (contd.)	Composition [mg/mL]
CaCl ₂ (anhyd.)	99.1	L-Phenylalanine	57.7
CuSO ₄ × 5H ₂ O	0.00125	L-Proline	20.12
FeSO ₄ × 7H ₂ O	0.415	L-Serine	18.37
KCl	276.8	L-Threonine	83.3
MgCl ₂ (anhyd.)	28.61	L-Tryptophan	17.8
NaCl	5500	L-Tyrosine × 2Na × 2H ₂ O	63
NaHCO ₃	2100	L-Valine	81.9
Na ₂ HPO ₄ (anhyd.)	71	Biotin	0.02
Na ₂ HPO ₄ × H ₂ O	62.5	D-Ca Pantothenate	4
ZnSO ₄ × 7H ₂ O	0.43	Choline chloride	18
KNO ₃	0.038	Folic acid	4
MgSO ₄ (anhyd.)	50.8	i-Inositol	25.2
Na ₂ SeO ₃	0.0085	Niacinamide	4
D-glucose	4000	Pyridoxine HCl	4
Linoleic acid	0.04	Riboflavin	0.4
Lipoic acid	0.105	Thiamine HCl	4
Phenol Red	8.1	Vitamin B12	1.413
Putrescine 2HCl	0.0805	Insulin	10
Sodium pyruvate	110	Transferrin	5.5
HEPES	2979	Sodium selenite	0.0134
L-Alanine	15.57	Arachidonic acid	0.014
L-Arginine × HCl	221.2	Cholesterol	1.54
L-Asparagine × H ₂ O	23.1	DL-alpha-tocopherol-acetate	0.49
L-Aspartic acid	23.27	Linoleic acid	0.07
L-Cystine × 2HCl	42	Linolenic acid	0.07
L-Glutamic acid	25.72	Myristic acid	0.07
L-Glutamine	584	Oleic acid	0.07
Glycine	13.12	Palitoelic acid	0.07
L-Histidine × HCl × H ₂ O	73.5	Palmitic acid	0.07
L-Isoleucine	91.7	Stearic acid	0.07
L-Leucine	91.7	Tween 80	15.4
L-Lysine × HCl	126.88	Pluronic	0.10%

Cell culture

The cell cultivation was performed in 37°C humidified incubators supplemented with 5% carbon dioxide. The seeding density was 2×10^5 cells/mL, and cell counts were performed every 4 days. The number of cells were determined using a hemocytometer. Spheroids would be enzymatically dissociated when spherical aggregates were observed. Approximately, 1.7 mL of sample were harvested from spinners and placed in a 2.0 mL microtube and centrifuged at 1,200 rpm for 6 min; 1.5 mL supernatant was saved

for antibody assays. Hundred microliters (μ L) of trypsin solution (2.5% (w/v) Trypsin in PBS) was added to resuspend the cells of 400 μ L. The cells were incubated at room temperature for 15 min and the cell density and viability were then determined by the trypan blue exclusion method.

Antibody assay

The concentration of the whole antibody, as well as the concentration of the light and heavy chain portions, was determined by an enzyme-linked

immunosorbent assay. Affinity purified rabbit anti-human IgG antibodies were diluted to 5 µg/mL in a coating buffer consisting of 100 mM NaHCO₃ and 100 mM NaCl (pH = 9.3). About 100 µL diluted antibody was added to 96 well plates (Nunc) and incubated overnight at 4°C. The plates were washed twice in a Tris buffer (20 mM Tris–HCl, 50 mM NaCl, pH = 7.2) containing 0.1% Tween 20, and washed twice in the Tris buffer alone. Blocking buffer (Tris buffer with 0.5% BSA) was added to the 96 well plates and incubated at 37°C for 1 h. Supernatant samples were diluted in the blocking buffer and samples were loaded into the 96 well plates in triplicate. Plates were incubated for 1 h at 37°C and the washing procedure was repeated. Hundred microliters (µL) of a goat anti-human IgG–HRP conjugate antibody diluted to 0.5–2 mg/mL in the dilution buffer was added to the plates. The plates were incubated for 1 h at 37°C and the washing procedure was repeated. Finally, 100 µL of 50 µg/mL ABTS in ABTS buffer (Roche) was added to the plates. The absorbance was determined at 405 nm using an ELx800 plate reader (Bio-Tek). This procedure was used for the whole antibody, the heavy chain (Fc specific), and the light chain (κ specific). In sandwich ELISA assays, rabbit anti-human IgG antibodies raised against the whole molecule, Fab-specific and kappa-chain specific, respectively, were used to coat the ELISA plates. HRP-conjugated rabbit anti-human IgG antibodies raised against the whole molecule, Fab-specific and kappa-chain specific, respectively, were used in the detection step of the ELISA assays.

Experimental design

Response surface methodology (RSM) was used to determine the influence of some medium components on the response of viable cell density. Our assumption is that since our product is growth-associated, increase in viable cell density will ultimately increase the antibody production. Theoretical and fundamental aspects of RSM have been extensively discussed elsewhere (Myers and Montgomery 1995). The experimental design adopted Box's central composite design for five variables at five levels each. The five independent variables were X1 = Glutamine, X2 = EAA, X3 = NEAA, X4 = ITS, X5 = Lipids. The independent variable coded regions were $-\alpha$ (–2,

Table 3 Actual factor levels corresponding to coded factor levels

Level	Symbol	Actual factor level at coded factor level of				
		–2	–1	0	1	2
Glutamine	X1	1 mM	2.5 mM	4 mM	5.5 mM	7 mM
EAA	X2	0×	0.5×	1×	1.5×	2×
NEAA	X3	0.25×	1×	1.75×	2.5×	3.25×
ITS	X4	0.25×	1×	1.75×	2.5×	3.25×
LIPIDS	X5	0.1×	0.4×	0.7×	1×	1.3×

Lowest Level), –1, 0 (middle level), 1, and $+\alpha$ (2, highest level). The actual values, which were chosen from preliminary studies, and the corresponding coded and uncoded values of the five independent variables are given in Table 3. The complete design has 42 experimental points, which including eight replications of the center point. The treatment combinations and observed responses are presented in Table 4. The 50 experimental medium runs were prepared in random order and the experiments were performed in four batches. The dependent variable (Y) was viable cell density and was assumed to be affected by the five independent variables. Based on data from this design, we fit a second order or higher order polynomial regression model described as follows:

$$Y = b_l + \sum_{i=1}^k b_i X_i + \sum_{i=1}^k b_{ii} X_i^2 + \sum_i \sum_j b_{ij} X_i X_j + \epsilon$$

where

Y = Viable cell density

b_l = Intercept for block l

X = Factors (X_1 = Glutamine, X_2 = EAA, X_3 = NEAA, X_4 = ITS, X_5 = Lipids)

b_z = Regression coefficient ($z = i, ii$ or ij , where $i < j$)

ϵ = Residual error

$k = 1, 2, 3, \dots$

Statistical analysis

Using ordinary least squares the regression model was fit to evaluate, the explanatory variables regarding linear, interaction, and quadratic effects of coded

Table 4 Treatment combinations with variables in coded values and the values of response

Run	X1	X2	X3	X4	X5	Y
1	1	1	-1	-1	1	8.1E + 05
2	1	1	-1	1	-1	7.2E + 05
3	-1	-1	-1	-1	-1	6.5E + 05
4	0	0	0	2	0	8.3E + 05
5	-1	1	1	-1	-1	1.0E + 06
6	1	-1	1	-1	1	8.0E + 05
7	2	0	0	0	0	7.3E + 05
8	0	0	0	0	2	7.1E + 05
9	1	1	-1	1	1	5.4E + 05
10	0	0	0	0	0	9.5E + 05
11	1	-1	-1	1	1	6.6E + 05
12	0	0	0	0	0	9.3E + 05
13	-1	-1	1	1	1	8.40E + 05
14	1	-1	1	1	-1	8.80E + 05
15	1	-1	1	1	1	6.25E + 05
16	-1	-1	1	-1	-1	7.00E + 05
17	0	0	0	0	0	1.18E + 06
18	-1	-1	-1	1	1	8.15E + 05
19	0	0	0	0	-2	1.15E + 06
20	1	-1	-1	1	-1	6.55E + 05
21	0	0	0	0	0	1.16E + 06
22	-1	-1	-1	-1	1	2.00E + 05
23	-2	0	0	0	0	1.52E + 06
24	0	0	0	-2	0	5.15E + 05
25	0	-2	0	0	0	3.10E + 05
26	-1	-1	1	1	-1	8.20E + 05
27	1	-1	-1	-1	1	5.10E + 05
28	1	1	1	-1	-1	8.10E + 05
29	-1	1	-1	-1	-1	7.85E + 05
30	0	0	0	0	0	1.18E + 06
31	-1	-1	-1	1	-1	8.30E + 05
32	0	0	2	0	0	9.55E + 05
33	-1	1	1	-1	1	1.29E + 06
34	0	0	-2	0	0	1.80E + 05
35	1	1	-1	-1	-1	2.00E + 05
36	1	1	1	-1	1	2.20E + 05
37	0	0	0	0	0	1.21E + 06
38	-1	1	1	1	1	1.03E + 06
39	-1	1	-1	1	-1	9.00E + 05
40	1	-1	1	-1	-1	9.85E + 05
41	-1	1	1	1	-1	8.65E + 05
42	1	-1	-1	-1	-1	8.75E + 05
43	1	1	1	1	-1	7.70E + 05

Table 4 continued

Run	X1	X2	X3	X4	X5	Y
44	1	1	1	1	1	1.08E + 06
45	0	0	0	0	0	9.90E + 05
46	-1	1	-1	-1	1	9.35E + 05
47	-1	1	-1	1	1	8.55E + 05
48	-1	-1	1	-1	1	1.00E + 06
49	0	2	0	0	0	8.90E + 05
50	0	0	0	0	0	9.85E + 05

Where $\alpha = '2'$, Response (Y) = Viable cell density in cells/mL, '-1', '0', '+1' are coded factorial levels

levels of Glutamine, EAA, NEAA, ITS, Lipids on cell density. The R^2 value was used to evaluate model sufficiency and the α -level was set as 5%, at which point every term in the selected model should be significant. The reduced model was evaluated using the R^2 . Lack of fit was used to attempt to find optimal conditions for all the variables maximizing the cell density. Canonical analysis was then used to evaluate the nature of the stationary point (maximum, minimum or saddle) and to find the ridge of steepest ascent. Further experiments were carried out in the direction of the maximum response along with alternate experiments where Glutamine was set to different coded levels from '0' to '-3', keeping EAA constant at coded level '2' and '4'. All statistical computations were done using SAS/STAT procedures, and optimum conditions were found through SAS data-step programming. Response surface plots were generated by SAS/GRAPH.

Results and discussion

Regression analysis revealed that linear (X_i) and quadratic effects ($X_i \times X_i$) were more significant than cross product interactions ($X_i \times X_j$), as based on the p -values obtained (Table 5). Among all independent variables, Glutamine (negative effect, X_1 : -2.79) and NEAA (positive effect, X_3 : 3.07) had the greatest effects on the cell density, while EAA showed an effect when combined with Glutamine ($X_2 \times X_1$). Among the pairwise interactions, EAA and Glutamine exhibited the greatest effect. Although NEAA squared ($X_3 \times X_3$) and NEAA (X_3) by itself were significant, they did not have a great effect when

Table 5 The *t*- and *p*- values of full model with *X*₁, *X*₂, *X*₃, *X*₄, *X*₅ as independent variables

Parameter	<i>t</i> -value	<i>p</i> -value
Intercept	13.75	<0.001
<i>X</i> ₁	−2.79	0.0092
<i>X</i> ₂	1.53	0.1376
<i>X</i> ₃	3.07	0.0046
<i>X</i> ₄	1.2	0.2404
<i>X</i> ₅	−0.81	0.425
<i>X</i> ₁ × <i>X</i> ₁	0.66	0.5167
<i>X</i> ₂ × <i>X</i> ₁	−2.1	0.1446
<i>X</i> ₂ × <i>X</i> ₂	−2.67	0.0124
<i>X</i> ₃ × <i>X</i> ₁	−0.27	0.7855
<i>X</i> ₃ × <i>X</i> ₂	−0.1	0.9206
<i>X</i> ₃ × <i>X</i> ₃	−2.86	0.0078
<i>X</i> ₄ × <i>X</i> ₁	0.31	0.758
<i>X</i> ₄ × <i>X</i> ₂	0.22	0.8304
<i>X</i> ₄ × <i>X</i> ₃	−0.74	0.4662
<i>X</i> ₄ × <i>X</i> ₄	−2.21	0.0352
<i>X</i> ₅ × <i>X</i> ₁	−0.8	0.4292
<i>X</i> ₅ × <i>X</i> ₂	1.3	0.2026
<i>X</i> ₅ × <i>X</i> ₃	0.25	0.8061
<i>X</i> ₅ × <i>X</i> ₄	0.2	0.8427
<i>X</i> ₅ × <i>X</i> ₅	−0.58	0.5675
Linear		0.0046
Quadratic		0.0055
Cross Product		0.6621

*R*² = 0.6339 for the total model

compared with the other variables, as judged by the *p*-value. The response surface plots were then plotted to see the effect of EAA and NEAA (Fig. 1), EAA

and Glutamine (Fig. 2), NEAA and Glutamine (Fig. 3) on the response which is the viable cell density (*Y*). ITS and Lipids were found to have no effect. The *R*² value for the total model is 0.6339. To simplify the model, the variables of ITS and Lipids were removed from the model and the data were re-analyzed using the reduced model. The polynomial regression model used for three variables was

$$Y = b_0 + b_1X_1 + b_2X_2 + b_3X_3 + b_{11}X_1^2 + b_{22}X_2^2 + b_{33}X_3^2 + b_{12}X_1X_2 + b_{13}X_1X_3 + b_{23}X_2X_3 + \epsilon$$

When the variables ITS and Lipids were kept constant, the lack of fit is found to be insignificant, suggesting that the model was adequate to explain the effect of these three variables on the response. The *R*² value of the reduced model decreases to 0.502, as denoted in Table 6. As Table 6 shows, the Glutamine (*X*₁) had significant linear effects and interacted with EAA (*X*₂). EAA and NEAA had significant quadratic effects (*X*₂ × *X*₂ and *X*₃ × *X*₃) while NEAA (*X*₃) also had a linear effect on the cell density.

Canonical analysis

Canonical analysis is a mathematical approach used to examine the overall shape of the response surface and to determine if the estimated response point is a maximum, minimum or a saddle point. If the stationary point is maximum or minimum, a corresponding increase or decrease will result in the response. In the case of a saddle point, the response may increase or decrease when we move away from

Fig. 1 Response surface plot showing the effect of EAA, NEAA, and their mutual effect on the response (viable cell density). Other variables were held at zero level

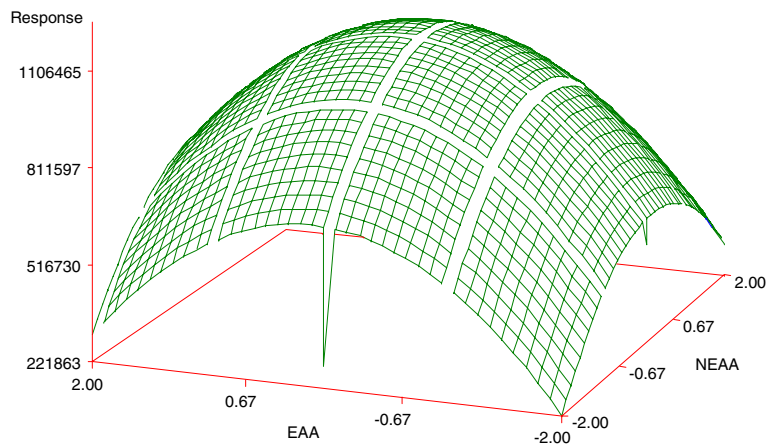


Fig. 2 Response surface plot showing the effect of EAA, glutamine and their mutual effect on the response (viable cell density). Other variables were held at zero level

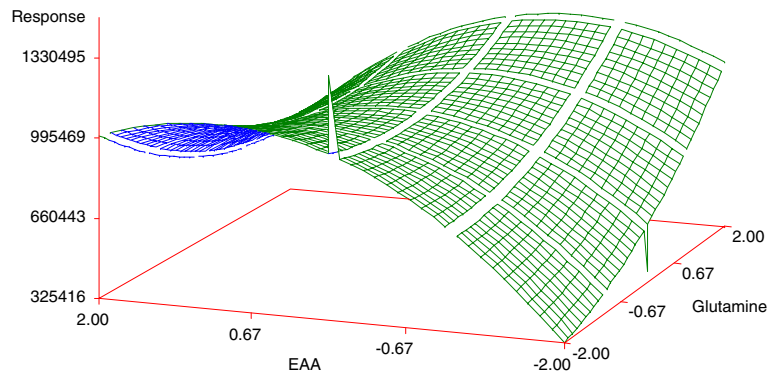


Fig. 3 Response surface plot showing the effect of NEAA, glutamine and their mutual effect on the response (viable cell density). Other variables were held at zero level

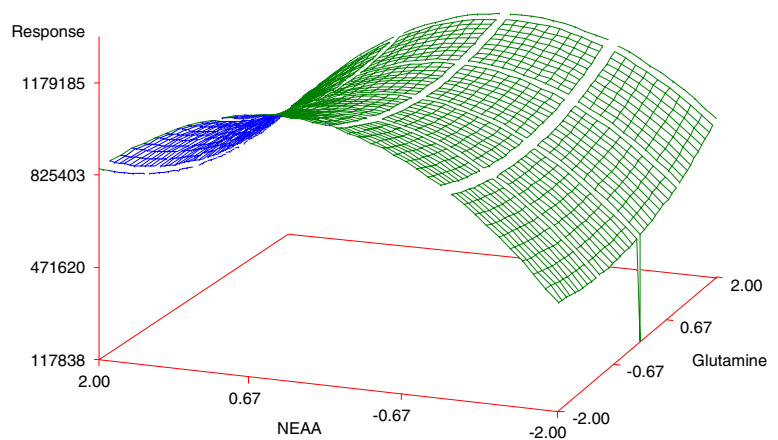


Table 6 The *t*- and *p*- values of the reduced model with X1, X2, X3 as independent variables

Parameter	<i>t</i> -value	<i>p</i> -value
Intercept	15.56	<0.0001
X1	-2.81	0.0076
X2	1.54	0.132
X3	3.1	0.0036
X1 × X1	0.66	0.5123
X2 × X1	-2.11	0.0407
X2 × X2	-2.69	0.0105
X3 × X1	-0.28	0.7835
X3 × X2	-0.1	0.9199
X3 × X3	-2.88	0.0064
Linear		0.001
Quadratic		0.0035
Cross product		0.2242

$R^2 = 0.5022$ for the reduced model

the stationary point, depending on which direction is taken. Maximizing the viable cell density is of interest; however the stationary point was a saddle point, so we move on the ridge in the direction to get the maximum response.

The points on the ridge that increased the response were found using the RIDGEMAX option of the SAS/RSREG procedure, and are shown in Table 7. From Tables 6 and 7, glutamine showed a negative effect on cell density while EAA, NEAA showed a positive effect. Therefore the glutamine values for the ridge moved in the negative direction and the values for EAA and NEAA moved in the positive direction. Following the ridge in Table 7, the highest cell density was $1.37E + 06$ cells/mL, but this prediction was not very reliable due to a large standard error obtained (144554). Based on the ridge analysis, the glutamine concentrations at high cell densities were found to decreasing, to a level of 1 mM or lower. Glutamine values smaller than 1 mM were thought to

Table 7 Ridge of steepest ascent for X1, X2, X3 independent variables, and estimated response and standard error

X1	X2	X3	Estimated cell density (10^5)	SE
-0.15	0.07	0.11	10.1	62346
-0.33	0.13	0.18	10.4	61381
-0.52	0.20	0.23	10.7	60421
-0.71	0.26	0.26	11.0	60558
-0.91	0.32	0.28	11.4	63278
-1.10	0.38	0.30	11.8	70025
-1.29	0.43	0.31	12.2	81626
-1.49	0.49	0.33	12.7	98153
-1.68	0.55	0.34	13.2	119266
-1.87	0.60	0.35	13.7	144554

be unreasonable and therefore additional experiments were conducted on the ridge below glutamine values of 1 mM.

To further explore the surface, we used the reduced model from Table 7, and obtained predicted cell densities at constant glutamine concentrations at different coded levels from '0' to '-3', NEAA at '0.5' and various values of EAA. The results are shown in Table 8. The results suggest that cell density increases as EAA (X2) increases when glutamine values are low. Figure 4 shows the effect of EAA and NEAA on VCD when glutamine is controlled at coded level '-1'.

EAA values up to a coded level of 14.5 (uncoded value = 12.625 X) are unfeasible because of osmotic effects or inhibition of metabolic pathways due to overfeeding the nutrients. According to the above results, however, it appears that with reduced Glutamine levels and concentrations of EAA and NEAA at a 0.5 coded level, large cell densities could possibly be obtained.

Table 8 Effect on EAA, NEAA and VCD when glutamine is controlled at different coded levels

X1(level)	X2(Level)	X3(Level)	VCD
-0.5	4.5	0.5	1.27E + 06
-1	6.5	0.5	1.56E + 06
-1.5	8.5	0.5	1.95E + 06
-2	10.5	0.5	2.43E + 06
-2.5	12.5	0.5	3.01E + 06
-3	14.5	0.5	3.69E + 06

Alternate experiments

To further evaluate the surface we ran some alternate experiments at different levels of glutamine (from coded level '0' to '-3'), keeping EAA constant at coded levels 2 and 4. We expected low cell growth at a glutamine value less than '-1.5 coded level' and no cell growth at zero ('-2' coded level) glutamine concentration. We also expected the EAA coded level of '2' to result in higher cell densities compared to EAA coded level '4,' due to osmotic effects and the inhibition of metabolic pathways from overfeeding. Therefore 14 additional experiments were conducted, four on the ridge, four at different levels of Glutamine keeping EAA at coded level '2,' four at EAA coded level '4,' and two controls (basal medium) as shown in Table 9.

These experiments were conducted under the same conditions as the initial experiments. The starting density of the cultures was 2×10^5 cells/mL, and the cells were allowed to adapt to the medium in four passages. The final viable cell densities were derived as an average of the third and the fourth passage, as shown in Table 9.

Medium-6 had a higher cell density compared to the controls (13 and 14), but the last passage of medium 3, 9, and 5 were nearly equal to the control medium, as shown in Fig. 5. Media which had results equivalent to, or better than medium-0, were carried out for one more passage (up to 8 days), to validate the data. The results are shown in Fig. 6. From passage 5, the viable cell density attained in medium 6 after 5 days of culture was about 1.6 times higher than the control.

Replicate experiments

To validate the above results, the cells in the control medium were taken out of a frozen state, and the experiment was repeated three times with control medium and medium-6. Cells were allowed to go for four passages and the final viable cell densities were taken as an average of passage 3 and passage 4. The results are presented in Fig. 7.

In medium-6, a viable cell density of 1.45×10^6 cells/mL was attained, which was found to be 1.4 times higher than for the control medium and within two standard errors of $1.23E + 06$ from the original run of medium 6 (standard error = 215,928 cells/mL

Fig. 4 Response surface plot showing the effect of EAA (X2), NEAA (X3) and their mutual effect on the Y (viable cell density), when glutamine (X1) is controlled at -1 coded level

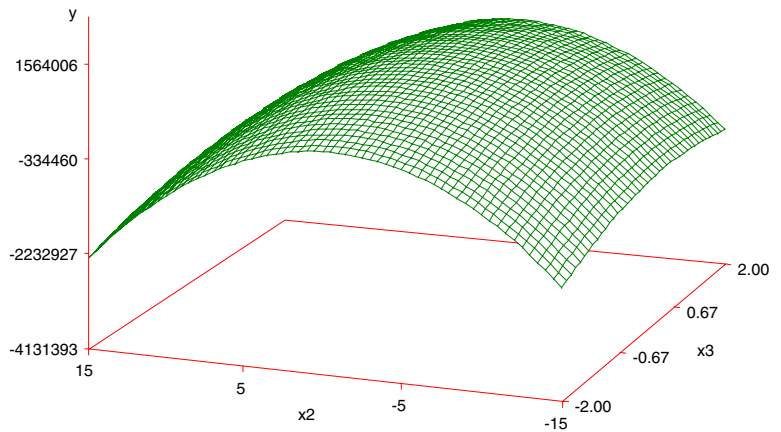


Table 9 Alternate experiments carried out on ridge and glutamine controlled at different coded levels, keeping EAA constant at coded levels 2 and 4

Medium	X1	X2	X3	VCD (10 ⁵)	SD
1	-0.52	0.196	0.225	8.13	203490
2	-1.1	0.375	0.299	8.81	179368
3	-1.487	0.491	0.329	8.61	60052
4	-1.874	0.604	0.352	8.01	152664
5	0	2	0.5	9.35	42230
6	-1	2	0.5	1.23	215928
7	-2	2	0.5	7.23	277564
8	-3	2	0.5	1.79	45162
9	0	4	0.5	7.93	102429
10	-1	4	0.5	4.49	152555
11	-2	4	0.5	5.7	144684
12	-3	4	0.5	2	52915
13	0	0	0	9.15	134722
14	0	0	0	9.48	121484

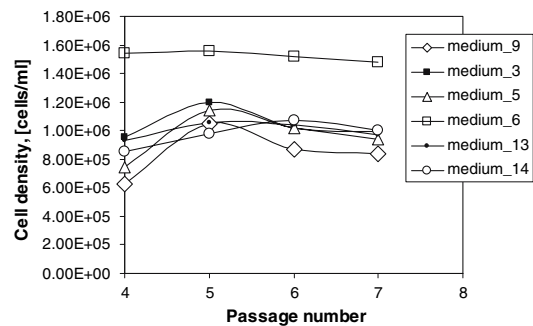


Fig. 6 Results of the alternate experiments conducted to further explore the surface

for medium 6). After four passages, if the cells were allowed to grow to passages of 5–7 the viable cell density obtained in medium-6 increased to 1.6×10^6 cells/ml. The viable cell density in the control experiment in these passages (passages 5–7) was between 9.2×10^5 cells/mL and 1.1×10^6 cells/mL, with in the standard deviation of 128,103 cells/mL (data not shown).

Assuming the specific antibody production (mg protein per cell per day) depends on the cell density, we expect to see an increase in the specific antibody production with an increase in the cell density. As detailed in the methods section, transformed cells were cultured in control and the optimized medium, medium-6, and the antibody titer was estimated in the supernatant, the specific antibody production was determined. Standard deviation was calculated in these media from triplicate experiments. The results are shown in Fig. 8. The antibody titer was determined for all the initial medium (~50) experiments, and the results were analyzed using SAS/

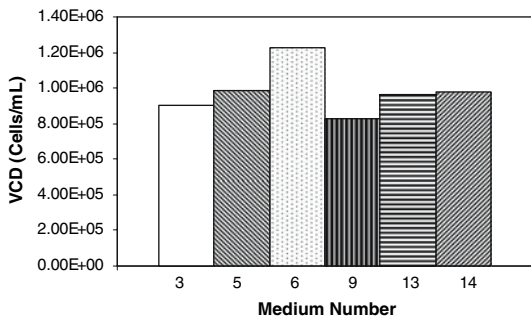


Fig. 5 Viable cell density versus medium number tested. The plot shows the viable density obtained in the last passage (passage 4) of alternate experiments undertaken. Details on media are listed in Table 9

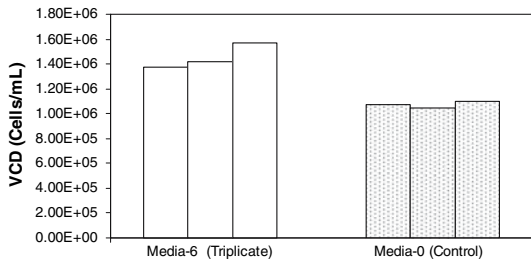


Fig. 7 Results of the triplicate experiments of Medium-6 and the Control Medium

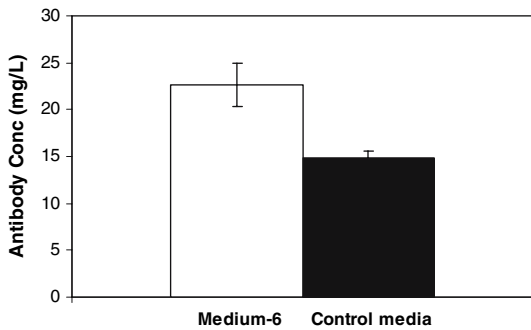


Fig. 8 Antibody production in Medium-6 and the Control Medium

STAT procedures. We observed the same trend for antibody production as well, where the stationary point is a saddle point, and the ridge values for the glutamine were moving in the negative direction and in positive direction for EAA and NEAA, shown in Table 10. The R^2 value for the model was 0.75 which shows adequacy of the model in explaining the effect of the variables on the response which is antibody production. Thus the antibody titer in medium-6 was estimated to be 1.6 times higher than the control medium and the composition of the variables in medium-6, that was found to be optimal in our present study, is listed in Table 11.

Conclusions

The increase in the Viable cell density (Cells/mL), and the production of the antibody against BoNT-A was accomplished by using Box-Wilson’s Central Composite Design.

In the optimal media, a 1.4 times higher in the viable cell density and a 1.6 times higher found antibody titer was obtained. Lower levels of glutamine

Table 10 Ridge of steepest ascent of the reduced model for getting maximum antibody production with independent variables X1, X2, X3. X4 and X5 are kept constant at their zero-level

X1	X2	X3	Estimated response (mg/L)
0	0	0	12.73
-0.172	-0.01	0.101	13.46
-0.34	-0.001	0.2	14.21
-0.522	0.03	0.293	15
-0.698	0.08	0.38	15.82
-0.874	0.149	0.461	16.69
-1.049	0.226	0.536	17.61
-1.22	0.31	0.607	18.58
-1.39	0.4	0.673	19.6
-1.56	0.496	0.736	20.6
-1.73	0.595	0.796	21.8

Table 11 Concentrations of the five variables of the optimal medium

Variables	Concentration
Glutamine	2.5 mM
EAA	3.25 × (1.67 g/L)
NEAA	2.125 × (0.168 g/L)
ITS	1 × (0.22 g/L)
Lipids	0.7 × (0.71 g/L)

Total concentration of all amino acids in EAA and NEAA are given in g/L. Concentrations of all other components are as given in Table 2

and higher levels of EAA are preferred. From the results presented here, we expect that one of the amino acids is replacing the role of amino acid-glutamine, and may be acting as limiting nutrient. Thus in our future and ongoing efforts, we will undertake amino acid analysis, to identify the limiting amino acid. We speculate that the addition of the limiting amino acid, albeit separately, separately, a decrease in the concentration of the total amino acid can be engineered, thus make the medium more economical. We also anticipate increased cell densities and higher antibody titers.

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