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# $^{1}\mathrm{H},\,^{13}\mathrm{C},\,\mathrm{and}\,^{15}\mathrm{N}$ assignments for the Archaeglobus fulgidis protein AF2095

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## LETTER TO THE EDITOR: <sup>1</sup>H, <sup>13</sup>C, and <sup>15</sup>N assignments for the *Archaeglobus fulgidis* protein AF2095

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#### **Biological context**

Structural genomics is providing a means to determine the molecular and cellular function for the vast amount of proteins in the Human proteome that lack any explicit experimental information by characterizing the complete range of protein folds (Montelione, 2001). The Northeast Structural Genomics Consortium (NESG; http://www.nesg.org/) is a pilot project funded by the National Institutes of Health Protein Structure Initiative, focusing on proteins from eukaryotic model organisms including humans. The thermophillic archaea Archaeglobus fulgidis AF2095 protein is an example of a protein of unknown biological function targeted for structural analysis by NESG. AF2095 belongs to the Pfam family PF01981 - UPF0099, protein domain family of unknown function that has been found in yeast, archaebacteria and eubacteria. AF2095 has been assigned to NESG Cluster ID:17431, a set of fourteen proteins with high (>~30%) sequence identity with human, Drosophila, Caenorhabditis elegans, Arabidopsis, yeast, archaeal and eubacterial origin (Liu, 2004). A total of fifty-six proteins are identified when the analysis is expanded to include all available genomes, where determining the NMR solution structure of AF2095 can be leveraged to infer 3D structural information for these proteins. Here we report the near complete <sup>1</sup>H, <sup>15</sup>N, and <sup>13</sup>C NMR assignments and secondary structure of AF2095. These data provide a basis for determining the solution structure of AF2095, for further investigation of the function of this protein and for providing representative structural and functional information for the protein domain family that includes AF2095.

#### Methods and experiments

Uniformly <sup>13</sup>C, <sup>15</sup>N-enriched AF2095 (123 amino acids) was cloned, expressed and purified following standard protocols used in the NESG consortium. Briefly, the full length gene (YK95 ARCFU) from Archaeglobus fulgidis was cloned into a pET21d (Novagen) derivative, yielding the plasmid pGR4-21. The resulting AF2095 open reading frame contains eight nonnative residues at the C-terminus (LEHHHHHH) of the protein. Escherichia coli BL21 (DE3) pMGK cells, a rare codon enhanced strain, were transformed with pGR4-21, and cultured in MJ9 minimal medium (Jansson et al., 1996) containing  $({}^{15}NH_4)_2SO4$  and  $U{}^{13}C$ -glucose as sole nitrogen and carbon sources. Initial growth was carried out at 37° C until the  $OD_{600}$  of the culture reached ~ 0.8 units. The incubation temperature was then decreased to 17°C and protein expression was induced by the addition of IPTG (isopropyl- $\beta$ -D-thiogalactopyranoside) at a final concentration of 1 mM. Following overnight incubation at 17°C, the cells were harvested by centrifugation and lysed by sonication. U-13C, 15N AF2095 was purified in a two step protocol consisting of Ni-NTA affinity column (Qiagen) and gel filtration column (Hi-Load 26/60 Superdex 75 pg, Amersham Biosciences) chromatography. The final yield of pure  $U^{-13}C$ , <sup>15</sup>N AF2095 (> 97%) by SDSPAGE; 13.5 KDa by MALDI-TOF mass spectrometry)



**Figure 1.** Two-dimensional <sup>15</sup>N-HSQC spectrum of AF2095 at 313 K and pH 6.5. The backbone amide and Trp side-chain resonances are assigned. The side-chain amide groups of Asn and Gln are connected by horizontal lines.

was approximately 68 mg/l. Samples of  $U^{-13}$ C,<sup>15</sup>N AF2095 for NMR spectroscopy were prepared at a protein concentration of 1.0 mM in 95% H<sub>2</sub>O/5% D<sub>2</sub>O solution containing 20 mM MES, 100 mM NaCl, 10 mM DTT, 5 mM CaCl<sub>2</sub>, 0.02% NaN<sub>3</sub> at pH 6.5.

All NMR spectra were recorded at 40°C on Bruker and Varian 500, 600, 750, and 800 MHz NMR systems. Spectra were processed using the NMRPipe software package (Delaglio *et al.*, 1995) and analyzed with PIPP (Garrett *et al.*, 1991) and AutoAssign (Zimmerman *et al.*, 1997). The assignments of the <sup>1</sup>H, <sup>15</sup>N and <sup>13</sup>C resonances were based on the following experiments: CBCA(CO)NH, CBCANH, C(CO)NH, HC(CO)NH, HNCO, HNHA, HNCA, HCCH-COSY and HCCH-TOCSY (Clore and Gronenborn, 1998; Ferentz and Wagner, 2000).

The secondary structure of AF2095 was based on characteristic NOE data involving the HN, H $\alpha$  and H $\beta$  protons,  ${}^{3}J_{HN\alpha}$ coupling constants, slowly exchanging HN protons and  ${}^{13}C\alpha$ and  ${}^{13}C\beta$  secondary chemical shifts (Wishart and Sykes, 1994; Wüthrich, 1986). The AF2095 NMR structure is composed of three helical regions corresponding to residues 17–45 ( $\alpha_1$ ); 58–70 and ( $\alpha_2$ ) and 100–109 ( $\alpha_3$ ); and a four stranded  $\beta$ -sheet region corresponds to residues 3–11 ( $\beta_1$ ); 50–55 ( $\beta_2$ ); 75–80 ( $\beta_3$ ) and 93–99 ( $\beta_4$ ).

#### Extent of assignments and data deposition

Extensive assignments for the backbone and side-chain resonances of AF2095 were obtained (208/220 HN-N, 109/115 C $\alpha$ , 104/114 C', 99/105 C $\beta$ , excluding the LEHHHHHH purification tag) (Figure 1). Initial NMR sample preparations for AF2095 suggested partial aggregation of the protein as evident by relatively low spectral signal-to-noise and a significant number of missing NMR resonance assignments. Optimizing the NMR sample conditions, particularly including data collection at elevated temperature (40° C) significantly improved the quality of the data enabling the completion of the assignments. This higher temperature is consistent with the extreme thermophillic nature of A. fulgidis and is probably closer to the functional conditions for which this protein has evolved. The amino-acid composition of AF2095 (19 L, 14 V, 7 I and 7 A) also complicated the analysis because of the relatively poor dispersion in the methyl region. Compounding this problem is the structural disorder observed for the 12 C-terminal residues, including the hexaHis purification tag (data not shown). Additionally, residues L83 to V86 and K14 to L15 are partially or completely unassigned presumably due to loop mobility since these residues are located between secondary structure elements. Unusually shifted resonance assignments for K49 and P98 were observed presumably due to a close interaction with W43 based on preliminary structural analysis. A table of chemical shifts has been deposited in the BioMagRes-Bank Database (accession code 6058).

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**Supporting Information:** Table containing the <sup>1</sup>H, <sup>15</sup>N, and <sup>13</sup>C resonance assignments for AF 2095 is included as an Appendix.

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SUPPLEMENTARY MATERIAL for <sup>1</sup>H, <sup>13</sup>C and <sup>15</sup>N Assignments for Archaeglobus fulgidis Protein AF2095. by

Robert Powers, Thomas B. Acton, Yiwen Chiang, Rajan Paranji, John R. Cort, Michael A. Kennedy, Jinfeng Liu,

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Residue N (		CO	Cα	Св	Others
M1	-(-)	-	-(-)	-(-)	
T2	-(-)	179.7	61.7(4.15)	69.3(4.20)	$C\gamma 21.5(1.33)$
L3	126.5(9.39)	173.9	54.2(4.93)	44.7(1.04.1.94)	$C_{\gamma} 27.3(1.83):C_{\delta} 27.1(0.88):C_{\delta} 23.5(0.69)$
<u>к</u> 4	117.1(8.95)	178.7	55.5(4.91)	37.2(1.72.1.56)	$C_{\chi}$ 24 1(1 17 1 06):C $\delta$ 30 0(1 57):
	11/11(0.50)	17017	0010(11)1)	5,12(11,2,1100)	$C_{\rm E} 41.1(2.63)$
05	123.1(8.42)	176.4	54,4(4,68)	35.1(1.82.1.73)	$C_{\gamma}$ 35 5(1 58):N <sub>E</sub> -(7 36 5 29)
۷6	125.8(8.80)	176.9	60 6(4 92)	34 7(1 70)	$C_{y} 21 5(0.73); C_{y} 22 2(0.82)$
17	123.0(0.00) 128.0(8.94)	176.9	60.9(4.79)	40.7(1.67)	$C_{y} = 30 (3(1) (32) (1) (22) (20) (20) (20) (20) (20) (20) (20$
17	120.0(0.94)	170.7	00.9(4.79)	40.7(1.07)	$C_{1}^{5} = 0.5(1.52, 1.13), C_{1m}^{5} = 20.5(0.73),$
V8	127 5(9 16)	175 4	60 1(4 68)	323(192)	$C_{\rm V} = 20.9(0.63)$
VO	127.3(9.10) 1107(870)	175.3	50.1(4.00)	32.3(1.72) 34.4(2.10)	$C_{\gamma} = 10.2(0.00); C_{\gamma} = 23.0(0.70)$
<b>P</b> 10	119.7(0.79) 110.5(7.68)	173.3	55 5(4 36)	34.4(2.10) 32 8(1.62)	$C_{\gamma} 26 A(1.74); C_{\gamma} 21.2(0.05)$
D11	119.3(7.08) 122 1(9.04)	174.7	54 3(4.30)	43 4(2 55 2 48)	C \ 20.4(1.74), C0 44.4(3.20)
D12	115.4(8.88)	174.2	54.7(4.40)	39.1(2.81)	
L13	118.0(6.80)	-	54.2(4.31)	43.0(1.31.1.67)	Cy 27 4(1 50):C8 22 8(0 76)
K14	119 9(8 20)	_	55.6(4.07)	30 0(1 81)	$C_{\gamma} 24 8(-):C_{\delta} 29 4(-):C_{\delta} 42 4(-)$
L15	120.0(7.69)	_	53.9(4.48)	43.8(1.29)	
S16	-(-)	175.8	57.8(4.44)	64.3(3.22.3.13)	
R17	120.5(8.83)	171.9	60.5(3.91)	30.7(1.84)	Cγ 27.4(1.61):Cδ 43.7(3.23)
G18	106.5(8.67)	175.1	47.0(3.63,4.0)	3)	
K19	120.8(7.37)	170.7	58.6(4.10)	32.5(2.00,1.81)	Cγ 25.8(1.54,1.48);Cδ 28.4(1.71);
			~ /		Ce 42.4(3.00)
L20	121.5(8.81)	172.0	58.6(3.86)	41.4(1.84.1.54)	Cγ 27.1(1.57):Cδ 25.1(0.84):Cδ 24.1(0.84)
A21	119.3(7.76)	171.9	55.5(3.76)	17.5(1.45)	
V22	115.3(7.41)	174.1	67.5(3.08)	31.2(2.16)	Сү 22.4(0.57);Сү 24.2(0.98)
O23	116.0(7.62)	172.0	58.1(3.76)	27.6(2.08)	Cγ 33.6(2.34):Nε -(6.43,5.51)
V24	117.6(7.80)	173.7	65.9(3.38)	31.0(2.16)	Cy 20.9(0.69):Cy 22.8(0.85)
A25	122.6(7.97)	171.4	55.4(3.96)	18.6(1.27)	
H26	116.3(8.68)	171.8	56.9(4.35)	31.5(3.40,2.77)	$C\delta_2$ 116.7(6.69)
A27	119.5(7.85)	172.5	54.7(3.46)	19.9(1.35)	- ` ` /
A28	119.4(8.31)	169.7	54.6(4.10)	19.4(1.53)	
I29	117.6(7.53)	173.0	63.8(3.55)	36.1(2.47)	Сү 28.4(1.70,1.39);Сү <sub>т</sub> 16.6(0.52);

Table S1 <sup>15</sup>N, <sup>13</sup>C, <sup>13</sup>CO and <sup>1</sup>H resonance assignments for AF2095 at pH 6.5 and 40°C.<sup>a</sup>

					Cδ 11.2(0.83)
I30	119.4(7.94)	172.2	65.7(3.54)	37.8(1.76)	Сү 29.3(1.50,1.09):Сүт 16.6(0.69):
			,		$C\delta 13.0(0.78)$
G31	104.5(8.47)	174.7	47.3(3.48)		
Y32	122.6(7.98)	172.9	63.4(3.50)	38.7(3.37,3.03)	C§ 132.6(7.05);Ce 118.1(6.86)
L33	118.7(8.63)	171.4	57.5(3.76)	42.2(1.88,1.46)	Cγ 26.9(1.89);Cδ 26.4(0.80);Cδ 22.5(0.90)
K34	115.1(7.39)	174.0	56.9(4.14)	33.8(1.88.1.49)	Cγ 25.1(1.40):Cδ 28.7(1.70):Cε 42.1(3.02)
S35	114.8(7.08)	178.1	59.9(4.35)	64.8(3.73,3.54)	
D36	122.9(9.01)	173.3	54.9(4.35)	43.4(2.80,2.58)	
S37	119.9(8.59)	173.8	61.8(3.85)	62.8(3.96)	
S38	118.6(8.32)	173.6	61.4(4.36)	62.4(4.01,3.93)	
L39	125.6(8.20)	170.9	58.6(4.09)	42.9(1.81,1.52)	Cγ 27.9(1.65);Cδ 25.4(0.82);Cδ 25.7(0.98)
R40	116.2(8.56)	173.4	59.1(4.15)	28.5(2.00,1.37)	Cγ 25.4(1.69);Cδ 43.4(3.00)
R41	121.8(7.25)	172.4	59.1(4.34)	30.2(2.08)	Сү 26.7(1.84,1.79);Сб 43.0(3.32)
K42	121.1(7.64)	172.0	59.4(4.15)	32.0(1.96)	Cγ 24.8(1.51);Cδ 29.0(1.72);Cε 42.1(3.00)
W43	118.6(8.44)	171.0	60.8(4.19)	27.6(3.85,2.93)	$C\delta_1$ 126.7(6.99);N $\epsilon_1$ 132.6(10.80)
					$C\zeta_3 120.6(7.90);C\zeta_2 114.9(7.08);$
					Cη <sub>2</sub> 122.5(7.14)
L44	121.2(8.16)	170.0	58.9(3.44)	41.9(2.14,1.57)	Cγ 27.0(2.06);Cδ 23.5(0.58);Cδ 26.4(0.98)
D45	121.8(8.41)	172.5	57.3(4.35)	40.2(2.97,2.77)	
E46	116.8(7.62)	174.1	56.1(4.32)	30.4(2.44,2.11)	Сү 35.5(2.74,2.41)
G47	106.7(7.59)	176.6	45.0(3.46,4.2	21)	
Q48	119.7(8.30)	176.1	52.4(-)	28.8(-)	Cγ 31.0(1.03)
K49	118.0(8.38)	171.7	58.8(4.09)	33.0(2.00)	Сү 25.4(0.20,-0.34);Сб 29.3(1.73);
					Cε 42.1(2.39)
K50	125.1(9.05)	175.6	54.7(6.01)	39.5(1.52,1.73)	Cγ 23.8(1.24);Cδ 30.6(1.57);Cε 41.4(2.63)
V51	119.0(8.25)	177.0	61.1(4.29)	36.1(1.92)	Сү 20.9(0.95)
V52	125.6(8.13)	175.6	60.8(5.33)	32.9(1.90)	Сү 22.5(0.85)
L53	128.6(8.99)	175.4	53.0(4.95)	45.7(1.59,1.37)	Сү 27.4(1.48);Сб 26.4(0.83);Сб 23.1(0.75)
K54	117.4(8.22)	174.9	54.5(5.51)	36.3(1.59)	Cγ 23.8(1.24);Cδ 29.7(1.57);
					Сє 41.7(2.85,2.96)
V55	111.7(8.12)	174.2	58.9(4.79)	34.9(2.43)	Сү 18.9(0.68);Сү 22.5(0.80)
K56	117.8(8.50)	174.9	57.8(4.35)	33.6(1.90)	Cγ 24.4(1.50);Cδ 29.0(1.70);Cε 41.7(3.00)
S57	107.6(7.14)	176.3	56.7(4.84)	67.0(4.25,3.92)	
L58	122.3(8.98)	172.6	57.7(3.96)	41.3(1.87,1.43)	Cγ 26.7(1.54);Cδ 25.4(0.99);Cδ 23.1(0.84)
E59	118.3(8.77)	170.3	60.5(3.87)	28.7(2.08,1.92)	Сү 36.2(2.37,2.27)
E60	119.5(7.91)	171.4	59.4(4.06)	30.6(2.00)	Сү 36.8(2.34,2.27)
L61	121.4(7.62)	172.0	59.0(3.74)	42.4(2.09,1.54)	Cγ 26.5(1.44);Cδ 25.8(0.66);Cδ 26.4(0.59)
L62	117.1(8.63)	170.4	57.1(3.91)	40.6(1.82,1.29)	Cγ 26.4(1.76);Cδ 25.1(0.85);Cδ 21.2(0.71)
G63	109.2(8.14)	173.5	47.4(3.95)		
I64	123.6(7.77)	173.2	63.5(3.76)	36.6(2.09)	Сү 28.7(1.51,1.39);Сүт 17.6(0.73);
					Cδ 13.1(0.78)
K65	120.7(7.75)	172.7	60.5(3.69)	33.4(2.00,1.71)	Cγ 24.8(1.22);Cδ 30.0(1.57);Cε 41.7(2.90)
H66	115.3(8.33)	172.7	58.3(4.54)	28.3(3.36)	

K67 A68	121.6(8.26) 120.8(8.43)	171.1 170.9	59.4(4.03) 55.5(3.96)	32.4(1.99,1.92) 18 3(1 37)	Cγ 25.1(1.46);Cδ 29.0(1.74);Cε 42.1(2.95)
F69	118 6(8 32)	169.3	59.3(3.70)	28.9(2.25.2.05)	$C_{\rm N}$ 36 5(2 55 2 44)
S70	117 6(8 09)	105.5	615(428)	62.8(4.01.3.92)	0 30.3(2.33,2.44)
I 71	120.9(7.18)	173.4	54.6(4.43)	42 9(1 66)	Cv 27 1(1 70)·C8 23 1(0 83)·C8 25 8(0 79)
G72	107.4(7.86)	175.8	45.8(4.08,3.7)	6)	
L73	119.9(7.43)	174.0	53.8(4.25)	42.8(1.43,1.13)	Сү 27.3(1.48);Сб 26.4(0.76);Сб 24.1(0.84)
V74	123.8(9.06)	174.8	64.1(3.75)	32.0(1.67)	Сү 21.5(1.00);Сү 22.8(0.72)
T75	116.6(7.78)	176.4	58.8(5.70)	73.4(3.96)	Сү 22.2(1.20)
G76	105.2(8.02)	179.3	44.6(4.41,3.3	0)	• • •
L77	121.7(8.47)	173.5	54.0(4.98)	45.1(1.95,1.40)	Cγ 27.4(1.71);Cδ 24.8(0.80);Cδ 26.4(0.80)
V78	121.2(8.15)	176.3	62.5(3.96)	33.0(2.05)	Сү 21.2(0.71);Сү 22.5(0.90)
Q79	124.8(7.83)	176.1	54.5(4.68)	32.5(1.82,1.73)	Cγ 34.2(2.09,1.91);Nε -(6.73,7.31)
D80	123.5(9.19)	174.2	53.4(4.74)	43.3(2.88,2.74)	• • • • • • • •
A81	128.6(8.69)	171.8	53.7(4.25)	18.6(1.43)	
G82	105.4(8.79)	176.7	45.7(4.14,3.7	1)	
L83	122.7(7.63)	-	54.7(-)	41.7(-)	
T84	-(-)	-	-(-)	-(-)	
E85	-(-)	-	-(-)	-(-)	
V86	-(-)	-	-(-)	-(-)	
P87	-(-)	-	57.8(4.85)	32.3(2.29)	Сү 27.8(1.99);Сб 50.6(3.56,3.84)
P88	-(-)	172.9	64.0(4.28)	31.7(2.33,1.88)	Cγ 27.4(2.10);Cδ 50.9(3.62,3.96)
G89	112.1(8.89)	175.7	45.0(3.50,4.1	0)	
T90	118.0(7.40)	176.9	64.9(3.98)	69.6(3.82)	Сү 21.8(1.02)
I91	129.5(8.59)	174.2	60.6(4.57)	38.6(1.68)	$C\gamma 27.4(1.80, 1.55); C\gamma_m 18.9(0.83);$
					Cδ 13.4(0.91)
T92	116.4(8.56)	175.9	62.2(4.20)	69.5(3.99)	Cγ 22.2(0.98)
A93	118.6(7.12)	175.3	51.4(5.28)	21.3(1.08)	
V94	117.3(8.81)	177.9	59.2(5.13)	36.6(1.76)	Сү 20.5(0.83);Сү 21.8(0.82)
V95	124.3(8.58)	178.4	58.0(5.18)	35.2(1.65)	Сү 16.0(0.80);Сү 22.2(0.80)
I96	125.5(9.02)	174.7	59.7(4.84)	41.2(1.54)	Сү 27.1(1.39);Сүт 19.9(0.72);Сб 14.3(0.56)
G97	111.7(8.31)	-	43.4(4.15,3.35)		
P98	-(-)	175.7	60.2(4.26)	35.0(0.09,-0.04)	Сү 24.1(1.07,0.42);Сб 49.2(3.11,1.81)
D99	118.2(8.44)	175.9	52.7(4.40)	44.7(2.55,2.45)	
E100	123.0(11.15)	171.7	57.4(4.29)	30.2(2.17,1.84)	Сү 35.9(2.45,2.38)
E101	125.8(8.64)	-	60.9(3.71)	29.8(2.09,2.03)	Сү 36.2(2.21)
R102	116.2(8.85)	172.0	59.0(4.10)	29.9(1.86,1.93)	Cγ 27.1(1.69);Cδ 43.4(3.21)
K103	117.5(7.12)	172.6	59.1(4.06)	33.8(1.81)	Cγ 25.4(1.47);Cδ 29.7(1.65);Cε 42.1(3.00)
I104	117.7(7.64)	171.7	64.7(3.47)	37.9(1.62)	$C\gamma 29.3(1.59,0.91);C\gamma_m 18.2(0.77);$
					Cδ 13.7(0.65)
D105	119.7(8.71)	172.0	56.6(4.34)	39.4(2.61)	
K106	118.6(7.16)	172.7	58.8(4.04)	32.5(1.96)	Cγ 25.1(1.50);Cδ 29.0(1.71);Cε 42.1(2.99)
V107	115.2(7.89)	173.4	64.0(3.90)	32.5(2.25)	Сү 22.5(0.89);Сү 19.6(0.79)
T108	106.4(7.77)	174.2	62.5(4.14)	68.4(4.39)	Cγ 22.2(0.89)
					• • • • •

G109	109.6(7.93)	176.4	46.8(3.88)		
N110	116.9(8.16)	175.1	52.8(4.84)	38.8(2.82,2.69)	Νδ 112.2(7.41,6.74)
L111	122.1(7.72)	-	53.8(4.59)	41.6(1.84,1.49)	Cγ-(1.71);Cδ-(0.88)
P112	-(-)	174.6	62.4(4.87)	34.8(1.71)	Cγ 24.4(1.96,1.78);Cδ 50.2(3.58,3.47)
L113	122.7(8.64)	172.7	55.3(4.35)	42.8(1.59)	Cγ 27.4(1.59);Cδ 23.8(0.90);Cδ 25.1(0.90)
L114	121.3(7.98)	173.8	56.0(4.17)	41.7(1.60)	Cγ 27.1(1.40);Cδ 24.4(0.78)
K115	122.3(7.90)	174.6	56.3(4.40)	33.6(1.76)	Cγ 24.1(1.38);Cδ 29.0(1.69);Cε 42.1(2.98)
L116	123.7(8.11)	173.6	55.3(4.29)	42.5(1.56)	Cγ 26.7(1.56);Cδ 23.5(0.99);Cδ 24.8(0.81)
E117	122.2(8.24)	174.4	56.2(4.26)	30.7(1.86)	Cγ 35.9(2.20)
H118	119.6(8.40)	176.9	55.3(4.67)	29.5(3.19,3.07)	
H119	125.4(8.24)	171.7	57.2(4.45)	29.7(3.22,3.13)	
H120	-(-)	-	-(-)	-(-)	
H121	-(-)	-	-(-)	-(-)	
H122	-(-)	-	-(-)	-(-)	
H123	-(-)	-	-(-)	-(-)	

## Footnotes to Table S1

<sup>a</sup>In each column, <sup>15</sup>N and <sup>13</sup>C shifts are listed first, and the corresponding <sup>1</sup>H shifts are given in parentheses. <sup>1</sup>H, <sup>13</sup>C and <sup>15</sup>N chemical shifts are referenced according to the method of Wishart et al. 1995