

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Faculty Publications in Food Science and
Technology

Food Science and Technology Department

2010

Co-localization of Amanitin and a Candidate Toxin-Processing Prolyl Oligopeptidase in Amanita Basidiocarps

Hong Lou

University of Nebraska, Lincoln

Heather E. Hallen-Adams

University of Nebraska at Lincoln, hhallen-adams2@unl.edu

John S. Scott-Craig

Michigan State University

Jonathan D. Walton

Michigan State University

Follow this and additional works at: <https://digitalcommons.unl.edu/foodsciefacpub>



Part of the [Food Science Commons](#)

Lou, Hong; Hallen-Adams, Heather E.; Scott-Craig, John S.; and Walton, Jonathan D., "Co-localization of Amanitin and a Candidate Toxin-Processing Prolyl Oligopeptidase in Amanita Basidiocarps" (2010). *Faculty Publications in Food Science and Technology*. 69.
<https://digitalcommons.unl.edu/foodsciefacpub/69>

This Article is brought to you for free and open access by the Food Science and Technology Department at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Faculty Publications in Food Science and Technology by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

1 September 9, 2010

2

3 **Co-localization of Amanitin and a Candidate Toxin-Processing Prolyl**
4 **Oligopeptidase in *Amanita* Basidiocarps**

5

6 Hong Luo, Heather E. Hallen-Adams¹, John S. Scott-Craig, and Jonathan D. Walton*

7

8

9 *Department of Energy Plant Research Laboratory, Michigan State University, E. Lansing*

10 *MI 48824*

11

12 ¹Current address: Department of Food Science and Technology, 319 Food Industry

13 Complex, University of Nebraska, Lincoln, NE 68583

14 * Corresponding author. Mailing address: Department of Energy Plant Research

15 Laboratory, Michigan State University, E. Lansing MI 48824. Phone: (1) 517 353 4885.

16 Fax: (1) 517 353 9168. E-mail: walton@msu.edu

17

18

19

20

21

22

23

1 Fungi in the basidiomycetous genus *Amanita* owe their high mammalian toxicity
2 to the bicyclic octapeptide amatoxins such as α -amanitin. Amatoxins and the related
3 phallotoxins (such as the heptapeptide phalloidin) are encoded by members of the
4 “MSDIN” gene family and synthesized on ribosomes as short (34 - 35 amino acid)
5 proproteins. Anti-amanitin antibodies and confocal microscopy were used to
6 determine the cellular and subcellular localization of amanitin accumulation in
7 basidiocarps (mushrooms) of the Eastern North American destroying angel
8 (*Amanita bisporigera*). Consistent with previous studies, amanitin is present
9 throughout the basidiocarp (stipe, pileus, lamellae, trama, and universal veil), but it
10 is present in only a subset of cells within these tissues. Restriction of amanitin to
11 certain cells is especially marked in the hymenium. Several lines of evidence
12 implicate a specific prolyl oligopeptidase, AbPOPB, in the initial processing of the
13 amanitin and phallotoxin proproteins. The gene for AbPOPB is restricted
14 taxonomically to the amatoxin-producing species of *Amanita* and is clustered in the
15 genome with at least one expressed member of the MSDIN gene family.
16 Immunologically, amanitin and AbPOPB show a high degree of co-localization,
17 indicating that toxin biosynthesis and accumulation occur in the same cells and
18 possibly in the same sub-cellular compartments.

19
20

1 More than 90% of all fatal mushroom poisonings are due to species of *Amanita* in
2 section *Phalloideae* such as the death cap (*A. phalloides*), and the destroying angel
3 species complex (*A. bisporigera*, *A. verna*, *A. virosa*, and *A. ocreata*) (3). The toxic
4 agents are a family of bicyclic octapeptides known as amatoxins, which survive cooking
5 and the human digestive tract and are actively taken up by liver and other cells. The
6 biochemical target of the amatoxins is RNA polymerase II, the major enzyme
7 transcribing protein-coding mRNAs in eukaryotic cells (4). The human LD₅₀ for α -
8 amanitin is ~0.1 mg/kg, and one mushroom can contain a fatal dose of 10-12 mg (36).
9 Irreversible liver failure and death may result 3-7 days after ingestion.

10 The amatoxins (and the chemically related phallotoxins such as phalloidin and
11 phalloidin) are synthesized on ribosomes and not, like all other known fungal cyclic
12 peptides, by nonribosomal peptide synthetases (10, 32). Amatoxins are translated as 35-
13 amino acid propeptides and must undergo several posttranslational modifications,
14 including proteolytic cleavage, cyclization, hydroxylation, and formation of a unique
15 tryptophan-cysteine cross-bridge called tryptathionine (19). The amatoxin and
16 phallotoxin genes are members of a large family, called the “MSDIN” family, that are
17 predicted to encode propeptides of 34 to 37 amino acids with conserved upstream and
18 downstream sequences flanking a hypervariable region of 7 to 10 amino acids. The
19 hypervariable region gives rise to the linear peptides corresponding to the mature toxins
20 (18). Certain species of *Amanita* have thus evolved the genetic potential to synthesize a
21 large variety of small, cyclic peptides using the same fundamental biochemical
22 mechanism.

1 In the MSDIN family of genes, both the amino acid immediately preceding the toxin
2 region and the last amino acid in the toxin region itself are invariant Pro residues (10).
3 All known amatoxins and phallotoxins, as well as other cyclic peptides that have been
4 identified in *A. phalloides*, contain at least one Pro (36). A peptidase that cleaves a
5 synthetic phalloidin peptide at the carboxy-side of both Pro residues to release the linear
6 phalloidin heptapeptide was purified from the phalloidin-producing mushroom *Conocybe*
7 *apala* (= *C. lactea* or *C. albipes*) (18). This processing peptidase was identified as a
8 member of the prolyl oligopeptidase (POP) family of proteases (6, 29). Due to limiting
9 amounts of biological material (*Amanita* does not form fruiting bodies in culture), and the
10 instability of the enzyme, it has not yet been possible to determine if a POP enzyme also
11 processes the amanitins in species of *Amanita*.

12 Temporal and structural sequestration is a common feature of secondary metabolism
13 in micro-organisms and plants, serving to prevent these often biologically active
14 compounds from interfering with primary metabolism and to insure their presence at the
15 ecologically most appropriate time and place (16). Mushrooms (here defined as the
16 basidiocarps of macrofungi in the phylum Basidiomycota) produce a number of
17 biologically active secondary metabolites, and some mechanism of separation from
18 primary metabolism would seem to be essential for both the amatoxins and the
19 phallotoxins, since their sites of action (RNA polymerase II and F-actin, respectively) are
20 present in all eukaryotic cells.

21 Many secondary metabolites are known to be synthesized in specialized cells in
22 other organisms, such as plant trichomes and cone snail secretory ducts (16, 34).
23 However, little is known about the relationship between chemical and structural cellular

1 specialization in relation to natural product production in macrofungi. Besides the
2 amatoxins and phallotoxins, examples of biologically active molecules from mushrooms
3 include psilocybin, lovastatin, muscarine, ibotenic acid, muscimol, strobilurins,
4 pleuromutilin, and illudins (1, 27). Specialized mushroom structures known or suspected
5 to contain toxic compounds include cystidia in genera such as *Inocybe*, *Russula*, and
6 *Strobilurus*, which are poisonous to mammals or insects (13, 15, 17, 23). *Conocybe apala*,
7 which makes phalloidin, has specialized secretory cells that are toxic to nematodes (12).
8 *Pleurotus cystidiosus* makes specialized cells (toxocysts) consisting of a liquid droplet
9 containing nematocidal compounds surrounded by an elastic adherent envelope (30).
10 Differential cytological staining indicates that many mushrooms contain biochemically
11 specialized hyphae, called secretory hyphae, that are intermingled with the structural
12 hyphae (5). However, the relationship between structure and chemistry has rarely been
13 established in any mushroom, i.e., whether the active compounds are actually located in
14 the specialized cell types.

15 In this paper, we use a specific anti-amanitin antibody to localize the sites of
16 amanitin accumulation in relation to the cellular distribution of AbPOPB, an enzyme
17 implicated in the post-translational processing of the amatoxin proproteins. The results
18 indicate that amanitins are synthesized and stored in a subset of cells found throughout
19 the mushrooms.

20

21

MATERIALS AND METHODS

22

Biological material. Mushrooms of the Eastern North American destroying angel

23 (*Amanita bisporigera*) were collected in Ingham County, Michigan, in July and August of

1 2008 and 2009 (Fig. 1). Very young fruiting bodies (button stage, still enclosed in the
2 universal veil, and not yet emerged above the ground) were also collected when possible
3 (Fig. 1). Mushrooms were fixed in 4% paraformaldehyde within 2 to 4 hr of collection.
4 All the mushrooms used in this study were analyzed by HPLC to confirm the presence of
5 amanitins (7, 11). Amanitin concentrations varied from mushroom to mushroom, and
6 samples with higher concentrations were chosen for the immunohistochemical analyses.

7 Amatoxin and phallotoxin-lacking species of *Amanita* (*A. citrina* and *A. muscaria*)
8 were also collected in the same location as controls. The other *Amanita* species used as a
9 source of DNA for blotting were previously described (10).

10 **Immunological and microscopic methods.** To prepare paraffin sections, blocks of
11 tissue (~3 mm cubes) were fixed in 4% paraformaldehyde for 1 h at room temperature.
12 Following graded dehydration in ethanol, tissue blocks were transferred to xylene and
13 incubated for 10 min with stirring. This step was repeated and tissue blocks were then
14 embedded in paraffin by standard methods. Sectioning to 5–10 μm thickness was
15 performed with a rotary microtome (Leica RM2155). Selected sections were mounted on
16 slides and the paraffin removed by washing twice in xylene (5 min each wash). Xylene
17 was then removed by washing the slides in ethanol twice (5 min each wash), after which
18 a graded rehydration step was conducted in ethanol to phosphate-buffered saline (PBS).
19 The slides were boiled for 20 min in a staining dish in the presence of 100 mM sodium
20 citrate buffer, pH 6.0. After cooling at room temperature for 20 min, the slides were
21 washed twice in PBS for 5 min.

22 Before hybridization, tissue sections on the slides were incubated with blocking
23 buffer (3% dry milk in PBS, 0.1% Tween-20, and whole goat polyclonal IgG (Santa Cruz

1 Biotechnology, Santa Cruz, CA, catalog sc-6654) at a 1:30 dilution for 4 h at room
2 temperature.

3 Primary antibodies were prepared in the same blocking buffer at 1:30 dilution. For
4 dual staining, the two primary antibodies were added at the same time. The slides plus
5 primary antibody were incubated typically for 4 h in a humidified chamber. The AbPOPB
6 antibody signal in mature mushrooms was consistently weaker than in immature
7 mushrooms. Therefore, tissue sections were incubated with anti-AbPOPB and anti-
8 amanitin antibodies for 15 hr instead of the usual 4 h. The slides were then rinsed twice
9 for 5 min each time in PBS buffer and incubated with secondary antibody, diluted
10 according to the manufacturer's instructions in blocking buffer, in a humidified chamber
11 for 2 h. The slides were rinsed twice for 10 min each in PBS, mounted with one drop of
12 Citifluor (Citifluor Ltd, Leicester, United Kingdom), and viewed with an Olympus
13 FV1000D confocal laser scanning microscope.

14 The anti-amanitin antibody was obtained from Heinz Faulstich (Heidelberg,
15 Germany) (14). The antibody had been raised in rabbits and purified by affinity
16 chromatography. It can detect 1.5 ng/ml α -amanitin in human serum and urine. It reacts
17 with α - and γ -amanitin but not β -amanitin or phallotoxins (14). Most specimens of *A.*
18 *bisporigera* produce α -, β -, γ -, and ϵ -amanitin as well as several phallotoxins (21).

19 The antibodies against AbPOPA and AbPOPB were raised in mouse (Cocalico
20 Biologicals, Reamstown, PA). Whole serum was used for immunoblots and
21 immunohistology. A monoclonal antibody against a synthetic actin C-terminal peptide
22 was produced in mouse (Sigma-Aldrich A3853). The secondary antibodies used were

1 Alexa Fluor 488 goat anti-mouse IgG (Invitrogen A11029) and Alexa Fluor 594 goat
2 anti-rabbit IgG (Invitrogen A11037).

3 The specificity of the antibody was tested by immunoblotting against inclusion
4 bodies of *Escherichia coli* expressing AbPOPB and against total extracts of *A.*
5 *bisporigera*. Total protein was obtained by grinding 0.5 g frozen basidiocarps of *A.*
6 *bisporigera* in liquid nitrogen. To the resulting powder was added 2 ml of a stock
7 solution of protease inhibitors. The stock solution was made by dissolving five tablets of
8 Complete Mini EDTA-free Protease Inhibitor (Roche Applied Science, Mannheim,
9 Germany) in 5 ml water and adding an additional 5 ml of Protease Inhibitor Cocktail
10 (Sigma P2714) diluted with water according to the manufacturer's instructions. The
11 homogenate was centrifuged at $14,000 \times g$ in 1.5-ml microtubes for 15 min. The
12 supernatant was fractionated by SDS-PAGE (pre-made gels of 7.5% acrylamide in Tris-
13 HCl, Bio-Rad, Richmond, CA). The SDS-PAGE markers were Bio-Rad Precision Plus
14 (Dual Color). In order to minimize proteolytic degradation of the POP proteins, the entire
15 extraction procedure was performed in less than 30 min and all solutions were kept on ice
16 or at 4°C. Proteins were transferred to nitrocellulose membrane (Bio-Rad, catalog 162-
17 0115) and detected using ECL Western Blotting Substrate according to the
18 manufacturer's instructions (Pierce, Rockford, IL, catalog 32106). Anti-POP antiserum
19 was incubated with the membrane for 30 min. The membranes were subsequently
20 exposed on Hyperfilm MP (Amersham/GE Healthcare, Buckinghamshire, United
21 Kingdom) for 15 to 60 sec.

22 **Isolation and expression of prolyl oligopeptidase genes.** For cloning *AbPOPA* and
23 *AbPOPB*, PCR primers corresponding to the N- and C-termini of both putative genes

1 (forward primers: 5'-GAAACGAGAGGGCGAAGTCAAGGTG-3' and 5'-
2 TCAAATGAAGTAGACGAATGGAC-3'; reverse primers: 5'-
3 AAGTGGATGACGATTATGCGGCAG-3' and 5'-
4 CACACGGATGAGCAATGGATGAG-3') were used in both combinations and the
5 amplicons cloned into *E. coli* DH5 α . These primers were based on the partial genomic
6 sequence of *A. bisporigera* (10). The amplicons were cloned into *E. coli* DH5 α and
7 sequenced. The full sequences of the genes were obtained from lambda clones (see
8 below). Full-length cDNA copies were obtained by 5' and 3' Rapid Amplification of
9 cDNA Ends (RACE) (18) using primers 5'-ATGTCTCGCCGAACTCGCCGCCTCCTC-
10 3' (5'-RACE) and 5'-GATTGGGTATTTGGCGCAGAAGTCACG-3' (3'-RACE) for
11 *AbPOPA* and primers 5'-GTGACGAACGATGAACATTGGAACCTT-3' and 5'-
12 GAAGATGGTAAATATGTGGCCCTGTA-3' for *AbPOPB*. Sequences generated from
13 the RACE reactions were used to assemble full-length cDNAs of *AbPOPA* and *AbPOPB*.
14 *AbPOPA* and *AbPOPB* cDNAs were cloned into the pET21 vector between the
15 *Bam*HI and *Not*I sites. T-7 tagged versions of both genes were also generated. The
16 expression constructs were transformed into *E. coli* strain BL21 (DE3) (Novagen, EMD
17 Chemicals, Inc., Darmstadt, Germany) and three positive clones for each construct were
18 selected. Expression was induced in a 50-ml culture at an OD₆₀₀ of 0.7 with 0.4 mM
19 isopropyl β -D-1-thiogalactopyranoside (IPTG) following the manufacturer's protocols
20 (Novagen, 11th edition). The cells were disrupted by freeze/thaw treatment and treated
21 with 0.5 U/ml Benzonase nuclease.
22 Both expressed POP proteins were present in inclusion bodies and yielded no
23 detectable activity with the chromogenic POP substrate Z-Gly-Pro-pNA (Sigma-Aldrich)

1 at 1 mM (18). Instead, we purified the inclusion bodies of expressed AbPOPA and
2 AbPOPB. The untagged POP proteins were then separated by SDS-PAGE and gel slices
3 containing the POP proteins were used for antibody production. Anti-AbPOPA antibody
4 was raised in chicken and anti-AbPOPB antibody was raised in mouse by Cocalico
5 Biologicals (Reamstown, PA) using their standard protocols.

6 **Nucleic acid manipulations.** DNA for blotting was digested with *Pst*I and
7 electrophoresed in 0.7% agarose. Hybridizations were performed overnight at 65°C in 4x
8 SET, 0.1% sodium pyrophosphate, 0.2% SDS, 10% dextran sulfate, and 625 µg/ml
9 heparin. SET (20×) is 3 M NaCl, 0.6 M Tris, and 0.04 M EDTA, pH 7.4. Full-length
10 cDNAs of *AbPOPA* and *AbPOPB* were labeled with [³²P]dCTP to use as probes. To
11 construct the genomic lambda library, genomic DNA of *A. bisporigera* was partially
12 digested with *Bfu*CI and electrophoresed in a 0.5% agarose gel at 5 V/cm. DNA fragments
13 of 12 to 20 kb were recovered with the QIAEX II gel extraction kit (Qiagen, Valencia,
14 CA, catalog 20021). The fragments were cloned into the λBlueSTAR Vector (Novagen).
15 The resulting library was screened with *AbPOPA* and *AbPOPB* cDNA probes by standard
16 methods (28). Positive clones were subcloned into *E. coli* strain BMC25.8 and sequenced.

17 **Nucleotide sequence accession numbers.** Sequence data from this article can be
18 found in the GenBank/EMBL database under the following accession numbers:
19 HQ225840 for *AbPOPA*, HQ225841 for *AbPOPB*, and EU196153 (updated) for *MSD-10*
20 of *A. bisporigera*.

21

22

23

RESULTS

1 **Immunolocalization of Amanitin.** In order to investigate the types of tissue in
2 which amanitin is distributed within a basidiocarp, samples were examined from several
3 representative tissue types, including pileus (cap), lamella (gill), stipe (stem), and
4 universal veil (Fig. 1). Multiple basidiocarps of different developmental stages and
5 various locations were examined.

6 The most intense and specific anti-amanitin signals were found in a sub-set of cells
7 located in the hymenium, i.e., on the lamella edges where the basidia are formed (Fig. 2A
8 – C). In some samples, there was a strong signal from the sub-hymenial layer in addition
9 to the cells of the hymenium (Fig. 2D). Under higher magnification of a single lamella,
10 the presence of amanitin in only certain hymenial cells and in the sub-hymenium was
11 clear (Fig. 2E, G). Some of the hymenial cells containing amanitin were true basidia
12 (arrows) because they have sterigmata (the slender projections on the basidia to which
13 basidiospores are attached before they are released) whereas others might be either sterile
14 cells or basidia at different stages of maturity (arrowheads) (Fig. 2E–I). Not all basidia
15 contained amanitin, nor did all sterile cells. Fig. 2I shows hymenial cells in which two
16 basidia with sterigmata did not show the presence of amanitin, whereas several adjacent
17 sterile cells did. The trama (inner part of the lamella) consistently showed weaker
18 staining than the hymenium or the sub-hymenium (Fig. 2A, B, D).

19 The pileus (cap) of *A. bisporigera* also gave strong signals with the anti-amanitin
20 antibody. Figure 3A shows a cross-section of the pileus of a young specimen of *A.*
21 *bisporigera* and Fig. 3B shows a mature pileus. The individual areas of signal were
22 irregular in shape and their locations did not correspond to cellular boundaries (Fig. 2B,
23 C). The areas giving amanitin signal were consistently smaller than the cells. In Fig. 3C,

1 the cellular boundaries of the section in Fig. 3B have been outlined in white in order to
2 better illustrate this point. The apparent subcellular localization of amanitin was also
3 visible at higher magnification, where the outlines of individual cells could be seen (Fig.
4 3D). A similar irregular and apparent subcellular pattern of amanitin distribution was
5 seen in the stipe (Fig. 3E) and in the universal veil (Fig. 3F).

6 Mushrooms of the amanitin-nonproducing species *A. citrina* and *A. muscaria* were
7 fixed, sectioned, and treated identically with the anti-amanitin antibody. No signals were
8 seen in any sections (data not shown).

9 In order to improve visualization of background cells not containing amanitin, a
10 universal anti-actin antibody was used as a general indicator of all cells. Figures 4A–C
11 show immunostaining of lamellae with anti-amanitin antibody, anti-actin antibody, and a
12 merge of the two. Figures 4D–F show higher magnification of a single lamella. Amanitin
13 was present in the trama, the subhymenium, and a subset of cells in the hymenium (Fig.
14 4C, F). Compared to amanitin distribution, the anti-actin antibody produced an even
15 staining throughout the lamellae, as expected for the distribution of a housekeeping
16 protein, with a somewhat stronger signal in the hymenium (Fig. 4B, E).

17 **Identification of a candidate amatoxin-processing peptidase in *A. bisporigera*.** It
18 is of particular interest to understand the relationship between the sites of amanitin
19 accumulation and amanitin biosynthesis. To date, no enzymes involved in amatoxin
20 biosynthesis except those coding the toxins themselves have been identified (10, 18). A
21 prolyl oligopeptidase (POP) from the phalloidin-producing mushroom *Conocybe apala*
22 cleaves a synthetic phalloidin proprotein to release the linear heptapeptide corresponding
23 to mature phalloidin, consistent with this enzyme being responsible for the initial

1 cleavage of the toxin propeptides (18). Using the *C. apala* POP protein (GenBank
2 ACQ65797) and the human POP protein (BAA86936) as queries, we identified partial
3 sequences for two POP genes, *AbPOPA* and *AbPOPB*, in the genome survey sequence of
4 *A. bisporigera*. Based on an analysis of a partial genome sequence of *Galerina marginata*,
5 which makes amatoxins (but not phallotoxins), this mushroom also has two POP genes
6 (22, and our unpublished results). In contrast, all other basidiomycetes whose full
7 genomes were available in GenBank or at the DOE Joint Genome Institute as of May,
8 2010, have a single POP gene (10). Highly similar orthologs of *AbPOPA* and *AbPOPB*
9 are present in plants, bacteria, and animals. No evidence was found for any POP genes in
10 ascomycetes, except perhaps weakly similar orthologs in *Nectria haematococca*
11 (EEU39197) and *Phaeosphaeria nodorum* (XP_001801532).

12 The two predicted POP genes of *A. bisporigera* and their corresponding cDNA
13 copies were cloned and sequenced. *AbPOPA* and *AbPOPB* each have 18 introns (Fig. 5).
14 The amino acid sequences of the predicted translational products of *AbPOPA* (761 amino
15 acids) and *AbPOPB* (730 amino acids) were 57% identical to each other and 62% and
16 57% identical, respectively, to the phalloidin-processing POP (733 amino acids) of *C.*
17 *apala* (18).

18 Within the genus *Amanita*, the genes encoding α -amanitin and phalloidin (*AMAI*
19 and *PHAI*, respectively) were found by DNA blotting to be present only in species that
20 make these compounds (10). We probed genomic DNA blots of 13 species in the genus
21 *Amanita*, representing all five sections, with *AbPOPA* and *AbPOPB*. DNA from all of the
22 tested species showed hybridization to a *AbPOPA* probe, whereas only DNA from
23 species in section *Phalloideae*, which contains all of the amatoxin- and phallotoxin-

1 producing species, showed hybridization to an *AbPOPB* probe (Fig. 6). Due to taxonomic
2 divergence and variation in loading, some of the signals with *AbPOPA* were weak (Fig.
3 6A). However, most significantly, DNA from none of the species in section *Validae*, the
4 sister group to section *Phalloideae*, showed hybridization to *AbPOPB* even with
5 prolonged exposure (Fig. 6A, lanes 5 – 8). DNA from these same specimens showed the
6 same pattern of hybridization or nonhybridization with *AMA1* and *PHAI* probes (10).

7 Because the genes of a particular secondary metabolite biosynthetic pathway tend to
8 be clustered in fungi, clustering of *AbPOPA* or *AbPOPB* with other genes involved in
9 amatoxin biosynthesis would suggest a role for one or both of these genes in amatoxin
10 biosynthesis (1, 27, 35). To test this, we screened a lambda phage clone library with
11 *AbPOPA* and *AbPOPB*, and two hybridizing clones were sequenced completely.
12 FGENESH (www.softberry.com) was used to predict proteins *ab initio* using the
13 *Coprinopsis cinerea* model. Each predicted protein was then used to search GenBank nr
14 using BLASTP. Five genes were predicted in the 13.5-kb *AbPOPA*-containing clone (Fig.
15 7). Gene 4 was *AbPOPA*. The predicted products of genes 1, 2, 3, and 5 had strong hits
16 (score < 1e-62) to hypothetical proteins in other basidiomycetes. Genes 3 and 5 were
17 predicted to encode mitochondrial transporters (pfam 00153) with orthologs in other
18 basidiomycetes.

19 Six genes were predicted by FGENESH in the 16.0-kb lambda clone containing
20 *AbPOPB*. None had expect scores less than 1e-26 against any protein in GenBank nr. The
21 best hits of genes a, c, e, and f were to hypothetical proteins in the ectomycorrhizal
22 basidiomycete *Laccaria bicolor*. Gene b was *AbPOPB*. Gene d was a member of the
23 MSDIN family previously identified in the genome survey sequence of *A. bisporigera*

1 (*MSD-10*; GenBank EU196153). A full-length cDNA copy of *MSD-10* was obtained
2 using 5' and 3' RACE, indicating that this gene is expressed and contains three introns,
3 like *AMA1* and *PHAI*. The predicted amino acid sequence of the hypervariable putative
4 toxin region of *MSD-10* is GAYPPVPMP, and therefore *MSD-10* is predicted to encode a
5 cyclic nonapeptide containing four Pro residues and lacking tryptathionine.

6 **Immunolocalization of *A. bisporigera* prolyl oligopeptidase B.** Based on these
7 several lines of evidence supporting a role for AbPOPB in the biosynthesis of amatoxins
8 in *A. bisporigera*, antibodies were raised against AbPOPB in order to determine the
9 relationship between cellular amatoxin accumulation and biosynthesis. Because neither
10 *AbPOPA* nor *AbPOPB* could be expressed in an enzymatically active form in either *E.*
11 *coli* or *Pichia pastoris* (unpublished results), we isolated AbPOPB from inclusion bodies
12 of *E. coli* (Fig. 8). The resulting anti-AbPOPB antibody did not recognize other proteins
13 in total extracts of *A. bisporigera* (Fig. 8). Like other fungal POPs (18), AbPOPB was
14 very sensitive to degradation during extraction. Full-length AbPOPB could not be
15 detected without the addition of high levels of protease inhibitors in the extraction buffer.
16 Even then, degradation usually occurred, as could be seen at high protein loadings (Fig.
17 8). Although antibodies against AbPOPA were successfully raised, the antibodies failed
18 to produce a clear and specific signal when tested on whole mushroom extracts (data not
19 shown). We therefore used only the anti-AbPOPB antibody for immunohistochemical
20 studies.

21 Dual labeling of pileus (cap) tissue from an immature mushroom indicated strong co-
22 localization of amanitin and AbPOPB, as indicated by the orange to yellow color (Fig. 9).
23 Figures 9D – F show higher magnification views of the area indicated by the white box in

1 Fig. 9C. Calculation of the overlap coefficients showed 52.0% of AbPOPB co-localizing
2 with amanitin and 99.9% of amanitin co-localizing with AbPOPB. Co-localization of
3 amanitin and AbPOPB were also examined in lamellae (Fig. 9G – I). The merge between
4 the two (i.e., orange to yellow color) showed significant overlap of the two signals (Fig.
5 9I). Amanitin and AbPOPB were both present in certain hymenial cells and not others
6 (Fig. 9G – I). Several areas were analyzed and the co-localization was calculated at better
7 than 75%.

8

9

DISCUSSION

10 Although amanitins have been known to be distributed throughout the tissue of
11 *Amanita* mushrooms, their cellular localization has not previously been determined (8).
12 Here we show that the distribution of amanitin is not uniform among cells of the
13 basidiocarp, nor are they sequestered in a morphologically distinct cell type, unlike the
14 anti-collembola toxins of *R. bella* or the anti-nematode compounds of *C. apala* (12, 23).
15 Selective distribution of amatoxins was particularly apparent in the organized cells of the
16 hymenium and sub-hymenium as opposed to the interwoven ground hyphae of the pileus
17 and stipe. Furthermore, our results indicate that amanitin is not evenly distributed within
18 the cells. Rather, amanitin appears to be compartmentalized at the sub-cellular level.
19 Although we did not resolve the structures containing amanitin, they are most likely
20 vacuoles, such as are known to be the sites of accumulation of secondary metabolites in
21 other organisms (16). Aflatoxins and some their biosynthetic intermediates accumulate in
22 vacuoles and vesicles in *Aspergillus parasiticus*, even though other steps in aflatoxin
23 biosynthesis occur in peroxisomes or the cytoplasm (26). However, a fundamental

1 difference between amanitins and aflatoxins (and most other ascomycete secondary
2 metabolites) is that whereas aflatoxins are ultimately secreted into the medium, amanitins
3 are not secreted and are therefore stored inside cells in a way that sequesters them from
4 primary metabolism (37).

5 A particular question that this paper begins to address is the relationship between
6 amanitin accumulation and amanitin biosynthesis. At this point we know little about the
7 enzymology of the post-translational processing steps of amanitin biosynthesis, which
8 include cleavage of the proprotein, cyclization, hydroxylation, formation of the cross-
9 bridge between Trp and Cys, and, in the case of the phallotoxins, epimerization of L-Thr
10 or L-Asp to the corresponding D isomer.

11 Several lines of evidence implicate prolyl oligopeptidase (POP) in cleavage of the
12 proprotein, which is the predicted first step of post-translational maturation of the
13 amatoxins and phallotoxins (18). First, all members of the MSDIN family, which
14 includes the genes encoding α -amanitin, β -amanitin, phallacidin, phalloidin, and a cyclic
15 decapeptide related to antamanide, have invariant Pro residues as the amino acid
16 immediately upstream of the hypervariable toxin region that contains the peptide
17 sequence of the mature toxins and as the last amino acid in the toxin region (10, 18, 36).
18 This indicates that posttranslational cleavage of the toxin proproteins is probably
19 catalyzed by a Pro-specific peptidase (6, 29). Second, an enzyme capable of cleaving a
20 phalloidin precursor protein at both Pro residues, resulting in release of the cognate linear
21 heptapeptide, was identified as a POP in the phalloidin-producing mushroom *C. apala*
22 (18).

1 In this paper we provide additional evidence to support the *in vivo* role of POP in
2 amanitin biosynthesis. First, *AbPOPB*, but not *AbPOPA*, is restricted taxonomically to
3 those species in the genus *Amanita* that make amatoxins. A plausible explanation for why
4 toxic *Amanita* species (and the amanitin-producing species *Galerina marginata*; our
5 unpublished data) have two POP genes whereas other basidiomycetes have only one is
6 that one of the POPs (*AbPOPA* in the case of *A. bisporigera*) is a housekeeping POP that
7 is functionally equivalent to the single POP found in other basidiomycetes, whereas the
8 second POP (*AbPOPB*) has a dedicated function in toxin biosynthesis. Although POPs
9 are widespread in nature, having been described in plants, animals, trypanosomes,
10 bacteria, and other basidiomycetes, little is known about their native substrates except in
11 mammals (2). Human POP cleaves several peptide hormones *in vitro* and *in vivo* and has
12 been implicated in human pathologies including dementia, amnesia, multiple sclerosis,
13 and sleeping sickness (24). The native substrates of POPs in other organisms have been
14 little studied. Our results point to the proproteins of the MSDIN family as the probable *in*
15 *vivo* substrates of a particular class of POPs in certain basidiomycetes.

16 A third line of evidence in support of a role of *AbPOPB* in amatoxin processing is
17 genetic linkage between *AbPOPB* and an expressed member of the MSDIN family
18 (*MSD-10*). Clustering of genes involved in a common pathway is the usual situation in
19 fungal secondary metabolism (31). Although there has been less work done on the
20 genetics of secondary metabolism in basidiomycetes compared to ascomycetes, this
21 appears to be the emerging consensus for this group of organisms as well (1, 27, 35).

22 Taken together, the evidence strongly implicates *AbPOPB* in the biosynthesis of the
23 amatoxins and phallotoxins, and therefore we raised antibodies against *AbPOPB* to use as

1 a marker to localize the site of amanitin biosynthesis. In fact, there was a high degree of
2 co-localization between amanitin and AbPOPB, which is consistent with synthesis and
3 accumulation occurring in the same cells.

4 Natural products serve as the major source of pharmaceutical agents against a wide
5 variety of old and emerging human diseases. However, most of our medicines are based
6 on only a few fundamental molecular scaffolds, which have been heavily exploited by
7 generations of tailoring (33). Historically, toxins have shown great value in drug
8 development and pharmaceutical use, especially the highly potent ones that have specific
9 targets. *Amanita* toxins are highly potent and highly specific, which has led to their
10 widespread use as research reagents. In several ways, the peptide toxins of *Amanita* are
11 chemically and biochemically unique among known natural products, e.g., they are the
12 only ones that contain the Cys-Trp cross-bridge (tryptathionine), and they are the smallest
13 known ribosomally synthesized peptides. They may therefore provide a novel scaffold on
14 which to design new pharmaceutical agents (9, 19, 20, 25). Identification of the genes and
15 enzymes involved in the biosynthesis of the *Amanita* toxins may make it possible to
16 produce a large number of new compounds based on the naturally occurring toxins.

17 Compared to the other main groups of micro-organisms that make secondary
18 metabolites (namely, bacteria and ascomycetes), relatively little is known about the
19 chemistry, biochemistry, or biology of secondary metabolites made by higher
20 basidiomycetes, commonly known as mushrooms (27). Compared to ascomycetes and
21 bacteria, mushrooms have more complex anatomies and consequently more cell
22 specialization. The results presented in this paper illustrate this cellular specialization for
23 both toxin accumulation and biosynthesis.

1

2

ACKNOWLEDGMENTS

3 We thank Heniz Faulstich (Heidelberg, Germany) for the generous gift of anti-
4 amanitin antibodies, Melinda Frame of the Michigan State University (MSU) Microscopy
5 Facility and Markus Günl (MSU) for scientific advice, and Federica Brandizzi (MSU) for
6 comments on the manuscript. This work was funded by grant DE-FG02-91ER200021
7 from the U.S. Department of Energy Office of Basic Energy Sciences, Division of
8 Chemical Sciences, Geosciences and Biosciences.

9

10

REFERENCES

11

- 12 1. **Agger, S., F. Lopez-Gallego, and C. Schmidt-Dannert.** 2009. Diversity of
13 sesquiterpene synthases in the basidiomycete *Coprinus cinereus*. *Mol. Microbiol.*
14 **72**:1181–1195.
- 15 2. **Bastos, I. M., F. N. Motta, S. Charneau, J. M. Santana, L. Dubost, K.**
16 **Augustyns, and P. Grellier.** 2010. Prolyl oligopeptidase of *Trypanosoma brucei*
17 hydrolyzes native collagen, peptide hormones and is active in the plasma of
18 infected mice. *Microbes Infect.* **12**:457–466.
- 19 3. **Bresinsky, A., and H. Besl.** 1990. *In* A Colour Atlas of Poisonous Fungi: A
20 Handbook for Pharmacists, Doctors and Biologists. Wolfe, Würzburg, Germany.
- 21 4. **Bushnell, D. A., P. Cramer, and R. D. Kornberg.** 2002. Structural basis of
22 transcription: alpha-amanitin-RNA polymerase II cocrystal at 2.8 Å resolution.
23 *Proc. Natl. Acad. Sci. USA* **99**:1218–1222.

- 1 5. **Clémentçon, H.** 2004. *In* Cytology and Plectology of the Hymenomycetes. J.
2 Cramer, Berlin.
- 3 6. **Cunningham, D. F., and B. O'Connor.** 1997. Proline specific peptidases.
4 *Biochim. Biophys. Acta* **1343**:160–186.
- 5 7. **Enjalbert, F., C. Gallion, F. Jehl, H. Monteil, and H. Faulstich.** 1992.
6 Simultaneous assay for amatoxins and phallotoxins in *Amanita phalloides* Fr. by
7 high-performance liquid chromatography. *J. Chromatogr.* **598**:227–236.
- 8 8. **Enjalbert, F., G. Cassanas, S. L. Salhi, C. Guinchard, and J. -P. Chaumont.**
9 1999. Distribution of the amatoxins and phallotoxins in *Amanita phalloides*.
10 Influence of the tissues and the collection site. *C.R. Acad. Sci. III* **322**:855–862.
- 11 9. **Fischbach, M. A., and C. T. Walsh.** 2009. Antibiotics for emerging pathogens.
12 *Science* **325**:1089–1093.
- 13 10. **Hallen, H. E., H. Luo, J. S. Scott-Craig, and J. D. Walton.** 2007. Gene family
14 encoding the major toxins of lethal *Amanita* mushrooms. *Proc. Natl. Acad. Sci.*
15 USA **104**:19097–19101.
- 16 11. **Hallen, H. E., R. Watling, and G. C. Adams.** 2003. Taxonomy and toxicity of
17 *Conocybe lactea* and related species. *Mycol. Res.* **107**:969–979.
- 18 12. **Hutchison, L. J., S. E. Madzia, and G. L. Barron.** 1996. The presence and
19 antifeedant function of toxin-producing secretory cells on hyphae of the lawn-
20 inhabiting agaric *Conocybe lactea*. *Can. J. Bot.* **74**:431–434.
- 21 13. **Jenkinson, T. S., G. J. Celio, M. Padamsee, B. T. M. Dentinger, M. L. Meyer,**
22 **and D. J. McLaughlin.** 2008. Conservation of cytoplasmic organization in the
23 cystidia of *Suillus* species. *Mycologia* **100**:539–547.

- 1 14. **Kirchner, K., and H. Faulstich.** 1986. Purification of amatoxin-specific
2 antibodies from rabbit sera by affinity chromatography, their characterization and
3 use in toxicological studies. *Toxicon* **24**: 273–283.
- 4 15. **Kuo, M.** 2006. Using a microscope: Basidia and cystidia.
5 http://www.mushroomexpert.com/microscope_cystidia.html.
- 6 16. **Kutchan, T. M.** 2005. A role for intra- and intercellular translocation in natural
7 product biosynthesis. *Curr. Opin. Plant Biol.* **8**:292–300.
- 8 17. **Largent, D. L., and D. Johnson.** 1977. *In* How to Identify Mushrooms to Genus
9 III: Microscopic Features. Mad River Press, Eureka, California.
- 10 18. **Luo, H., H. E. Hallen-Adams, and J. D. Walton.** 2009. Processing of the
11 phalloidin proprotein by prolyl oligopeptidase from the mushroom *Conocybe*
12 *albipes*. *J. Bio. Chem.* **284**:18070–18077.
- 13 19. **May, J. P., and D. M. Perrin.** 2007. Tryptathionine bridges in peptide synthesis.
14 *Pept. Sci.* **88**:714–724.
- 15 20. **McIntosh, J. A., Donia, M. S., and E. W. Schmidt.** 2009. Ribosomal peptide
16 natural products: bridging the ribosomal and nonribosomal worlds *Nat. Prod. Rep.*
17 **26**:537–559.
- 18 21. **McKnight, T. A., K. B. McKnight, and M. C. Skeels.** 2010. Amatoxin and
19 phallotoxin concentration in *Amanita bisporigera* spores. *Mycologia* **102**:763-5.
- 20 22. **Nagendran, S., H. E. Hallen-Adams, J. M. Paper, N. Aslam, and J. D. Walton.**
21 2009. Reduced genomic potential for secreted plant cell-wall-degrading enzymes
22 in the ectomycorrhizal fungus *Amanita bisporigera*, based on the secretome of
23 *Trichoderma reesei*. *Fung. Genet. Biol.* **46**:427–435.

- 1 23. **Nakamori, T, and A. Suzuki.** 2007. Defensive role of cystidia against
2 Collembola in the basidiomycetes *Russula bella* and *Strobilurus ohshima*. Mycol.
3 Res. **111**:1345–1351.
- 4 24. **Nolte, W. M., D. M. Tagore, W. S. Lane, and A. Saghatelian.** 2009.
5 Peptidomics of prolyl endopeptidase in the central nervous system. Biochemistry
6 **48**:11971–11981.
- 7 25. **Oman, T. J., and W. A. van der Donk.** 2010. Follow the leader: the use of
8 leader peptides to guide natural product biosynthesis. Nat. Chem. Biol. **6**:9–18.
- 9 26. **Roze, L. V., A. Chanda, and J. E. Linz.** 2010. Compartmentalization and
10 molecular traffic in secondary metabolism: A new understanding of established
11 cellular processes. Fung. Genet. Biol., in press. doi:10.1016/j.fgb.2010.05.006.
- 12 27. **Schneider, P., S. Bouhired, and D. Hoffmeister.** 2008. Characterization of the
13 atromentin biosynthesis genes and enzymes in the homobasidiomycete *Tapinella*
14 *panuoides*. Fung. Genet. Biol. **45**:1487–1496.
- 15 28. **Scott-Craig, J. S., D. G. Panaccione, F. Cervone, and J. D. Walton.** 1990.
16 Endopolygalacturonase is not required for pathogenicity of *Cochliobolus*
17 *carbonum* on maize. Plant Cell **2**:1191–1200.
- 18 29. **Szeltner, Z., and L. Polgár.** 2008. Structure, function and biological relevance of
19 prolyl oligopeptidase. Curr. Protein Pept. Sci. **9**:96–107.
- 20 30. **Truong, B. -N., K. Okazaki, T. Fukiharu, Y. Takeuchi, K. Futai, X. -T. Le,**
21 **and A. Suzuki.** 2007. Characterization of the nematocidal toxocyst in *Pleurotus*
22 subgen. *Coremiopleurotus*. Mycoscience **48**:222–230.

- 1 31. **Walton, J. D.** 2000. Horizontal gene transfer and the evolution of secondary
2 metabolite gene clusters in fungi: An hypothesis. *Fung. Genet. Biol.* **30**:167–171.
- 3 32. **Walton, J. D., D. G. Panaccione, and H. E. Hallen.** 2004. Peptide synthesis
4 without ribosomes. *In* J. Tkacz and L. Lange (ed.), *Advances in Fungal*
5 *Biotechnology for Industry, Agriculture, and Medicine*, p. 127–162. Kluwer
6 Academic, New York, NY.
- 7 33. **Walsh, C. T., and M. A. Fischbach.** 2010. Natural products version 2.0:
8 connecting genes to molecules. *J. Am. Chem. Soc.* **132**:2469-2493.
- 9 34. **Watkins, M., D. R. Hillyard, and, B. M. Olivera.** 2006. Genes expressed in a
10 turrid venom duct: divergence and similarity to conotoxins. *J. Mol. Evol.* **62**:247–
11 256.
- 12 35. **Welzel, K., K. Eisfeld, L. Antelo, T. Anke, and H. Anke.** 2005.
13 Characterization of the ferrichrome A biosynthetic gene cluster in the
14 homobasidiomycete *Omphalotus olearius*. *FEMS Microbiol. Lett.* **249**:157–163.
- 15 36. **Wieland, T.** 1986. *In* A. Rich (ed.), *Peptides of Poisonous Amanita Mushrooms*.
16 Springer, New York, NY.
- 17 37. **Zhang, P., Z. Chen, J. Hu, B. Ei, Z. Zhang, and W Hu.** 2005. Production and
18 characterization of amanitin toxins from a pure culture of *Amanita exitialis*.
19 *FEMS Microbiol. Lett.* **252**:223–228.
- 20

1 **FIGURE LEGENDS**

2

3 FIG. 1. (A) Mature basidiocarps of *Amanita bisporigera* used in this study (collected
4 in Ingham County, Michigan, in August, 2009). (B) Immature basidiocarps collected
5 before emergence from the ground. Intact basidiocarps are shown on the right, and
6 longitudinal sections through the same basidiocarps on the left. Structures: a, pileus (cap);
7 b, stipe (stem); c, universal veil, giving rise to the volva at the base of the stem as
8 indicated in (A); d, lamella (gill).

9

10 FIG. 2. Amanitin immunostaining of lamellae (gills) of *A. bisporigera* observed by
11 confocal laser scanning microscopy (CLSM). (A) Low-magnification CLSM view
12 showing cross-section of two lamellae and part of a third. (B) Higher magnification of a
13 single lamella by CLSM. (C) Same gill section as in (B) viewed by differential
14 interference contrast (DIC). (D) A different basidiocarp showing a cross-section of three
15 lamellae and parts of two others by CLSM. (E) Anti-amanitin immunostaining of basidia
16 (arrows), indicated by the sterigmata (the two slender projections), sterile cells (arrow
17 heads), and the sub-hymenium (CLSM superimposed on DIC). Sterigmata are the slender
18 projections from the basidia that bear the basidiospores, which are no longer present in
19 these sections. Sterile cells are the cells in the hymenium that, in *Amanita*, resemble
20 basidia but do not bear spores. (F) Same section as in (E) viewed with DIC alone. (G)
21 Anti-amanitin immunostaining of hymenium and sub-hymenium showing basidia (arrows)
22 with sterigmata and sterile cells containing amanitin (CLSM superimposed on DIC). (H)
23 Same section as in (G) observed by DIC. (I) Basidia (arrows) showing absence of

1 amanitin and adjacent sterile cells (arrow heads) showing presence of amanitin (CLSM
2 superimposed on DIC).

3

4 FIG. 3. Amanitin immunostaining of different tissues of *A. bisporigera* by LSM. (A)
5 Section of the pileus of the immature basidiocarp shown in Fig. 1B. (B) Section of a
6 mature pileus. (C) Same view as in (B) with white lines manually drawn to highlight
7 outlines and boundaries of the cells. (D) Higher magnification of an individual hypha of a
8 mature pileus (CLSM superimposed on DIC). (E) Section of stipe (stem) of a mature
9 basidiocarp. (F) Section of a universal veil (volva) of a mature basidiocarp (see Fig. 1A).

10

11 FIG. 4. Dual immunostaining of *A. bisporigera* lamellae using anti-amanitin and
12 anti-actin antibodies. (A – C) Lamella cross-sections: left to right show amanitin staining,
13 actin staining and the merge of the two. (D – F) Higher magnification views of a cross-
14 section of a lamella (same staining order as above). (G – H) Higher magnification views
15 of a cross-section of a mature pileus.

16

17 FIG. 5. Gene structure of *AbPOPA* and *AbPOPB* of *A. bisporigera*.

18

19 FIG. 6. DNA blotting of 13 *Amanita* species probed with (A) *AbPOPA* or (B)
20 *AbPOPB*. The species are: Lane 1, *A. aff. suballiacea*; lane 2, *A. bisporigera*; lane 3, *A.*
21 *phalloides*; lane 4, *A. ocreata*; lane 5, *A. novinupta*; lane 6, *A. flavoconia*; lane 7, *A.*
22 *porphyria*; lane 8, *A. franchetii*; lane 9, *A. muscaria*; lane 10, *A. gemmata*; lane 11, *A.*
23 *hemibapha*; lane 12, *A. velosa*; lane 13, unidentified species of *Amanita* sect. *Vaginatae*.

1 Mushrooms represent sections *Phalloideae* (1–4), *Validae* (5–8), *Amanita* (9 and 10),
2 *Caesareae* (11), and *Vaginatae* (12 and 13). Approximately the same amount of DNA
3 was loaded in each lane (see reference 10).

4

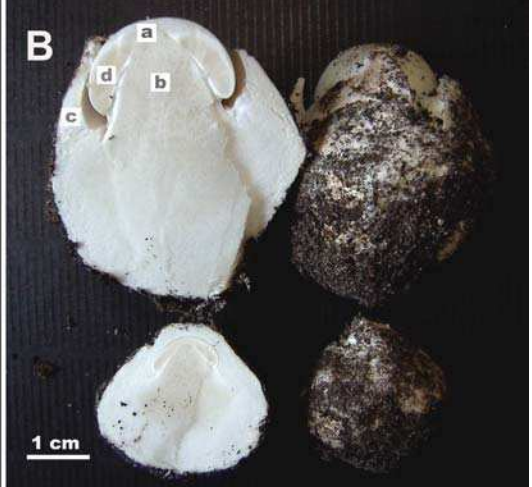
5 FIG. 7. Gene maps of lambda phage clones containing (A) *AbPOPA* or (B)
6 *AbPOPB*. Arrows indicate transcriptional direction. Genes were predicted using
7 FGENESH (www.softberry.com).

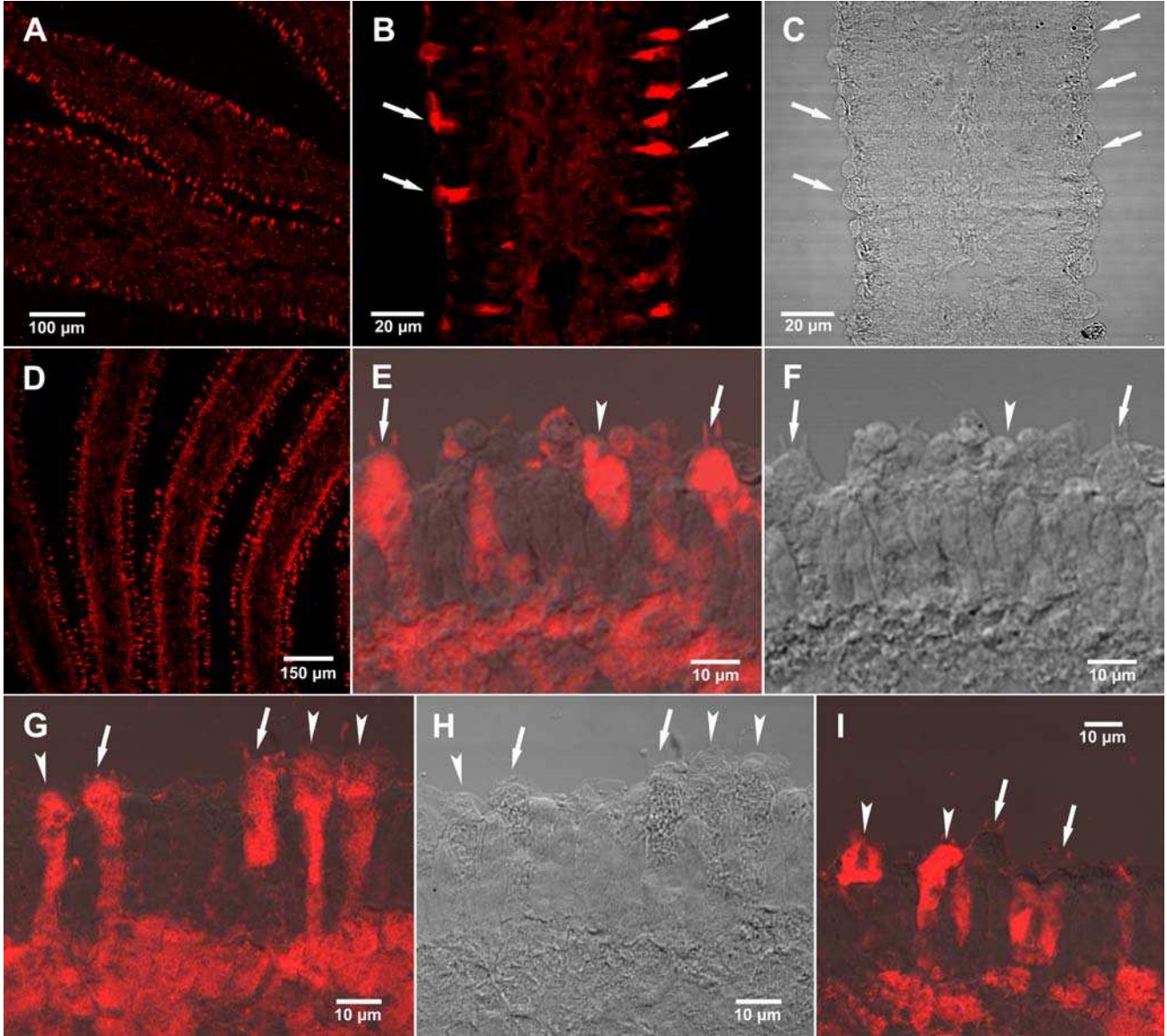
8

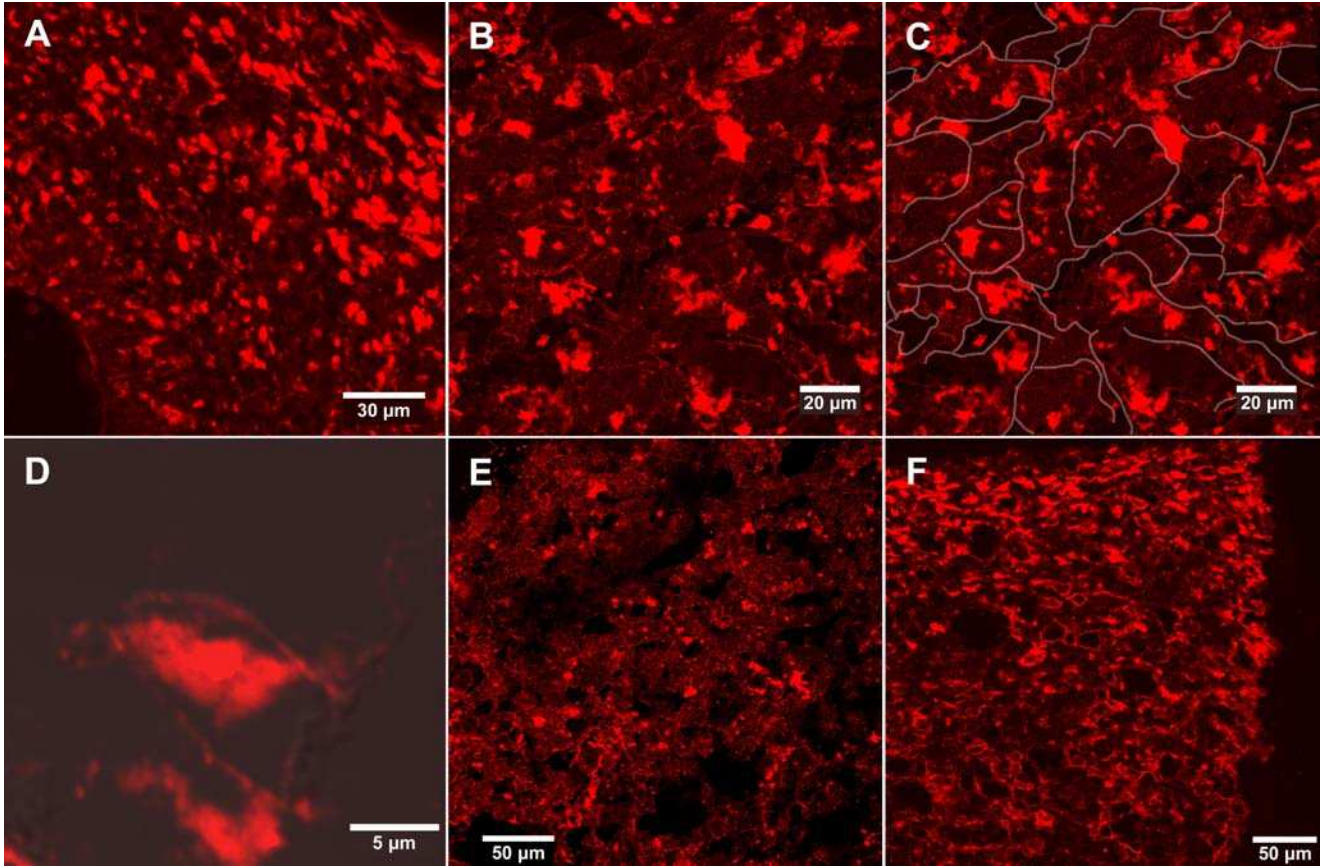
9 FIG. 8. Immunoblotting of *E. coli* inclusion bodies and total extracts of *A.*
10 *bisporigera* with anti-AbPOPB antibody. Lanes: 1, BL21 empty vector (PET21) control;
11 2, inclusion body (1 μ l = 0.05 μ g) of an *E. coli* BL21 line expressing AbPOPB; Lanes 3
12 through 6, total soluble protein from *A. bisporigera* loaded at 24 μ g, 47 μ g, 94 μ g and
13 141 μ g, respectively. All lanes were from the same gel, but some lanes between lanes 1
14 and 2 were removed. Size indicators (left) are in kDa.

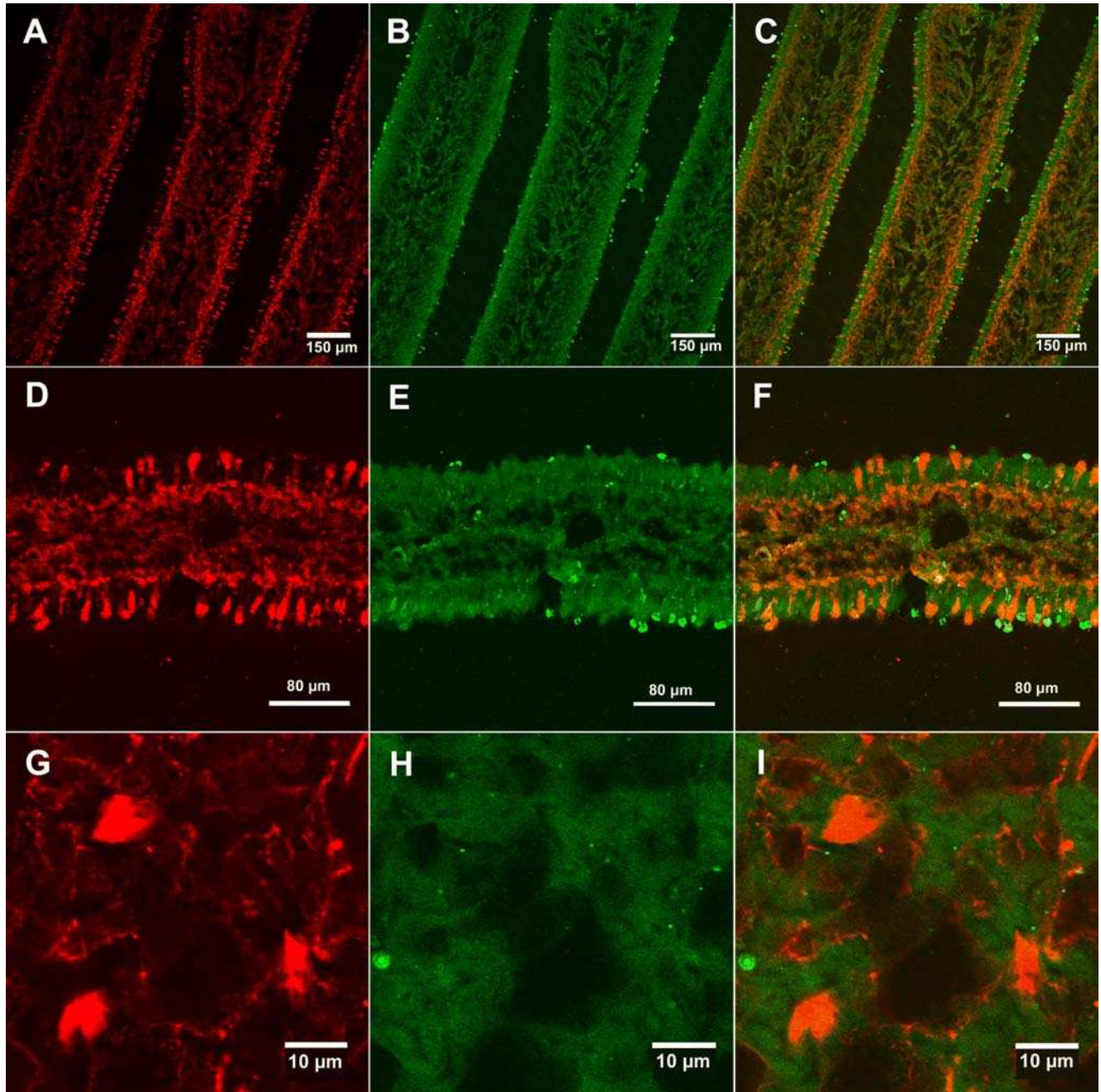
15

16 FIG. 9. Co-localization of amanitin and AbPOPB. (A – C) Co-localization in the
17 pileus. The stipe is on the left. From left to right: anti-amanitin, anti-AbPOPB, and merge
18 of (A) and (B). (D – F) Higher magnification of a portion of the pileus in (A – C). (G – I)
19 Co-localization in a single lamella, same order as (A – C).









AbPOPA



AbPOPB



0 1 2 kb

Key:

□ Exon — Intron ■ UTR

