Chlorella Virus PBCV-1 Encodes a Homolog of the Bacteriophage T4 UV Damage Repair Gene \textit{denV}

Masakazu Furuta  
\textit{University of Nebraska-Lincoln}

John Schrader  
\textit{University of Nebraska-Lincoln}

Holly Schrader  
\textit{University of Nebraska-Lincoln}

Tyler Kokjohn  
\textit{University of Nebraska-Lincoln}

Simon Nyaga  
\textit{University of Texas Medical Branch, Galveston, Texas}

\textit{See next page for additional authors}

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Authors
Masakazu Furuta, John Schrader, Holly Schrader, Tyler Kokjohn, Simon Nyaga, Amanda McCullough, R. Stephen Lloyd, Dwight Burbank, Dorit Landstein, Leslie C. Lane, and James L. Van Etten
DNA exposure to UV radiation produces two major photoproducts, cyclobutane pyrimidine dimers (CPDs) and pyrimidine-pyrimidone (6-4) photoproducts (5). Surviving UV irradiation requires the capacity to repair both of these dipyrimidine lesions, and there are three repair processes for this purpose, as follows. (i) Photoreversal directly converts dimers and 6-4 photoproducts to monomers. (ii) Nucleotide excision repair removes both of these by excising a nucleotide and replacing the DNA segment that contains the lesion. (iii) Base excision repair initiates incision of the glycosyl bond of the 5′ pyrimidine of the dimers (5). Solar radiation damages DNA-containing viruses which are common in freshwater. This gene functions in vivo and also when cloned in Escherichia coli. Photodamaged virus DNA can also be photoreactivated by the host chlorella. Since the chlorella viruses are continually exposed to solar radiation in their native environments, two separate DNA repair systems, one that functions in the dark and one that functions in the light, significantly enhance their survival.

**Materials and Methods**

Chlorella, viruses, and plasmids. The hosts for the chlorella viruses, Chlorella strain NC64A and Chlorella strain Pbi, were grown on MBBM medium (40) and FE5 medium (25), respectively. Procedures for producing, purifying, and plating virus PBCV-1 and the other chlorella viruses and the isolation of host and virus DNAs have been described previously (38, 40, 42). Three PBCV-1 DNA restriction fragments containing ORF A50L were subcloned from cosmid cYLS (16) into the polylinker region of pBluescript KS(+)(Stratagene) as indicated below (see Fig. 1B). The resultant plasmids were transformed into E. coli AB2480 (araA434), kindly provided by Mary Berlyn of the E. coli Genetic Stock Center at Yale University, and grown in LB medium (17) at 37°C. The PBCV-1 A50L gene probe used in hybridization experiments was amplified from cosmid cYLS by PCR (21). Oligonucleotide primers complementary to the 5′ and 3′ ends of the gene (accession no. U42580) were designed to introduce BamHI restriction sites at the translation initiation codon and immediately 3′ of the translation stop codon. The PCR product was cleaved with BamHI and cloned into the pUC19 BamHI site.

**UV radiation source and measurement.** Samples were irradiated with a 15-W General Electric G15T8 germicidal lamp, and doses were quantified with a UVX radiometer and a UV-25 probe (Ultraviolet Products). This lamp was operated for 100 h prior to performing experiments to stabilize the UV output. The lamp was illuminated for 10 to 15 min before exposing samples. Lamp output was determined for each experiment and was typically 0.1 J m−2 s−1. Dose was varied by changing the exposure time.

**UV protection assays.** (i) **Complementation of E. coli AB2480 by the PBCV-1 A50L gene.** The protocol was a modification of the methods used by Kokjohn and Miller (13) to quantify CFU. Cells were grown to the mid-log phase in LB medium (17) at 37°C (50 Klett units at 660 nm), harvested by centrifugation, suspended in an equal volume of 0.85% saline, and irradiated for various time periods.
All manipulations after irradiation were conducted in the dark. Irradiated cells were diluted in saline, and duplicate samples were plated on LB agar. Plates were incubated in the dark at 37 °C, and colonies were counted after 16 h.

(ii) UV resistance of PBCV-1 wild type and deletion mutants. The UV sensitivity of PBCV-1 and two deletion mutants of PBCV-1 lacking the A50L gene (viruses named P1050 and P1210 [14]) was measured by procedures similar to those used by Simonson et al. (31) for determining host cell reactivation of UV-irradiated bacteriophages. Virus lysates were diluted in 50 mM Tris-HCl (pH 7.8) and exposed to increasing doses of UV radiation. UV sensitivity was quantified by plaque assay. The titration plates were incubated either in the dark (dark repair conditions) or in the light (photoreactivation conditions) for 3 to 5 days.

Pyrimidine dimer-specific nicking activity assays. PBCV-1 virions were disrupted by sonication, and chloroplast cells were disrupted in a French press in a buffer containing 25 mM NaH2PO4 (pH 6.8), 100 mM KC1, 10 mM EDTA, and 100 μg of bovine serum albumin ml−1. Cell extracts were normalized by chromatophorl content. The extracts and 0.4 ng of a γ-32P-labelled, 5'-end-labelled duplex 49-mer oligonucleotide containing a centrally located cyclobutane pyrimidine dimer (gift from J. S. Taylor, Washington University, St. Louis, Mo.) were incubated in cell disruption buffer at 37 °C for 30 min. The reactions were stopped by adding buffer containing 95% formamide, 20 mM EDTA, 0.02% (wt/vol) bromophenol blue, and 0.02% (wt/vol) xylene cyanol. The samples were heated to 90°C for 5 min prior to electrophoresis on a 15% polyacrylamide-urea gel.

Other procedures. Radioactive DNA probes were prepared by nick translation with a kit from Bethesda Research Laboratories. For dot blots, virus DNAs were denatured and applied to nitrocellulose membranes as described previously (27). Total RNA was isolated and analyzed from PBCV-1-infected cells as described previously (29).

RESULTS

Complementation of E. coli by the PBCV-1 A50L gene. Figure 1A shows the predicted amino acid alignment of PBCV-1 ORF A50L with bacteriophage T4 endonuclease V. The two ORFs are about the same size and have 41% amino acid identity. Critical amino acids required for T4 endonuclease V structure and function are known from crystal (20) and co-crystal structures (44) and from site-directed mutations (3, 4, 11, 18, 28). These critical amino acids include the αNH2 group of Thr-2 which serves as a nucleophile (3, 28) and Glu-23 (4, 11, 18). The predicted amino acid sequence of ORF A50L has these two key amino acids.

To determine if the A50L gene product was active, three DNA constructs containing the A50L gene (Fig. 1B) were transformed into UV repair-deficient E. coli AB2480 (uvrA recA) (12) and tested for repair of UV damage to the bacterial genome. All three transformants survived UV radiation better
than a transformant with the plasmid vector alone (Fig. 2A). Thus, the PBCV-1 A50L gene, like the denV gene (15, 36), complements the UV repair deficiencies of this E. coli strain. Furthermore, like many chlorella virus genes (e.g., see reference 19), A50L has an upstream region that functions as a promoter in E. coli.

**PBCV-1 mutants that lack the A50L gene.** We recently isolated four PBCV-1 mutants with 27- to 37-kb deletions in the left end of the 330-kb genome (14). These mutants, which lack the A50L gene, replicate like the parent virus except for slightly reduced burst sizes. We compared the UV sensitivity of wild-type PBCV-1 and two deletion mutants designated P1050 and P1210 to determine if the A50L gene product functions in vivo. When infected cells were incubated in the dark (host cells grow heterotrophically in the dark and support virus replication), both deletion mutants were roughly 1,000-fold more sensitive to UV radiation than the wild-type virus was (Fig. 2B). In contrast, when infected cells were incubated in the light, which permits host photoreactivation of DNA damage, the deletion mutants were as UV resistant as the wild-type PBCV-1 virus (Fig. 2C).

**Appearance of A50L gene product during virus replication.** In their native environment, chlorella viruses are constantly exposed to solar radiation. Therefore, it seemed likely that the A50L enzyme might be packaged in the virions to initiate DNA repair immediately upon entry into the host cell. However, two experiments indicate that the enzyme is not packaged in the virion. (i) Sodium dodecyl sulfate-polyacrylamide gel electrophoresis of total proteins from PBCV-1 and PBCV-1 mutants lacking the A50L gene did not reveal a protein band of the expected size in wild-type virions that was absent in the deletion mutants, even after overloading the gel and silver staining (results not shown). (ii) Purified virions were disrupted by sonication, and these viral extracts were assayed for pyrimidine dimer-specific nicking activity with a 32P-labelled oligonucleotide that contains a site-specific cyclobutane pyrimidine dimer. There was no difference between the untreated oligonucleotide (Fig. 3, lane 2) and the oligonucleotide treated with the
infection and disappeared about 120 min after virus infection. Viruses, the A50L enzyme in size and amino acid sequence. The A50L enzyme is widespread among the chlorella viruses. The A50L gene probe did not hybridize to host chlorella DNA (Fig. 4B).

**DISCUSSION**

The following observations and experimental results support the hypothesis that the chlorella virus PBCV-1 A50L gene is a homolog of the *E. coli* bacteriophage T4 denV gene. (i) The predicted PBCV-1 A50L ORF has a Z-score of 34 and 41% amino acid sequence identity to endonuclease V, including critical amino acids known to be required for endonuclease V structure and function. (ii) Structural modelling studies between endonuclease V and the A50L gene product, with the X-ray crystal structure of endonuclease V as a guide, predict similar folding patterns between the two proteins (26a). (iii) Cloned DNA fragments containing the A50L gene complement the DNA damage repair defects of *E. coli* AB2480. (iv) Deletion mutants of virus PBCV-1 lacking the A50L gene are more susceptible to inactivation by UV irradiation. (v) Enzyme extracts from PBCV-1-infected cells, but not from uninfected cells, exhibit pyrimidine dimer-specific nicking activity. (vi) Repair activity of the A50L enzyme substantially overlaps that of the photoreactivation system, indicating that pyrimidine dimers are being removed. (vii) The A50L gene product is expressed as an early viral function. Taken together, these findings indicate that PBCV-1 encodes a homolog of the bacteriophage T4 denV gene and that this homolog is expressed and functions to eliminate UV-induced dimers in the viral genome during infection of the host alga. Thus, PBCV-1 is the first example of an algal virus encoding a DNA repair gene and only the second example of a virus of any type known to encode host-independent DNA UV repair functions.

Until the discovery of the PBCV-1 A50L gene, there were no known homologs of the T4 denV gene. Southern blot analysis of many independently isolated chlorella viruses (Fig. 4B) indicate that T4 denV gene homologs are common in this group of viruses. While the effects of convergent evolution in producing the apparent relationship between the T4 denV and PBCV-1 A50L genes cannot be discounted, the discovery of DNA repair gene homologs in unrelated viruses parasitizing hosts from different kingdoms suggests that these genes are ancient and widely disseminated in the biosphere. Indeed, Bernstein and Bernstein (1) have suggested that enzymes involved in DNA repair, recombination, and replication probably arose early and have been retained in biotic evolution. From an enzymological standpoint, the chlorella viruses are a new source of denV-like genes for genetic and structural studies.

UV damage to the PBCV-1 genome can also be repaired by the host photoreactivation system. Thus, PBCV-1 has access to two independent repair systems to reverse DNA damage, i.e., photoreactivation using host-encoded gene products and a virus-encoded enzyme that initiates dark repair. The combined activities of these repair systems should enable PBCV-1 to effectively exploit their hosts under a range of environmental conditions and to retain infectivity in environments exposed to substantial solar radiation. Since the algal host for these viruses is a photosynthetic organism, the viruses are obviously exposed to solar radiation in their natural environment. Photoreactivation enables infecting damaged viruses to rapidly and continuously repair pyrimidine dimers, ensuring effective viral repli-
cation under conditions in which the host is exposed to sunlight. The presence of a separate light-independent repair system permits DNA repair in the absence of photoreactivation. Thus, PBCV-1 can replicate whenever suitable hosts are encountered, day or night. This finding helps explain the frequent occurrence and occasional high titer of these viruses in fresh water collected throughout the world (39, 43, 48, 50).

UV repair systems are widely distributed in aquatic microorganisms, and evidence suggests that some of their viruses might also encode DNA repair genes. The effects of virus-encoded DNA repair genes will need to be considered in developing models of aquatic ecosystems. Marine surface waters contain high concentrations of virus particles, $10^7$ to $10^8$ ml$^{-1}$, although the majority of these particles are probably not infectious at any one time (6, 34). Most of these particles consist of viruses that infect bacteria, cyanobacteria, and algae. Viruses exist in a dynamic state in these environments and influence the composition, activity, and genetic diversity of microbial communities (6). For example, solar UV radiation rapidly inactivates bacteriophage and cyanophage with loss rates as high as 0.4 to 0.8 h$^{-1}$ in full sunlight (33, 35). Not surprisingly, infectivity disappears faster in the light than in the dark, leading to the suggestion that the concentration of infectious viruses in seawater may exhibit a strong diel signal (35). Suttle and Chen (35) have noted that sunlight inactivates bacteriophages at different rates, implying that bacteriophages differ in the ability to repair DNA damage. The capacity to repair UV-damaged DNA is clearly widespread among bacteriophages (10). Viruses may capitalize on available host-encoded repair systems, including photoreactivation, to eliminate UV damage (10, 47) or, as exemplified by bacteriophage T4 and virus PBCV-1, also encode novel, host-independent DNA repair functions. Presumably, viruses that can use two separate DNA repair systems have a better chance to survive in environments exposed to intense sunlight. Thus, it seems likely that many dsDNA-containing viruses found in environments subjected to solar radiation might encode DNA repair genes.

In summary, this study has three major implications. First, chlorella virus PBCV-1 and many, if not all, chlorella viruses encode a homolog of the bacteriophage T4 endonuclease V UV repair enzyme. Thus, the enzyme is not unique but a member of a widely distributed family. Second, the chlorella viruses will be a valuable source of endonuclease V homologs for structural and genetic studies. Third, the discovery of two independent repair systems, a host-encoded photorepair system and a combined virus- and host-encoded excision dark repair system, helps explain the ease with which chlorella viruses are recovered from surface waters throughout the world. Furthermore, similar mechanisms may help explain the surprisingly high concentrations of virus particles observed in marine surface waters.

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ADDENDUM IN PROOF

After acceptance of this paper, S. Shiota and H. Nakayama (Proc. Natl. Acad. Sci. USA 94:593–598, 1997) reported the characterization of a gene from *M. luteus* that also encodes a bacteriophage T4 endonuclease V-like enzyme. The PBCV-1-encoded enzyme is more similar to T4 endonuclease V than is the enzyme from *M. luteus*.

REFERENCES


