

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Publications, Agencies and Staff of the U.S.
Department of Commerce

U.S. Department of Commerce

12-12-1969

Blind River Dolphin: First Side-Swimming Cetacean

Earl S. Herald

Steinhart Aquarium, California Academy of Sciences, San Francisco

Robert L. Brownell Jr.

Steinhart Aquarium, California Academy of Sciences, San Francisco, rlbcetacea@aol.com

Fredric L. Frye

Steinhart Aquarium, California Academy of Sciences, San Francisco

Elkan J. Morris

Steinhart Aquarium, California Academy of Sciences, San Francisco

William Evans

Naval Undersea Research and Development Center, San Diego, California 92132

See next page for additional authors

Follow this and additional works at: <https://digitalcommons.unl.edu/usdeptcommercepub>



Part of the [Environmental Sciences Commons](#)

Herald, Earl S.; Brownell, Robert L. Jr.; Frye, Fredric L.; Morris, Elkan J.; Evans, William; and Scott, Alan, "Blind River Dolphin: First Side-Swimming Cetacean" (1969). *Publications, Agencies and Staff of the U.S. Department of Commerce*. 138.

<https://digitalcommons.unl.edu/usdeptcommercepub/138>

This Article is brought to you for free and open access by the U.S. Department of Commerce at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Publications, Agencies and Staff of the U.S. Department of Commerce by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Authors

Earl S. Herald, Robert L. Brownell Jr., Fredric L. Frye, Elkan J. Morris, William Evans, and Alan Scott

Blind River Dolphin: First Side-Swimming Cetacean

Abstract. *The blind river dolphin (Platanista gangetica), first written about by Pliny the Elder in A.D. 72, was found (10 November 1968) to be the first known side-swimming cetacean. The rudimentary eye lacks the lens, but anatomical evidence suggests that the eye may serve as a light sensor. The underwater sound emissions of this species, although similar to those of the Amazon River dolphin (Inia geoffrensis), appear to be produced constantly.*

As part of a study of the only river dolphin long recognized by native peoples as "blind" (*Platanista gangetica*), E. S. Herald, R. L. Brownell, Jr., F. L. Frye, and E. J. Morris spent 11 days on the Indus River near Sukkur, 515 km north of Karachi, West Pakistan (November 1968). We found that the blind dolphin has a variety of native names, the most common of which are buhlaan (West Pakistan) and susu (India).

The river dolphin family, Platanistidae, contains only four species; all are long-snouted and similar in appearance. The least known species occurs in Tung Ting Lake, China (*Lipotes vexillifer*). Another is widely distributed in the Amazon and Orinoco basins (*Inia*

geoffrensis), and the third occurs in the La Plata area (*Pontoporia blainvillei*). All three of these species have small eyes and apparently effective vision. Such is certainly true of the Amazon species which has recently become a public aquarium favorite. The fourth species, the blind river dolphin, is limited to the muddy waters of the Ganges, Indus, and Brahmaputra river systems. Its vestial eye is usually not visible externally, and its concealed location is shown only by a small opening in the skin usually smaller than that of the auditory meatus. Our anatomical studies have shown that the animal is not totally blind.

We obtained the dolphins from Sindi fishermen who use them for food. They are captured at night with a throw net, and a living catfish is used for bait. Three young females were taken: No. 1 on 4 November (total length, 121 cm; 25.7 kg); No. 2 on 7 November (115 cm; 21.2 kg); and No. 3 on 8 November (107 cm; 19.5 kg). With the exception of abrasions on the lower jaw due to capture and subsequent tethering, the dolphins were remarkably free of injuries; even scratch marks were absent.

When the new arrivals were placed in our holding pond on the Indus River, after surface breathing or blowing they started to roll the body as they swam

downward. Because of the very muddy water, we could not see what was happening under the surface, but we did think that they were swimming upside down, as is sometimes done by the related Amazon freshwater dolphin, *Inia geoffrensis*. Later at Karachi (10 November 1968) when the dolphins were in the clear water of a small swimming pool for the first time, we were amazed to observe that they were swimming on their sides—a method of progression previously unknown among cetaceans.

The 25.7-kg female swam on her left side in a clockwise direction, whereas the 21.2- and 19.5-kg females swam on their right sides in a counterclockwise direction. While the dolphins were side-swimming, the pectoral flipper (right or left, depending upon swimming direction) either touched the bottom or trailed about 2 to 3 cm above it. The tail was higher in the water than the head, so that the body was at an oblique angle with respect to the bottom of approximately 10°. Thus the head is aimed downward, and moves constantly. If the susu were vertical in the water, the head would be bobbing up and down; but since the dolphin is on its side, the head motion becomes a lateral sweep over the bottom.

These three blind dolphins traveled from Sukkur to San Francisco in 5 days, a distance of some 17,721 km. Two of these days were spent in recuperative swimming, one at Karachi and one at Tokyo. At both these localities the susu showed their normal side-swimming pattern. Although they were continually presented with a wide variety of living and dead food, they would accept none. Consequently they were placed on a force-feed schedule.

In 1878, Anderson showed that *Platanista* lacked the crystalline lens of the eye (1). Our dolphins showed no reaction to drastic changes in light intensity, even when the initial intensity was at low illumination levels where rod activity would be seen. However, our anatomical study shows that the eye may be capable of serving as a light receptor, even if it is incapable of forming a clear image on the retina because of the flat shape of the cornea and the absence of an effective lens, with degeneration and absorption of the lens fiber. The retina is adapted to light-gathering rather than image-resolving, with a very densely packed receptor layer, a rather scanty bipolar and ganglion cell layer, and an extremely small optic nerve containing only a few hun-

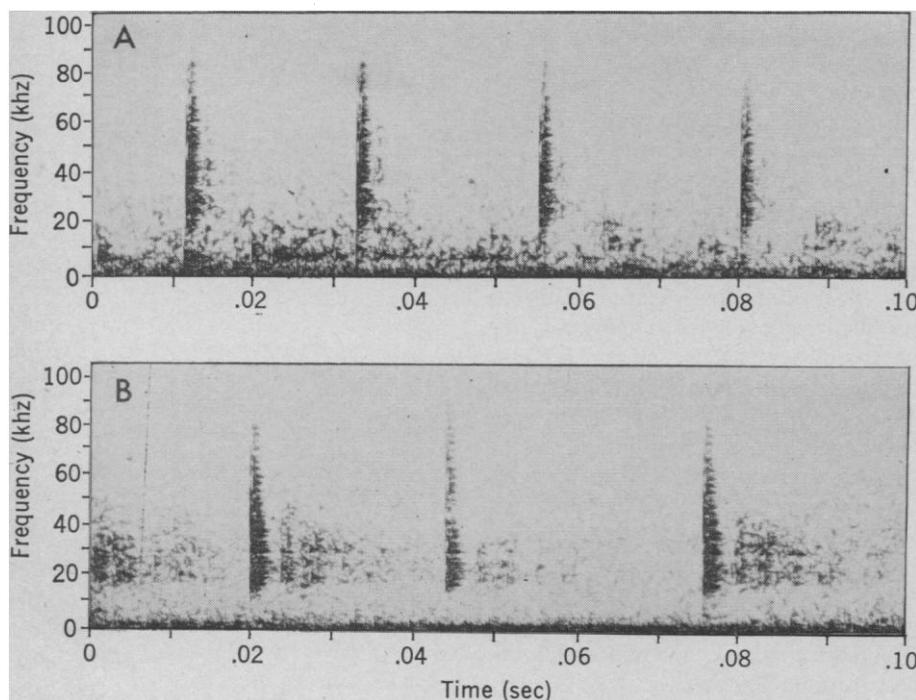


Fig. 1. Pulse trains produced by *Inia geoffrensis* (A) and *Platanista gangetica* (B). The ambient sound pressures in aquariums varied from 10 to 15 db referred to 1 μ bar. The animals were oriented with their rostrums pointed directly at the hydrophone at a distance of approximately 1 m (filter bandwidth, 480 Hz).

dred fibers; the pigment epithelium layer, which prevents light scatter within the eye itself, is absent. Because of the dense pigmentation of the overlying skin, light can reach the retina only through the pinhole sphincter-like lid structure. A cone-shaped muscle layer extends from the posterior orbit to the circumference of the skin overlying the eye, and its contraction would clearly open this sphincter; a definite circular sphincter-like arrangement of muscle fibers in the tissue certainly is the anatomical substrate for closure. Thus, the structure is present for sensing light and, maybe, even for telling its direction. Possibly the creature does use vision at night or at a seasonal migration or breeding time, although no utility is seen for vision in the muddy habitat where *Platanista* is usually found.

Echolocation in cetacea has been experimentally validated in only two species, *Tursiops truncatus* and *Phocoena phocoena* (2). However, every species of odontoceti studied thus far produces trains of clicks similar in many ways to those used by species known to echolocate (3). The blind river dolphin can now be added to this list. While the dolphins were in captivity at the Steinhart Aquarium, we monitored and recorded the sound emissions of all three *Platanista* as a group and of each individually, using a hydrophone (Chesapeake model CH26B), a matched high-gain amplifier system, and an instrumentation tape recorder (Lockheed model 417). The frequency response of the entire system at a tape speed of 30 inch/sec was flat (± 3 db from 100 hz to 100 khz).

The underwater sounds of the *Platanista* consisted of trains of pulses which were produced constantly during the sound-monitoring periods. The pulses were produced at a rate of 20 to 50 per second. The amplitude of the pulses was strongly dependent on the animals' orientation with respect to the hydrophone. When the dolphins were swimming toward the hydrophone, the amplitude of the emitted signal would drop by 8 to 10 db as the animal's rostrum would swing 10° on either side of the transducer. When the orientation of the rostrum was more than 40° away from the hydrophone, the signal level was reduced by 15 to 20 db. During the entire 16 hours of monitoring, there were no periods in which pulse trains were not observed. Variation in light intensity and the

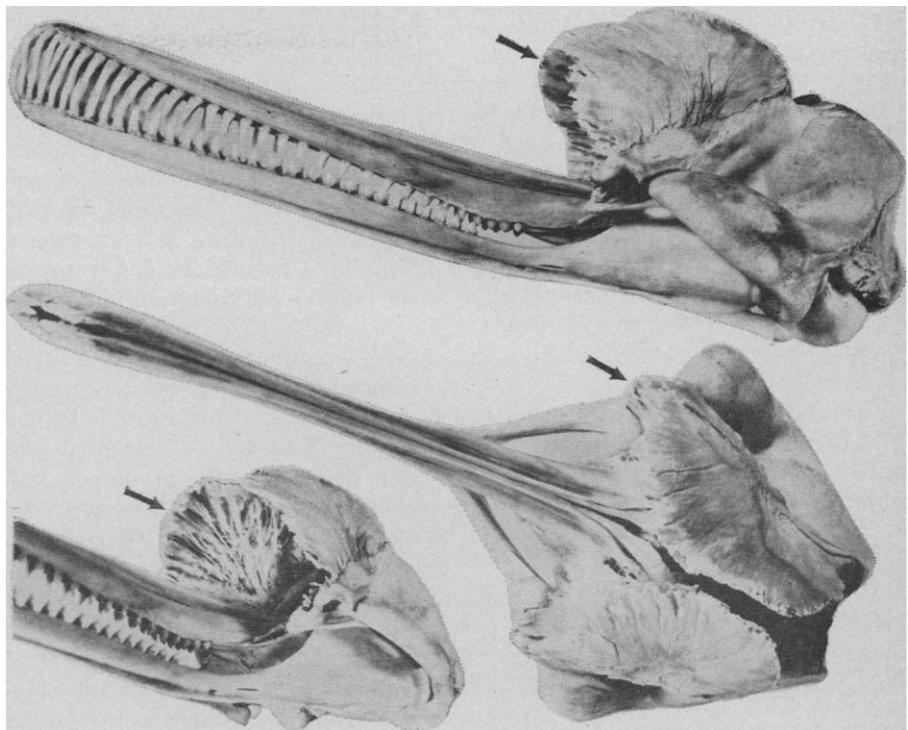


Fig. 2. Lateral, dorsal, and antero-oblique views of the skull of *Platanista gangetica*, showing the nature of the maxillary extensions (arrow).

presence of objects in the tank, for example, hydrophone or wood or pipe barriers, did not appear to influence the animal's sound emissions.

By contrast, the related Amazon dolphins, *Inia geoffrensis*, in an adjacent tank produced trains of pulses only during orientation toward food or strange objects, or both, placed in their tank. The pulse trains produced in these cases were similar in amplitude, repetition rate, and dependency on orientation to those emitted by *Platanista*. However, when the light intensity was reduced, *Inia* responded by more frequent sound emissions.

The wave form and frequency content of individual pulses produced by both *Platanista* and *Inia* are quite similar, with maximum energy between 15 and 60 khz (Fig. 1). The pulses produced by both of these freshwater odontoceti have considerably less energy below 15 khz than has been observed in most other delphinids. This could partially account for the observation reported by Norris (2) that *Inia* produce low-intensity signals. Most previous recordings of pulses produced by *Inia* have been made on systems with limited frequency response, that is, 50 hz to 20 khz. Unfortunately, the short time available for acoustic observation of this unique cetacean did not allow for any definitive test of

echolocation capability. The degenerate nature of the eye, constant pulse emission, and obstacle-avoidance behavior, both in the natural murky habitat of the dolphin and in captivity, suggest that *Platanista* is an effective echolocator.

Among cetaceans, the blind river dolphin has a unique skull in that there are broad maxillary flanges which extend upward into the forehead melon (Fig. 2). These paired flanges do not meet in the center dorsally, thus leaving a variable gap of about 5 to 10 mm (21.2-kg female). Although the outside of each flange is smooth, the inside supporting structure has a very intricate, radial, weblike appearance. Based upon the work of Evans *et al.* (4), which indicates that the topography of the delphinid skull has an effect on the sound field, we suspect that the *Platanista* maxillary flanges act as acoustic baffles to direct the sonic pulses into a narrow beam.

From the time of capture, blind dolphin No. 1 survived for 38 days, No. 2 for 24 days, and No. 3 for 44 days. Primary cause of death for the two larger animals was pneumonic infection; for the smallest dolphin the injury to the lower jaw at the time of capture was deemed responsible; other factors may also have been involved.

The three complete skeletons that have been prepared are the first from

the Indus River; the two larger skeletons are located at the California Academy of Sciences (CAS 14921 and 14922), and the smallest is at the Museum of Comparative Zoology (MCZ 52306).

EARL S. HERALD

ROBERT L. BROWNELL, JR.

FREDRIC L. FRYE, ELKAN J. MORRIS
*Steinhart Aquarium, California
Academy of Sciences, San Francisco*

WILLIAM E. EVANS

Naval Undersea Research

and Development Center,

San Diego, California 92132

ALAN B. SCOTT

Institute of Medical Sciences,

Pacific Medical Center,

San Francisco, California 94115

References and Notes

1. J. Anderson, *Anatomical and Zoological Researches: Comprising an Account of the Zoological Results of the Two Expeditions to Western Yunnan in 1868 and 1875* (Quaritch, London, 1878), vol. 1, pp. 417-550; vol. 2, pp. 25-26, 28-32, 34-41.
 2. K. S. Norris, in *The Biology of Marine Mammals*, H. T. Anderson, Ed. (Academic Press, New York, 1969), pp. 393-400, 416-417.
 3. W. E. Evans, in *Marine Bio-Acoustics*, W. N. Tavolga, Ed. (Pergamon, Oxford, 1967), vol. 2, pp. 159-186.
 4. W. E. Evans, W. W. Sutherland, R. G. Beil, in *Marine Bio-Acoustics*, W. N. Tavolga, Ed. (Pergamon, Oxford, 1964), vol. 1, pp. 353-372.
- 1 April 1969; revised 16 September 1969

Intracranial Drug Implants: An Autoradiographic Analysis of Diffusion

Abstract. *Labeled crystalline atropine, administered to the hypothalamus of rats, remained strictly localized in a sphere, 1.0 to 1.8 millimeters in diameter, during the first 3 minutes. A similar distribution obtained after 1 hour. At intermediate times, slightly elevated radioactivity, reflecting concentrations 2,000 to 10,000 times below behaviorally effective doses, was observed several millimeters from the implantation site.*

Local changes in the concentration of neurohumors and related substances have been used to selectively activate or inactivate functionally defined pathways in areas where the proximity of different neural systems has limited the effectiveness of traditional techniques (1). The usefulness of drug-injection procedures depends on the degree of localization achieved. MacLean demonstrated (2) that the injection of even small quantities of fluid under pressure produces fairly extensive diffusion. We

have attempted to circumvent the problems inherent in this technique by permitting the drug to go into solution in the brain, using the extracellular fluid as the solvent (3). This approach has its own problems in that dosage parameters are difficult to estimate, but our behavioral and electrophysiological data consistently show that diffusion is limited to a sphere about 1.0 mm in diameter. Dye-diffusion and autoradiographic studies have confirmed this estimate (4). Autoradiography, however, is limited by spread of radioactivity during histological processing and film mounting. This type of potential artifact is obviated in the dry-mount procedure with unfixed and unembedded freeze-dried sections (5), as applied in our experiment.

It is possible to obtain a map of positive and negative injection sites and thus to establish directly the extent of effective diffusion. However, the ubiquity of positive placements in some regions of the brain makes such an approach difficult for some drugs, and it has been suggested (6) that a ventricular distribution of these drugs may account for at least some of their effects. Such an interpretation may apply to the observation that an implant of atropine at any one of several carbachol-sensitive sites blocks the drinking which is normally elicited by the administration of carbachol (carbamylocholine chloride) to sites in the hypothalamus, preoptic area, septal area, and hippocampus (7). Contralateral as well as ipsilateral interacting sites have been described, and it is hard to account for the interaction in terms of an atropine blockade of direct neural projections to the carbachol implantation site.

Since the ventricular distribution hypothesis limits the usefulness of all procedures for injection of drugs into the central nervous system, we decided to use an autoradiographic method which makes it possible to limit spread of radioactivity after the tissue is dead (5). We studied the pattern of atropine diffusion under conditions which produce a blockade of the behavioral effects of intracranial carbachol.

In the first experiment, cannulas were implanted stereotaxically into the lateral hypothalamus of ten rats. One week after surgery, the placements were shown to be sensitive to carbachol, since the administration of 0.5 to 5.0 μg of carbachol to the lateral hypothalamus elicited an intake of at least 10 ml of water. At least 24 hours later, 1.0 to

5.0 μg of ^3H -labeled atropine (8) was administered to the same site. The animals were decapitated 2 to 16 minutes later (the behavioral effects of centrally applied atropine typically appear within 1 to 2 minutes).

In subsequent experiments, cannulas were implanted stereotaxically into the medial septal area as well as into the lateral hypothalamus of 14 rats. Both placements were shown to be carbachol-sensitive by the elicitation of a water intake of at least 10 ml within 30 minutes after the application of 0.5 to 5.0 μg of carbachol. At least 24 hours later, 0.5 to 5.0 μg of carbachol was again applied to the septal area of all rats. Thirty seconds after the animals began to drink, 1.0 to 5.0 μg of labeled atropine was placed into the lateral hypothalamus. The animals were then returned to the test situation and permitted to drink. Water intake ceased within 1 to 4 minutes after the implantation of atropine. The animals were decapitated at least 1 minute after the water intake ceased (dropped to zero). To obtain a representative sample of the drug diffusion, animals were decapitated 2, 3, 5, 6, 7, 8, 9, 12, and 60 minutes after the application of ^3H -atropine. After the animals were decapitated, the area surrounding the tips of both cannulas was excised while the brain was kept on a petri dish cooled on ice. Within 5 minutes after the decapitation, areas of the brain were frozen by immersion in liquid nitrogen. Drug diffusion may have continued between decapitation and freezing. The frozen tissue was used for (i) the preparation of 2- μm freeze-dried sections for dry-mount autoradiography or (ii) liquid-scintillation counting of radioactivity in blocks of tissue weighing approximately 10 mg which encompassed the site of atropine implantation. The radioactivity of comparable blood samples, taken at the time of decapitation, was also determined by liquid-scintillation counting. The autoradiograms were evaluated by counting silver grains and by obtaining densitometric estimates at exposure times from 2 weeks to 2 months.

The results (Fig. 1) are remarkably consistent, in view of the wide range of drug action and exposure times, in showing that the application of labeled atropine in crystalline form produced a pattern of intense radioactivity which remained restricted, even after 1 hour, to a sphere of about 1.0 to 1.8 mm in diameter. There was, in many cases,