

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Entomology papers

Museum, University of Nebraska State

3-10-2021

Defining and validating regenerative farm systems using a composite of ranked agricultural practices

Tommy L.D. Fenster
California State University East Bay

Claire E. LaCanne
University of Minnesota

Jacob R. Pecenka
Purdue University

Ryan B. Schmid
Blue Dasher Farm, Ecdysis Foundation

Michael M. Bredeson
Blue Dasher Farm, Ecdysis Foundation

See next page for additional authors

Follow this and additional works at: <https://digitalcommons.unl.edu/entomologypapers>

Fenster, Tommy L.D.; LaCanne, Claire E.; Pecenka, Jacob R.; Schmid, Ryan B.; Bredeson, Michael M.; Busenitz, Katya M.; Michels, Alex M.; Welch, Kelton D.; and Lundgren, Johnathan G., "Defining and validating regenerative farm systems using a composite of ranked agricultural practices" (2021). *Entomology papers*. 165.
<https://digitalcommons.unl.edu/entomologypapers/165>

This Article is brought to you for free and open access by the Museum, University of Nebraska State at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Entomology papers by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Authors

Tommy L.D. Fenster, Claire E. LaCanne, Jacob R. Pecenka, Ryan B. Schmid, Michael M. Bredeson, Katya M. Busenitz, Alex M. Michels, Kelton D. Welch, and Johnathan G. Lundgren



RESEARCH ARTICLE

Defining and validating regenerative farm systems using a composite of ranked agricultural practices [version 1; peer review: 2 approved]

Tommy L.D. Fenster^{1,2}, Claire E. LaCanne^{1,3}, Jacob R. Pecenka^{1,4}, Ryan B. Schmid¹, Michael M. Bredeson¹, Katya M. Busenitz^{1,5}, Alex M. Michels^{1,6}, Kelton D. Welch¹, Jonathan G. Lundgren ¹

¹Blue Dasher Farm, Ecdysis Foundation, Estelline, South Dakota, 57234, USA

²California State University East Bay, Hayward, California, USA

³Center for Agriculture, Food, and Natural Resources, University of Minnesota Extension, Minneapolis, Minnesota, USA

⁴Department of Entomology, Purdue University, West Lafayette, Indiana, USA

⁵Department of Entomology, University of Nebraska, Lincoln, Nebraska, USA

⁶Department of Natural Resource Management, South Dakota State University, Brookings, South Dakota, 57007, USA

V1 First published: 15 Feb 2021, 10:115
<https://doi.org/10.12688/f1000research.28450.1>

Latest published: 15 Feb 2021, 10:115
<https://doi.org/10.12688/f1000research.28450.1>

Abstract

Background: Ongoing efforts attempt to define farms as regenerative to aid marketers, policymakers, farmers, etc. The approach needs to balance precision with function, and must be transparent, simple, scalable, transferable, incorruptible, and replicable.

Methods: We developed practice-based scoring systems to distinguish regenerative cropland and rangeland, and validate them based on whether these scores scaled with regenerative goals on actual farm operations. Study systems included cornfields of the Upper Midwest, almond orchards of California, and rangeland systems of the Northern Plains. Response variables included soil carbon and organic matter, soil micronutrients, water infiltration rates, soil microbial communities, plant community structure, invertebrate community structure, pest populations, yields, and profit.

Results: Regenerative outcomes were strongly correlated with our approach to farm scoring. Soil organic matter, fine particulate organic matter, total soil carbon, total soil nitrogen, phosphorous, calcium and sulfur all increased alongside regenerative matrix scores in one or both of the cropping systems. Water infiltration rates were significantly faster in more regenerative almond orchards. Soil bacterial biomass and Haney soil health test scores were higher as cropland incorporated more regenerative practices. Plant species diversity and biomass increased significantly with the number of regenerative practices employed on almonds and rangelands. Invertebrate species diversity and richness were positively associated with regenerative practices in corn, almonds, and rangelands,

Open Peer Review

Reviewer Status  

Invited Reviewers

1

2

version 1

15 Feb 2021



report



report

1. **John M. Holland** , Game and Wildlife Conservation Trust, Fordingbridge, UK
Jenny Bussell, Game and Wildlife Conservation Trust, Leicestershire, UK

2. **Richard W. Teague**, Texas A&M AgriLife Research, Vernon, USA

Any reports and responses or comments on the article can be found at the end of the article.

whereas pest populations and almond yields were unaffected by the number of regenerative practices. Corn yields were negatively associated with more regenerative practices, while almond yields were unaffected by the number of regenerative practices. Profit was significantly higher on more regenerative corn and almond operations.

Conclusions: Our scoring system scaled positively with desired regenerative outcomes, and provides the basis for predicting ecosystem responses with minimal information about the farming operation. Natural clusters in the number of regenerative practices used can be used to distinguish regenerative and conventional operations.

Keywords

Biodiversity, Haney Test, Invertebrates, Regenerative Agriculture Label, Soil Carbon, Soil Health, Soil Microbiology

Corresponding author: Jonathan G. Lundgren (jonathan.Lundgren@ecdysis.bio)

Author roles: **Fenster TLD:** Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Writing – Original Draft Preparation, Writing – Review & Editing; **LaCanne CE:** Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Writing – Review & Editing; **Pecenka JR:** Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Writing – Review & Editing; **Schmid RB:** Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Writing – Original Draft Preparation, Writing – Review & Editing; **Bredeson MM:** Conceptualization, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Busenitz KM:** Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Michels AM:** Conceptualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Welch KD:** Conceptualization, Data Curation, Investigation, Writing – Original Draft Preparation, Writing – Review & Editing; **Lundgren JG** : Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Project Administration, Resources, Software, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This work was funded in part by USDA-Pest Management Alternatives Program [Grant 2013-34381-21245], NCR SARE [Grants GNC16-227, GNC15-207, LNC18-410], and W SARE [Grant GW19-193], Globetrotter Foundation, and Ecdysis Foundation. *The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

Copyright: © 2021 Fenster TLD *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Fenster TLD, LaCanne CE, Pecenka JR *et al.* **Defining and validating regenerative farm systems using a composite of ranked agricultural practices [version 1; peer review: 2 approved]** F1000Research 2021, 10:115 <https://doi.org/10.12688/f1000research.28450.1>

First published: 15 Feb 2021, 10:115 <https://doi.org/10.12688/f1000research.28450.1>

Introduction

The term “regenerative agriculture” was first coined by [Robert Rodale \(1983\)](#). The name highlighted how industrialized agriculture was severely reducing its natural resource base, and that without rebuilding that natural resource base, “sustainable agriculture” and “conservation agriculture” were insufficient for supporting the food and natural resource needs of a growing human population. Regenerative agricultural systems increase soil health and promote biodiversity while producing nutritious food profitably. Some outcomes of a regenerative operation are improved greenhouse gas relationships, balanced water relationships, reduced pollution from agrichemicals, increased resiliency of farms, more nutritionally robust foods, increased ecosystem services, etc. ([Elevitch et al., 2018](#); [Francis et al., 1986](#); [Sherwood & Uphoff, 2000](#)). A central challenge for researchers, policy makers, consumers, etc. is defining what a regenerative farm is and having a standardized method for discerning them from conventional farms ([Newton et al., 2020](#); [Schreefel et al., 2020](#)).

The principles and philosophy that define regenerative systems have been well formulated by practitioners and educators ([Lal, 2020](#); [Schreefel et al., 2020](#); [UnderstandingAG, 2020](#); [USDA-NRCS, 2020](#)). Essentially, regenerative practices can be distilled down to two central principles: 1) reduce uniform disturbance (such as tillage and agrichemical use), and 2) increase diversity (biodiversity, and the diversity of revenue streams from an operation). The first working formula for these principles was proposed by [LaCanne & Lundgren \(2018\)](#) and included four principles: 1) reduce or eliminate tillage, 2) never leave bare soil, 3) maximize plant diversity and productivity on a farm, and 4) integrate livestock and cropping operations. Here, we add a fifth principle: 5) reduce or eliminate synthetic agrichemicals. In croplands, practices such as no-till, diverse crop rotations lasting more than four seasons, cover cropping, intercropping and interseeding, and livestock integration all contribute to a regenerative farming model ([LaCanne & Lundgren, 2018](#); [Rhodes, 2017](#)). In rangelands, practices that actualize regenerative principles include reduction/elimination of synthetic agrichemicals and adaptive multi-paddock grazing systems that allow a rangeland to rest following the punctuated disturbance of grazing ([Teague & Kreuter, 2020](#); [Wagner et al., 2020](#)). Regenerative farms are also diverse and complementary in their enterprises, and adaptive in their management choices, ensuring that a farm is resilient and profitable in the face of adversity. These practices are dependent upon one another within a system for them to be optimally successful, and it is the system, not the individual practices, that drives the success of an operation.

Classifying regenerative farming operations has to balance precision with function. Adaptability and innovation are hallmarks of operations that are guided by regenerative principles. Therefore, while a matrix of responses or practices must have sufficient nuance to accurately reflect a regenerative system, it cannot be a rigid formula that inhibits that critical adaptability and ongoing innovation. Nor should a scoring system be so complex as to exclude adoption by regenerative farmers. Classification schema have been developed for other movements

within ecological agriculture, including conservation agriculture ([Hendrix et al., 1986](#); [Kassam et al., 2012](#)), organic agriculture ([USDA-AMS, 2020](#)), holistic management ([Alfaro-Arguello et al., 2010](#)), mob grazing ([Gurda et al., 2018](#)), etc. In these systems, the tangible manifestations of core principles are represented by a series of standards or practices. An inherent risk is that regenerative agriculture will follow the same pattern, and come to be defined by specific standards or practices, rather than by the principles and goals of a regenerative system. There are a growing number of certification programs and definitions for regenerative farming that capture key themes of regenerative agriculture and assemble various practices that may produce intended outcomes ([AGW, 2020](#); [Carbon_Underground, 2020](#); [HMI, 2020](#); [ROC, 2020](#); [TerraGenesis, 2020](#)). In the end, any labelling or certification system must be validated empirically to determine whether it results in the intended outcomes.

Regenerative agriculture needs to be defined by principles and supported by empirical data from farms displaying a wide range of management practices to ensure the implemented principles are achieving regenerative farming goals. The approach to defining a regenerative system must be transparent, simple, scalable, transferable, incorruptible, and replicable. Our team has created a series of questions for land managers that encapsulate the principles that drive regenerative agriculture as defined by the farmers and ranchers ([Fenster et al., 2021](#); [LaCanne & Lundgren, 2018](#); [Pecenka & Lundgren, 2019](#); [Schmid & Lundgren, 2021](#)). Each answer receives a score, and higher scores are indicative of more regenerative operations. A regenerative matrix score is attained by summing the scored results to these questions. The first goal of this paper is to determine whether key response variables pertaining to soil health, biodiversity, ecosystem function, and farm performance ([Elevitch et al., 2018](#); [Schreefel et al., 2020](#)) scale with the assignment of scores from our regenerative matrix. Essentially, does a higher regenerative score in cropland and rangeland consistently scale positively with the desired outcomes of regenerative farming? The second goal of this paper is to determine whether there is a threshold score that can be used to distinguish between regenerative and conventional systems? If this matrix performs these functions, then this approach could be used to advance the field of regenerative agriculture by providing a clear, scalable and transferable approach to characterizing a regenerative farm.

Methods

Two independent scoring systems based on a narrow suite of practices were used to define regenerative cropland and rangeland systems. Three food systems were examined in this project, corn in the Upper Midwest ([LaCanne & Lundgren, 2018](#)), almond orchards in California ([Fenster et al., 2021](#)), and rangelands of the Northern Plains ([Pecenka & Lundgren, 2019](#); [Schmid & Lundgren, 2021](#)). Farms included in these studies represented successful, established systems and were in their respective farming philosophies for at least three years. Some of the farms were targeted for their reputation as being leaders in regenerative farming, but the resulting fields and ranches in reality represented a wide continuum from very conventional to very regenerative farming systems.

We examined different suites of response variables that are considered representative of a regenerative system. In croplands (Table 1), the elimination of tillage, maintaining ground cover through planting cover crops or fostering resident vegetation, planting hedgerows, and use of organic amendments (compost, manure, mulch, compost teas), and grazing, were all considered regenerative (LaCanne & Lundgren, 2018; Soto *et al.*, 2021). Tillage, maintaining bare soil, and spraying synthetic insecticides, herbicides, fungicides, use of chemical fertilizers were all considered conventional practices. Regenerative practices were scored as 1, and conventional practices were scored as 0. In rangelands (Table 2), management systems were defined by their stocking density, rotation frequency, the duration that the pasture rested following grazing, and use of ivermectin products. Cattle operations were categorized based upon their combination of these practices to form the different systems (Colley *et al.*, 2019; Teague & Barnes, 2017). Use of ivermectin in the ranches was categorized as high (multiple applications during a year; scored as 0), low (single annual use, not applied during grazing period; scored as 1) and no ivermectin (scored as 2). Operations' stocking densities (animal units [AU] per ha), were categorized as fewer than 5 animal units (AU) per ha (scored as 0), 5–10 AU per ha (scored as 1), and more than 10 AU per ha (scored as 2). Operations were categorized as having a rotation frequency of 30 d or more (scored 0), between 10–30 d (scored as 1), and less than 10 d (scored as 2). Finally, rest periods on ranches were scored as continuously grazed (no rest; scored as 0), allowed to rest 1 > and < 30 d (1), or >30 d (2) over a growing season. A simple questionnaire to obtain the necessary information to populate our matrices can be found as in *Extended data* (Lundgren, 2021).

The study systems

Corn was examined on 20 farms (each with four fields) in North Dakota, South Dakota, Minnesota, and Nebraska in 2015 and 2016. Fields were a minimum of 4 ha in size, and the sampling area in each field was 61 × 61 m. All samples were taken at least 12 m into the field to minimize border effects. Only three of these farms were certified organic. Genetically modified (Bt hybrid) corn was universally treated with neonicotinoid seed treatments and was regarded as insecticide-treated. In corn, we examined soil organic matter (SOM), fine particulate organic matter (fPOM), soil bulk density, water infiltration, invertebrate communities in the soil, on the soil surface and in the plant canopy, pest abundance, yields, and profits.

Sixteen California almond orchards were studied in 2018 and 2019. Replicate plots ($n = 4$; 40 × 40 m) were established in each orchard. Plots were established 20 m into the field to avoid field margin effects. The orchards in the study ranged from the Northern half of the San Joaquin Valley through the Capay Valley to Chico. Orchards were 3–38 y old. Almond orchards are generally planted with at least two different varieties, with over-lapping bloom periods to promote pollination. Therefore, all of the orchards in the study contained at least two varieties, and almond varieties varied among the orchards. In each orchard, we examined a full range of soil and water characteristics, soil microbial, plant, and invertebrate community characteristics, pest damage, yields, and economics.

Dung invertebrate communities were examined on 16 ranches in a 7,935 km² region in eastern South Dakota during 2015 and 2016. All sites were grazed by cattle for at least 5 y, but annual grazing intensity and grazing period varied. Herds ranged from 20 to 120 individuals, and the cattle differed in size, breed, and administered ivermectin products. Focal pastures were at least 4 ha in size. The systems were ranked from regenerative to conventional based on several practices (Table 2). In our study, the designation of management systems was based upon the grouping of several management practices that were occasionally all present on the same cattle operation.

A second study of cattle ranches examined plant communities and helminth fecal parasites. Selected sites ($n = 20$ per grazing treatment) focused on rangelands in the Dakotas during 2019 and 2020. Grazing treatments were assigned based on a ranch management character matrix, which consisted of rancher-defined best management practices for regenerative and conventional ranching systems (Table 2). This character matrix was adapted from a similar study conducted in the region by Pecenka & Lundgren (2019). Grazing treatments were paired within each region and year. Each grazing system had been practiced on the site for at least 4 years.

Soil quality

Soil physical and chemical properties were assessed in corn fields and in almond orchards. SOM and bulk density (BD) were assessed in both of these systems. In cornfields and almonds, four soil cores (8.5 cm deep, 5 cm in diam) were randomly collected from a single field on each of 20 farms; cores were collected at locations that were at least 10 m apart. On each sample, vegetative material was removed from 60 g of soil by hand, and the resulting samples were stored in aluminum containers at 105°C overnight. SOM in the soil sample was measured using the weight loss on ignition (LOI) technique (Davies, 1974). To measure bulk density, four soil cores of known volume per field were weighed before and after drying at 100°C for 55 h.

In cornfields, we also measured fine particulate organic matter (fPOM). Approximately 30 g of soil was weighed in sterile aluminum pans. The soil was soaked in 90 mL of hexametaphosphate for 24 h, mixed for 5 min with a stainless-steel mixer, and were then sieved through screens with 500 µm and then 53 µm holes; the finer screen isolated the fPOM fraction. The isolated sample was then weighed, dried for 24 h at 105°C. Samples were then cooled in a desiccator cabinet, weighed, baked in a furnace for 4 h at 450°C. The samples were then cooled in a desiccator cabinet and weighed a final time.

In almond orchards, soil samples were collected at random locations in the inter-row area of each plot to determine total soil carbon (TSC) and total soil nitrogen (TSN). The probe (2.54 cm × 91.44 cm) was inserted 60 cm deep. Each core was placed into a plastic bag that was stored on ice until it could be transferred to a paper bag in the laboratory. Samples were weighed, dried, and weighed again. All visible pieces of rock and organic matter were removed from the samples. The samples were ground and were then passed through

Table 1. A matrix of farm practices in cropland/orchards that can be used to distinguish regenerative and conventional farms. Shaded rows refer to corn farms in the Upper Midwest, and unshaded rows refer to almond orchards in California. Fungicide use, organic amendments, and field margins/hedgerows were not described in the cornfields. A '1' was assigned for each practice if it was considered regenerative and '0' if it was considered conventional. Eliminating synthetic fertilizers, herbicides, fungicides, insecticides, and tillage were considered regenerative. Including cover crops, organic amendments (e.g., composts, compost teas, etc), livestock integration, and diversified field margins and hedgerows were also considered regenerative practices.

Field locations (County or nearest town, State)	Fertilizers	Herbicides	Fungicides	Insecticides	Tillage	Cover crops	Field margins/hedgerows	Organic amendments	Grazers	Composite score
Merced, CA	1	1	1	1	0	1	1	1	0	7
Butte, CA	1	1	0	1	1	1	1	1	0	7
Merced, CA	1	1	1	1	1	1	1	1	0	8
Merced, CA	1	1	1	1	1	1	1	1	0	8
Yolo, CA	1	1	0	1	0	1	1	1	1	7
Butte, CA	0	0	0	0	1	1	0	1	0	3
Butte, CA	0	0	0	0	1	0	0	0	0	1
Yolo, CA	0	0	0	0	1	0	0	0	0	1
Yolo, CA	0	0	0	0	1	0	0	0	0	1
Butte, CA	1	1	0	1	1	1	1	1	0	7
Merced, CA	1	1	0	1	1	1	0	1	0	6
Merced, CA	0	0	0	0	1	0	0	0	0	1
Yolo, CA	1	1	1	1	1	1	0	0	1	7
Merced, CA	0	0	0	0	1	0	0	1	0	2
Merced, CA	0	0	0	0	1	0	0	1	0	2
Merced, CA	0	0	0	0	1	1	0	1	0	3
Bladen, NE	0	0	0	0	0	0	0	0	0	0
York, NE	0	0	0	0	0	0	0	0	0	0
Bismarck, ND	0	0	0	0	1	0	0	0	0	1
Bismarck, ND	0	0	0	0	1	0	0	0	0	1
White, SD	0	0	0	0	0	0	0	0	0	0
Pipestone, MN	0	0	0	0	0	0	0	0	0	0
Toronto, SD	0	0	0	0	0	0	0	0	0	0
Gary, SD	0	0	0	0	0	0	0	0	0	0

Field locations (County or nearest town, State)	Fertilizers	Herbicides	Fungicides	Insecticides	Tillage	Cover crops	Field margins/hedgerows	Organic amendments	Grazers	Composite score
Arlington, SD	0	0		0	0	0			0	0
Lake Norden, SD	0	0		0	0	0			0	0
Bladen, NE	1	0		1	1	1			0	4
York, NE	1	0		1	1	1			0	4
Bismarck, ND	0	1		1	1	1			1	5
Bismarck, ND	1	0		1	1	1			1	5
White, SD	1	1		1	0	1			0	4
Pipestone, MN	1	1		1	0	1			0	4
Toronto, SD	1	0		0	1	1			0	3
Gary, SD	1	1		1	0	1			1	4
Arlington, SD	1	0		1	1	1			1	5
Lake Norden, SD	1	0		1	1	1			0	4

Table 2. A matrix of farm practices in rangelands that can be used to distinguish regenerative and conventional ranches. Unshaded rows refer to ranches that were included in the dung invertebrate assessment. Shaded rows refer to ranches that were examined in the fecal parasite and plant community study.

Ranch location (Nearest town, State)	Ivermectin	Stocking density	Rotation frequency	Rest period	Composite matrix score
Bruce, SD	1	0	0	0	1
Castlewood, SD	1	2	1	2	6
Clear Lake, SD	2	2	2	2	8
Estelline, SD	1	1	1	2	5
Estelline, SD	0	1	0	0	1
Flandreau, SD	0	0	0	0	0
Gary, SD	2	2	2	2	8
Goodwin, SD	2	2	2	2	8
Madison, SD	0	1	1	2	4
Milbank, SD	1	1	1	2	5
Milbank, SD	0	0	0	2	2
Sioux Falls, SD	2	2	2	2	8
Summit, SD	1	1	1	2	5
Thomas, SD	1	1	2	2	6
Twin Brooks, SD	2	2	1	2	7
Volga, SD	0	0	0	0	0
Hayti, SD	0	0	0	0	0
Hayti, SD	0	0	0	0	0
Strandburg, SD	0	0	0	0	0
Estelline, SD	0	0	0	0	0
Watertown, SD	1	2	2	2	7
Hayti, SD	1	1	2	2	6
Estelline, SD	2	1	2	2	7
Gary, SD	2	1	2	2	7
Goodwin, SD	1	1	2	2	6
Tuttle, ND	1	0	0	0	1
Tuttle, ND	0	0	0	0	0
Wing, ND	0	0	0	0	0
Moffit, ND	1	0	0	0	1
Fort Rice, ND	2	2	2	2	8
ND7; Wing, ND	2	1	2	2	7
Bismarck, ND	2	2	2	2	8
Wing, ND	2	2	2	2	8

Ranch location (Nearest town, State)	Ivermectin	Stocking density	Rotation frequency	Rest period	Composite matrix score
Moffit, ND	2	2	2	2	8
Summit, SD	0	0	2	2	4
Summit, SD	0	0	0	0	0
Milbank, SD	2	1	1	2	6
Milbank, SD	0	0	0	0	0
Watertown, SD	2	2	2	2	8
Castlewood, SD	0	0	0	0	0
Summit, SD	2	0	2	2	6
Milbank, SD	0	0	0	0	0
Castlewood, SD	2	2	2	2	8
Castlewood, SD	0	0	1	2	2
Sheldon, ND	0	1	2	2	5
Sheldon, ND	0	0	0	0	0
Napoleon, ND	0	2	2	2	6
Napoleon, ND	0	0	0	0	0
Ellendale, ND	2	2	2	2	8
Forbes, ND	1	0	1	2	4
Forbes, ND	2	2	2	2	8
Berlin, ND	0	1	1	0	2

Management practices of cattle operations were scored 0-2 based, with higher numbers reflecting practices that promote biodiversity and soil quality. Ivermectin application frequency was divided into multiple applications (0), single application not during grazing season (1), and no ivermectin use (2). Stocking density (animal units; AU) was divided into <5 AU/ha (0), 5-10 AU/ha (1), and >10 AU/ha (2). Rotation frequency was divided into >30 d rotation (0), 10-30 d rotation (1), and <10 d rotation (2). Rest period was considered as continuously grazed during the growing season (0), a rest period of 1-30 d (1), and a rest of > 30 d (2).

a sieve with 0.180 mm openings. Elemental analysis was conducted on three subsamples (12–15 mg) that were housed on tin capsules (5 × 9 mm) (ECS 8020, NC Technologies, Milan, Italy). To control for the relative compaction and other circumstances, the mass (Mg) of TSC per ha was assessed using the Equivalent Soil Mass (ESM) method, in which a cubic spline of Mg of TSC per depth layer was calculated (Wendt & Hauser, 2013). This resulted in the assessment of carbon as Mg of TSC/ha at 6,000 Mg (59.2 cm deep).

Soil macro- and micronutrients were also quantified in the almond orchards. The samples were ground to pass a 2 mm sieve and divided into three subsamples (two were 4 g each

and one weighed 40 g). The 40 g soil sample is analyzed with a 24 h incubation test at 24°C. This sample is wetted through capillary action by adding 20 mL of deionized water to a 237 mL glass jar and then capped. After 24 h, the gas inside the jar was analyzed using an infrared gas analyzer (IRGA) (Li-Cor 840A, LI-COR Biosciences, Lincoln NE) for CO₂-C. The two 4 g samples were extracted with 40 mL of deionized water and 40 mL of H³A to extract the NO₃-N, NH₄-N, and PO₄-P from the samples. The samples were shaken for 10 min, centrifuged for 5 min, and filtered through filter paper (Whatman 2V, Cytiva, Marlborough, MA). The water and H³A extracts were analyzed on a flow injection analyzer (Lachat 8000, Hach Company, Loveland CO). The water extract was also analyzed on a

Teledyne-Tekmar Torch C:N analyzer for water-extractable organic C and total N. The H³A extract was also analyzed on a Thermo Scientific ICP-OES instrument for P, K, Mg, Ca, Na, Zn, Fe, Mn, Cu, S and Al. The Haney soil health score combines five independent measurements of a soil's biological properties to provide a general estimate of the overall health of a soil system (Haney *et al.*, 2018). It is calculated as 1-d-CO₂-Carbon/10 plus water extractable organic carbon (WEOC)/50 plus water extractable organic nitrogen (WEON)/10.

Water infiltration

We used the rainfall infiltration rate kits of the Natural Resource Conservation Service (NRCS) to sample water infiltration rates in northern cornfields and California almonds. A metal ring (15 cm diam, 13.5 cm tall) was hammered 6.5 cm into the soil. Water (444 mL) was poured into the ring, and the time it required to saturate into the soil was recorded. A second container of 444 mL of water was poured into the ring, and the time to saturation was recorded again. In cornfields, this process was recorded during anthesis.

Microbial diversity

Microbial communities were only sampled in almond orchards using the Phospholipid fatty acid (PLFA) test. Soil cores (15 cm depth, 1.9 cm diam; N = 16), were taken from four replicates per site per farm during the fruiting period. The samples were taken at random locations within each replicate, at least 5 m apart, using a transect that diagonally bisected the plot. Soil cores for each orchard were combined in a sealed plastic bag and placed in a cooler with dry ice (Drenovsky *et al.*, 2010). Soil samples were stored at -80°C until they could be freeze-dried and ground to 2 mm particle sizes. Phospholipid fatty acid (PLFA) testing provided an index of a soil's microbial biomass and composition (Frostegård *et al.*, 2011). The microbial biomass and community composition were recorded as total microbial biomass, undifferentiated microbial biomass, total bacteria, Gram-positive bacteria, Actinomycetes, Gram-negative bacteria, *Rhizobia* bacteria, total fungi, arbuscular mycorrhizal fungi, saprophytic fungi, and Protozoa.

Plant community structure

Plant community structure was assessed solely as ground cover in the almond study. The ground cover height and composition in each of the replicates/plots was recorded during each of the three sampling periods. The percent ground cover was categorized as 0–25%, 25–50%, 50–75%, and 75–100%. Percent ground cover in the overall orchard was assessed using visual assessments of the percent ground cover in each invertebrate quadrat.

Plant community diversity and green canopy cover were examined during the 2019 and 2020 grazing seasons of the rangeland study, while vegetation biomass was assessed only during the 2020 season. All three plant community metrics were measured during three periods of the grazing season; early June, mid-July, and late-August/early-September. At each pasture site, two 50 m transect lines were established 15 m

apart and perpendicular to the slope of the land. Plant community diversity was assessed using the belt transect method for monitoring native prairie vegetation in the Dakotas (Grant *et al.*, 2004). Briefly, plant community structure was recorded for a randomly selected 5 × 0.5 m² belt along each transect line. Vegetation is classified in each 5 m segment according to a hierarchical breakdown of plant groups common to the region (Grant *et al.*, 2004). Green canopy cover was documented using the smartphone app Canopeo, which quantifies fractal green vegetation canopy cover area per unit area (Patrignani & Oschner, 2015). Quadrats (50 × 25 cm²) were established at 0, 25, and 50 m along transect lines to quantify green canopy cover with Canopeo. Lastly, a disc pasture meter was used to estimate above-ground vegetation biomass along each transect line and within their immediate surrounding area (<15 m). Compression height of vegetation was recorded by dropping a 0.13 m² plate weighing 1.34 kg from a height of 183 cm.

Insect diversity

Insect surveys in cornfields and almonds involved sampling the epigeal community from 0.25 m² sheet-metal quadrats (15 cm tall) inserted into the soil (Lundgren *et al.*, 2006). Quadrats were placed in randomly selected locations between corn rows (n = 5 per plot) during anthesis. Invertebrates were exhaustively collected from the surface of the soil and beneath plant debris using handheld mouth aspirators. In almonds, sampling of the invertebrate communities occurred during the bloom, fruit development, and harvest periods. The invertebrate communities that could be collected from the soil surface and top 2 cm of the soil with mouth-operated aspirators in 15 min were preserved in 70% ethanol.

In cornfields, we also sampled the foliar invertebrate community. The foliar invertebrate community was sampled using a destructive whole plant assessment. Corn plants (n = 25 per plot) were randomly selected, thoroughly examined, and invertebrates that were not sight-identified were collected using mouth aspirators. Plants were then severed at the ground level and were transported to the field margin where their leaves, stems, ears, whorls, and tassels were inspected and dissected on white sheets.

Soil invertebrate and dung invertebrate communities were assessed using soil cores in corn fields and rangelands. In corn fields, five soil cores (10 cm diameter, 10 cm height) were collected per field in randomly selected locations within and between corn rows during anthesis. In rangelands, the same core approach was used to collect invertebrates from 2–5 d old dung pats and the soil directly beneath the pat (n = 10 per ranch). Samples were collected monthly from May to September. All cores were refrigerated for < 36 h, and then were placed into a Berlese collection system over 7 d, when they reached a constant weight.

All extracted invertebrates were identified microscopically and cataloged; each unique specimen was identified to the lowest taxonomic level possible representing a functional morphospecies. Each morphospecies was placed into a trophic guild

(coprophage, predator, parasitoid, herbivore, or pest) based upon previous descriptions of the biology of arthropod community in these systems. Voucher specimens are housed in the Mark F. Longfellow Biological Collection at Blue Dasher Farm, Estelline, SD, USA.

Pests

In cornfields, pests included aphids, lepidopteran larvae, and corn rootworm adults that were identified from the foliar bioinventories. In almonds, pest incidence was assessed on 500 almonds per farm in 2018 and 600 almonds per farm in 2019 (< 20 from any one tree) (Bentley *et al.*, 2001; Doll, 2009; Legner & Gordh, 1992). The almonds were each categorized as having no pest damage, navel orange worm damage (*Amyelois transitella* [Walker]; Lepidoptera: Pyralidae), ant damage (Formicidae), oriental fruit moth damage (*Grapholita molesta* [Busck]; Lepidoptera: Tortricidae), peach twig borer damage (*Anarsia lineatella* Zeller; Lepidoptera: Gelechiidae), leaf footed plant bug or stinkbug damage (Hemiptera: Coreidae, Pentatomidae), and unknown pest damage (Bentley *et al.*, 2001; Doll, 2009; Symmes, 2018).

Fecal oocyst counts of coccidia and egg counts of gastrointestinal nematodes belonging to the superfamilies Trichostrongyloidea and Strongyloidea were conducted on herds from the 2019 – 2020 rangeland study. Three grams of dung was collected from 2 – 4 d old cattle dung pats present in pastures ($n = 5$ pats sampled/site). Fecal samples were processed according to the Modified Wisconsin Sugar Float Technique (Cox & Todd, 1962). Coccidia oocysts and eggs of Trichostrongyles and Strongyles were quantified for 1 g of fecal sample. Sampling for internal parasites in the herds was conducted twice in 2019 (mid-July and late-August/early September) and three times in 2020 (early-June, mid-July, and late-August/early September).

Yield and profit

In cornfields and almonds, responses from producer surveys were used to determine management practices, costs, and revenues that went into the direct net profitability of each operation. Gross profit analysis included yield, return on grain, and additional revenue that included livestock grazing and production. In cornfields, yield information provided by the farmers were confirmed by yields that were hand-gathered from three 3.5-m row sections from each replicate-field. Costs associated with corn production included corn seed, cover crop seed, drying/cleaning grain, crop insurance, tillage, planting corn, planting cover crop, fertilizers, herbicides, and irrigation (additional information on profit/loss analysis can be found in LaCanne & Lundgren 2018). Operating costs in almonds included winter sanitation, sampling for tree nutrient status and soil salinity, pH, and nutrient levels, irrigation and frost protection, fertilizers, insecticides, herbicides, fungicides, disease treatment sprays, trapping vertebrate pests, cover crop seed, tillage, mowing, flammers, grazers, and harvest.). Harvest in the almonds included the hourly labor to conduct the harvest and the price paid to external contractors (additional information on the almond profit/loss analysis can be found in Fenster *et al.* in prep). In both systems, only direct costs and revenues were used to calculate profitability.

Data analysis

Unless otherwise described, we used linear regressions to search for correlations between the regenerative matrix scores and response variables. In the almond study, clay percentages of the soils were considered as co-factors in all models that examined TSC. Plot values were composited across plots into single values in the almond study and linear regression analyses were used to compare regenerative scores and response variables. In instances where resampling a single farm was performed (either spatially or temporally) Linear Mixed Models were used to remove pseudoreplication and account for this dependence; field and farm were included as random factors in the corn models, and month was included as a fixed factor and farm as a random factor in the rangeland studies. All statistics were conducted using Systat 13.1 (Systat Software Inc., San Jose, CA)

Results

Soil physical and chemical properties

In cornfields, fPOM of the soil was strongly and positively associated with regenerative matrix scores ($F_{1,17} = 4.61$; $P = 0.047$) (Figure 1A), but SOM ($F_{1,17} = 0.11$; $P = 0.75$), bulk density ($F_{1,17} = 2.32$; $P = 0.15$), and water infiltration rates ($F_{1,11} = 0.47$; $P = 0.51$) of the soil were not correlated with matrix scores. The slowest water infiltration rates in the higher regenerative scoring farms were those that practiced tillage.

In almond orchards, higher matrix scores correlated to significantly higher levels of TSC and TSN (500–6000 ESM layers) and SOM (15 cm depth). At the 0–6000 Mg ESM layer, there was a significant correlation between the orchards' matrix scores and TSC (model: $F_{2,13} = 13.77$, $P = 0.001$; score: $t = 2.26$, $P = 0.04$; clay: $t = 4.73$, $P < 0.001$) (Figure 2). At the 6000 Mg ESM layer there was a significant correlation between the orchards' matrix scores and TSN (model: $F_{2,13} = 17.73$, $P < 0.001$; score: $t = 1.94$, $P = 0.08$; clay: $t = 5.63$, $P < 0.001$). In contrast to what we found in cornfields, in the top 15 cm of soil there was a significant correlation between the orchards' SOM%, matrix scores, and clay percentage (model: $F_{2,13} = 47.56$, $P < 0.001$; score: $t = 5.50$, $P < 0.001$; clay: $t = 8.05$, $P < 0.001$) (Figure 1B).

In almonds, higher matrix scores and soil clay percentages correlated to higher levels of WEON ($F_{2,13} = 9.16$, $P = 0.003$) and WEOC ($F_{2,13} = 15.61$, $P = 0.001$) in the organic matter of the soils. Total phosphorus (model: $F_{2,13} = 4.32$, $P = 0.04$; score: $t = 2.65$, $P = 0.02$, clay: $t = -1.28$, $P = 0.23$) and inorganic phosphorus (model: $F_{2,13} = 4.56$, $P = 0.03$, score: $t = 2.74$, $P = 0.02$, clay: $t = -1.28$, $P = 0.22$) were significantly and positively correlated with an orchard's matrix score. Higher matrix scores correlated to higher levels of Calcium ($F_{2,13} = 5.97$, $P = 0.01$) and Sulfur ($F_{1,14} = 10.80$, $P = 0.01$), but lower levels of Aluminum ($F_{1,14} = 6.13$, $P = 0.03$). There were no correlations between other micronutrients and the regenerative matrix score ($P > 0.05$). Haney soil health test scores were positively correlated with regenerative matrix scores (model: $F_{2,13} = 7.82$, $P = 0.01$, Score $t = 2.99$, $P = 0.01$, clay: $t = 2.59$, $P = 0.02$) (Figure 4). Soil respiration was unaffected by regenerative matrix scores (model: $F_{2,13} = 2.91$, $P = 0.09$, Score $t = 1.62$, $P = 0.13$, clay: $t = 1.77$, $P = 0.10$).

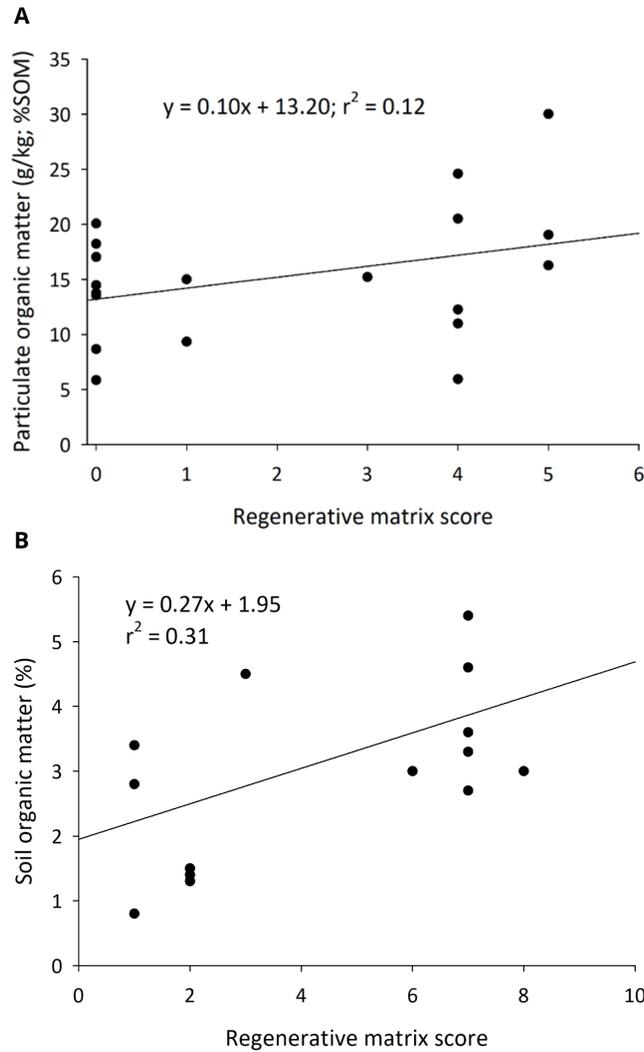


Figure 1. The relationships of regenerative farming intensity on fine particulate organic matter in cornfields of the Upper Midwest (A) and percent soil organic matter in California almond orchards (B).

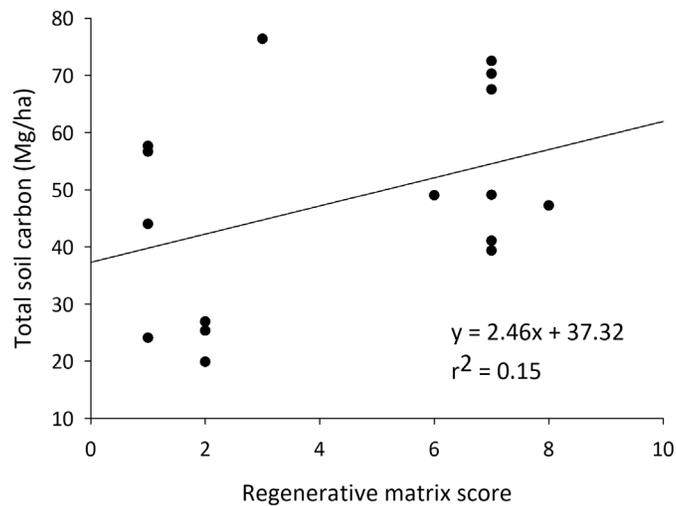


Figure 2. Total soil carbon in California almond orchards as the orchards increase the number of regenerative practices.

Higher regenerative-conventional matrix scores in almonds correlated to lower bulk densities (model $F_{2,13} = 8.95$, $P = 0.004$; score: $t = -3.91$, $P = 0.002$, clay: $t = -1.61$, $P = 0.13$). Water infiltration rates increased alongside matrix scores for these orchards (model: $F_{2,5} = 11.43$, $P = 0.01$; score: $t = -4.50$; $P = 0.009$, clay: $t = 2.56$, $P = 0.05$) (Figure 3).

Soil microbiology

In almond orchards, higher matrix scores correlated to higher amounts of bacterial biomass ($F_{1,14} = 6.25$, $P = 0.03$) and higher Gram (+) biomass ($F_{1,14} = 9.12$, $P = 0.01$). The percent composition in the microbial community was not affected by regenerative score ($F_{1,14} = 2.47$, $P = 0.14$), indicating greater overall microbial biomass rather than bacterial dominance. The remainder of these microbial community metrics were not correlated to the regenerative conventional matrix score ($P > 0.05$).

Plant community

More regenerative cropland and rangeland supported more diverse and robust plant communities. In almond orchards, higher matrix scores correlated to more ground cover ($F_{1,14} = 127$, $P < 0.001$), more plant species ($F_{1,14} = 27.61$, $P < 0.001$) (Figure 5A), and greater biomass height ($F_{1,14} = 9.90$, $P = 0.01$). In rangelands of the Northern Plains, there was a significant relationship between matrix score and plant biomass (Score: $F_{1,53} = 8.47$, $P = 0.01$; month: $F_{2,53} = 0.45$, $P = 0.64$) (Figure 5B), plant diversity (Score: $F_{1,106} = 14.22$, $P < 0.001$; month: $F_{3,106} = 1.02$, $P = 0.39$) (Figure 5C), and ground cover (Score: $F_{1,106} = 10.06$, $P = 0.002$; month: $F_{3,106} = 14.76$, $P < 0.001$).

Invertebrate community

Across cropland and rangeland, more regenerative farms had more robust invertebrate communities. Several aspects of invertebrate community structure in cornfields were strongly and positively correlated with how regenerative a field was scored. The abundance ($F_{1,55} = 6.53$, $P = 0.01$), species richness

($F_{1,55} = 38.31$, $P < 0.001$), and diversity ($F_{1,55} = 7.34$, $P = 0.01$) (Figure 6A) of the epigeal invertebrate community was positively affected by matrix score. Similarly, matrix scores were positively associated with the species richness of the invertebrate community found in the soil column ($F_{1,56} = 12.96$, $P = 0.001$), but the abundance was only marginally associated with matrix score ($F_{1,56} = 3.41$, $P = 0.07$), and the diversity of this community was unaffected by matrix score ($F_{1,56} = 2.45$, $P = 0.12$). Foliar invertebrate abundance ($F_{1,55} = 0.99$, $P = 0.32$), species richness ($F_{1,55} = 1.35$, $P = 0.25$), and species diversity ($F_{1,55} = 0.20$, $P = 0.66$) were not related to the matrix scores.

The invertebrate community on the soil surface was positively affected by the number of regenerative practices on almond orchards. Invertebrate abundance ($F_{1,14} = 10.28$, $P = 0.01$), biomass ($F_{1,14} = 70.10$, $P < 0.001$), species richness ($F_{1,14} = 14.44$, $P = 0.002$) and species diversity indices (H: $F_{1,14} = 12.37$, $P = 0.003$; DS: $F_{1,14} = 11.11$, $P = 0.01$) (Figure 6B) were positively correlated with matrix score. There was a positive correlation between earthworm (square rooted to normalize residuals) abundance and matrix score on these orchards ($F_{1,14} = 9.87$, $P = 0.007$).

In rangelands of the Northern Plains, the invertebrate community associated with cattle dung was strongly and positively associated with the matrix score attained. The species richness (Score $F_{1,74} = 15.38$, $P < 0.001$; month $F_{4,74} = 6.54$, $P < 0.001$), and species diversity (Score $F_{1,74} = 16.10$, $P < 0.001$; Date: $F_{4,74} = 1.50$, $P = 0.21$) (Figure 7A) of the dung invertebrate community were positively associated with matrix score. Abundance of dung invertebrates was uncorrelated with matrix score (Score $F_{1,74} = 0.01$, $P = 0.99$; month: $F_{4,74} = 4.83$, $P = 0.001$). The abundances of two critical functional groups, dung beetles (Score $F_{1,60} = 10.05$, $P = 0.006$; month: $F_{4,60} = 5.70$, $P = 0.001$) (Figure 7B) and invertebrate predators, (Score $F_{1,74} = 7.41$, $P = 0.01$; month: $F_{4,74} = 4.09$; $P = 0.01$) increased as a rangeland became more regenerative.

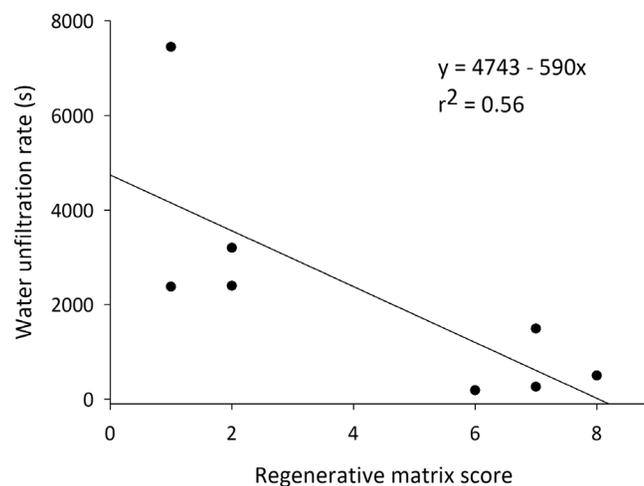


Figure 3. Water infiltration rates in the soils of California almond orchards in relation to the number of regenerative practices on each orchard.

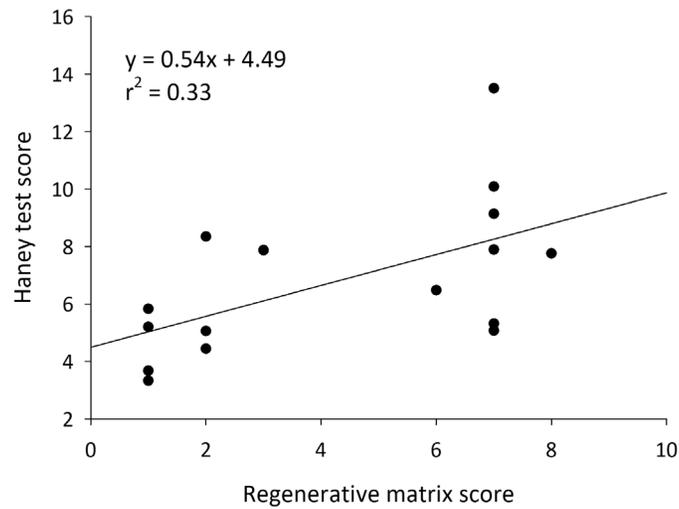


Figure 4. Haney test scores on soils from California almond orchards with increasing numbers of regenerative practices.

Pest populations

In all of these systems, pest populations were uniformly below any recognized economic thresholds. Pests were not correlated with the number of regenerative practices used in cornfields (pest abundance: $F_{1,55} = 2.18$, $P = 0.15$) or in almond orchards (% damaged nuts: $F_{1,14} = 0.69$; $P = 0.42$). In rangelands, there was no relationship between the matrix score and *Trichostrongyles* (Score: $F_{1,88} = 0.09$, $P = 0.77$; month: $F_{3,88} = 0.39$, $P = 0.76$) or *Coccidia* (Score: $F_{1,88} = 1.78$, $P = 0.19$; month: $F_{3,88} = 0.70$, $P = 0.56$) fecal parasite loads.

Yields

Corn yields were negatively correlated with the regenerative matrix score ($F_{1,56} = 3.88$, $P = 0.05$). Despite this, three of the top ten yielding cornfields were regenerative, indicating that regenerative practices are not necessarily tied with yield reductions. Standard deviations became greater proportions of the mean as a farm became more regenerative (variability in yields were higher among regenerative farms) ($R^2 = 0.79$; $F_{1,4} = 5.12$, $P = 0.11$). In almonds there was no relationship between almond yield and how regenerative an orchard was ($F_{1,12} = 0.86$, $P = 0.37$). On a treatment level, regenerative farms had twice the variance in yield relative to the conventional orchards.

Economics

Although there was a positive relationship between matrix score and profitability in cornfields, this relationship was not significant ($F_{1,53} = 2.49$, $P = 0.12$). In cornfields, there was a significant increase in the standard deviation of profit as a farm became more regenerative (e.g., standard deviation as a proportion of the mean; $R^2 = 0.72$; $F_{1,4} = 7.68$, $P = 0.07$). This indicates more variability in profits among the more regenerative producers. The top ten highest netting farms were all regenerative farms (scores of four and five). In almonds, profit

was strongly and positively correlated with matrix scores ($F_{1,11} = 7.41$, $P = 0.02$) (Figure 8). Operating costs were unaffected by matrix score ($F_{1,13} = 0.70$, $P = 0.42$), but net income increased as regenerative scores increased ($F_{1,11} = 8.50$, $P = 0.01$).

Discussion

Our approach to defining farms based on a small suite of carefully chosen practices was strongly associated with several key metrics that define regenerative systems, including soil health, biodiversity promotion and profitability. With our matrix, we have distilled complex agricultural systems down to fewer than 10 practices that serve as useful indicators of regenerative operations in cropland and rangeland. These practices represent each of the five principles that define regenerative agriculture in cropland and in rangeland. We have determined that these practices are regenerative across cropping systems; the cropland matrix works in row crop systems as well as a perennial system including orchards. Also, it was noteworthy that none of the regenerative farm attributes attained an asymptote, which suggests that farms could become more regenerative if they continue to incorporate additional regenerative practices. This matrix can be used to categorize a farm as regenerative or conventional based on a threshold score.

Regenerative farming practices inherently increase plant biomass and diversity in both cropland and rangeland systems, and this plant community is a central mechanism for improving the soil health, biodiversity, and resilience of these farming operations. Farmers in this study enhanced their plant diversity and biomass (Figure 5) using a variety of tools. In almonds and cornfields, many of the farmers planted annual cover crops to improve the health of their soils. In some regenerative orchards, farmers allowed the native vegetation to persist. All of the regenerative orchards stopped the use of

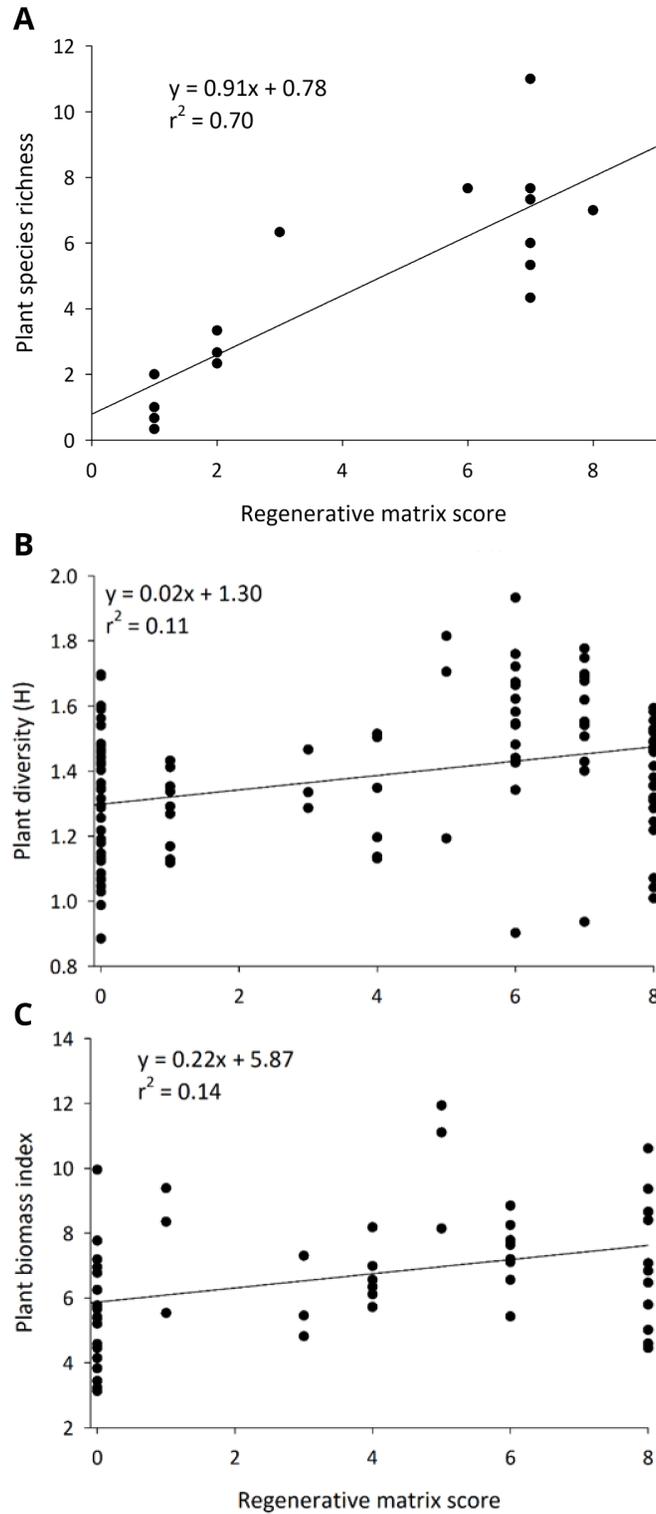


Figure 5. Plant species richness on the orchard floor in California almonds (**A**), and plant diversity (Shannon H; **B**) and biomass index (**C**) in rangelands of the Northern Plains as they relate to how regenerative a farm is.

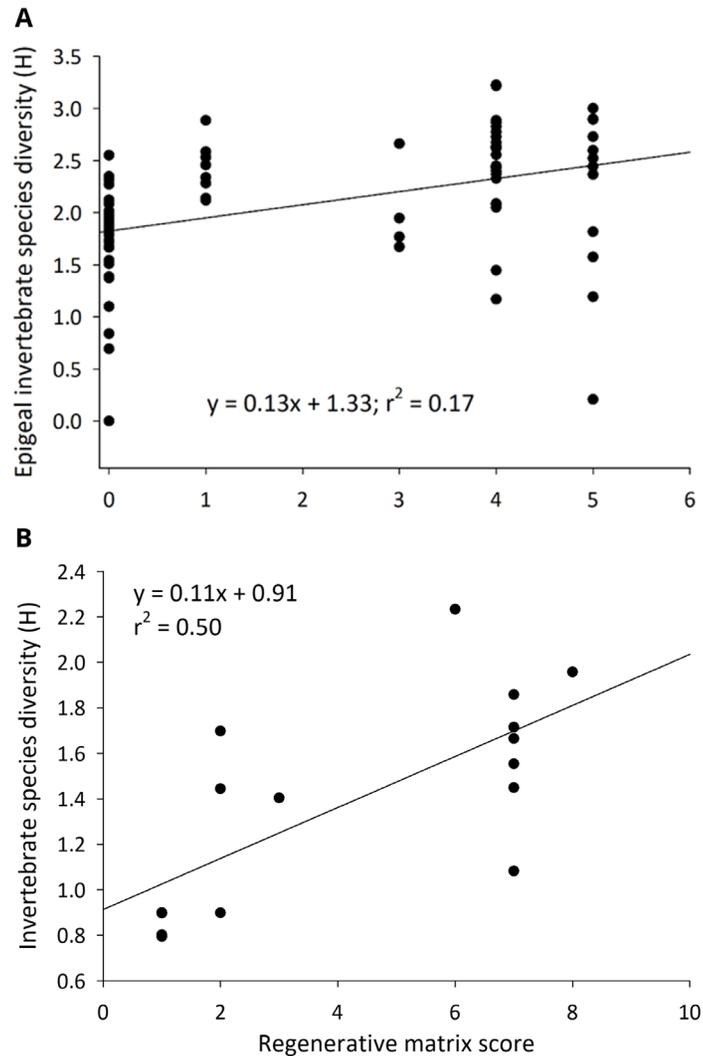


Figure 6. The invertebrate species diversity (Shannon H) on the soil surface in cornfields of the Upper Midwest (**A**), and on the orchard floors of California almonds (**B**), as they relate to the number of regenerative practices implemented on farms.

synthetic herbicides, and this likely contributed to greater plant diversity. In rangelands, short-term, intense grazing stimulates plant communities to diversify and grow if the plant community is allowed to rest following a grazing event (Hillenbrand *et al.*, 2019; Teague *et al.*, 2011). Plants are a vital part of the carbon cycle, and as such they are the primary means whereby energy enters into an ecosystem (Weil & Brady, 2017). Additionally, plant communities are requisite directly and indirectly to the genesis of healthy soils (Lucas, 2001). Plant roots provide the needed polysaccharides for microbial communities to grow and perform key nutrient cycling services (Philippot *et al.*, 2013). Physical properties of soils are also influenced by plants, including water infiltration rates, soil aggregate structure, bulk density, etc. (Gulick *et al.*, 1994; Liu *et al.*, 2005). Plant diversity and biomass scales positively

with the diversity of nearly every other group of organisms (Bianchi *et al.*, 2006; Saunders *et al.*, 2013; Zak *et al.*, 2003), and thus plant communities are a driver of biodiversity within agricultural habitats. Taken in sum, the relationships between plant communities and regenerative score illustrate an important mechanism for how regenerative farms positively enhance many intended regenerative outcomes.

Animal integration is another important tool for managing farms that improve regenerative outcomes. The most regenerative cropping operations observed in this set of studies always integrated livestock (chickens, sheep, or cattle), and these farms also had the greatest biodiversity, soil health, water infiltration rates, and economic metrics. Regeneratively-managed livestock in cropland improve soil chemical and physical

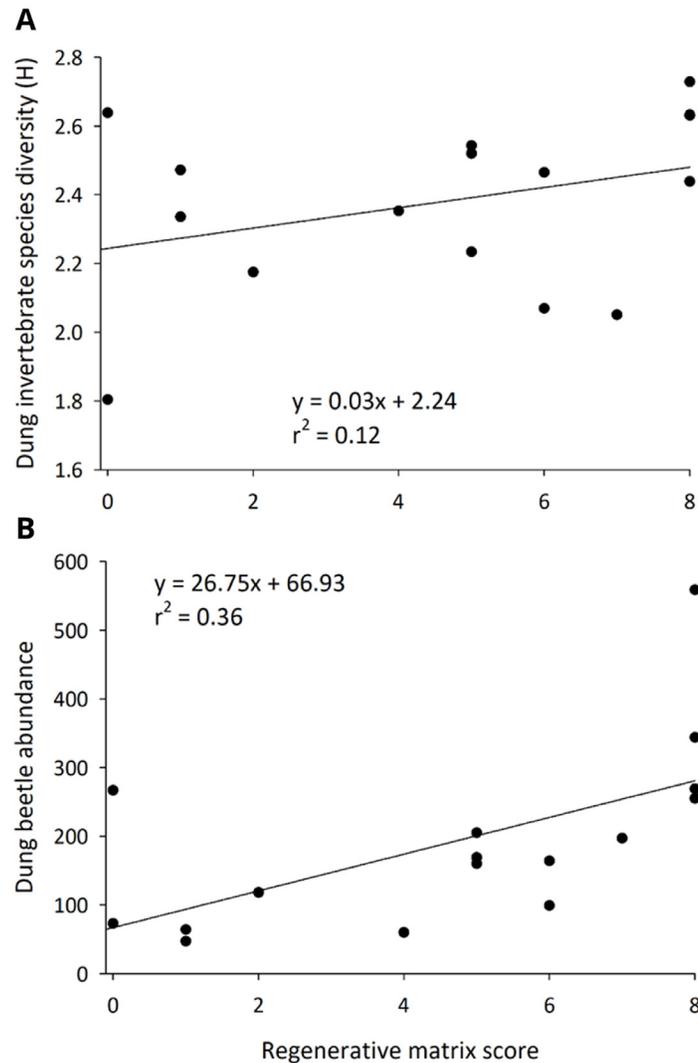


Figure 7. Invertebrate species diversity (Shannon H) in dung pats (A) and the abundance of dung beetles (B) as they relate to regenerative intensity on ranches in the Northern Plains.

properties, increase niche diversity which allows other organisms to thrive, and provides additional revenue and an additional plant management tool that can increase farm resilience (Colley *et al.*, 2019; Delgado *et al.*, 2011; Niles *et al.*, 2018; Teague *et al.*, 2016). Important logistical constraints influence the circumstances of integrating livestock into cropland, especially in standing crops like almonds. Barriers preventing farmers from integrating livestock into cropland should be removed so this important management tool is more commonly adopted.

Soil chemical and physical properties were positively affected by the number of regenerative practices in cropland. Bulk density was significantly lower in the more regenerative almond orchards. Soil carbon and soil organic matter were strongly associated with more regenerative farms. Although

we did not conduct a carbon life cycle assessment on corn or almonds, these results support the idea that regenerative farming systems can help our cropland sequester more carbon and help to offset greenhouse gas emissions (Lal, 2020; Soto *et al.*, 2021). Organic and inorganic phosphorous are also enhanced as almond orchards become more regenerative. Phosphorous, a mined agricultural nutrient, is becoming increasingly rare and is available at significant economic and environmental costs (Alewell *et al.*, 2020), so enhancing the plant-available forms of phosphorous that are present in the soils is an important outcome of regenerative agriculture. Micronutrients calcium and sulfur were also enhanced by increasing the number of regenerative practices in almonds. In almonds, water infiltration rates were significantly improved by regenerative production practices. Water is becoming more scarce (Burri *et al.*, 2019), especially in California (Mall & Herman, 2019), as climates

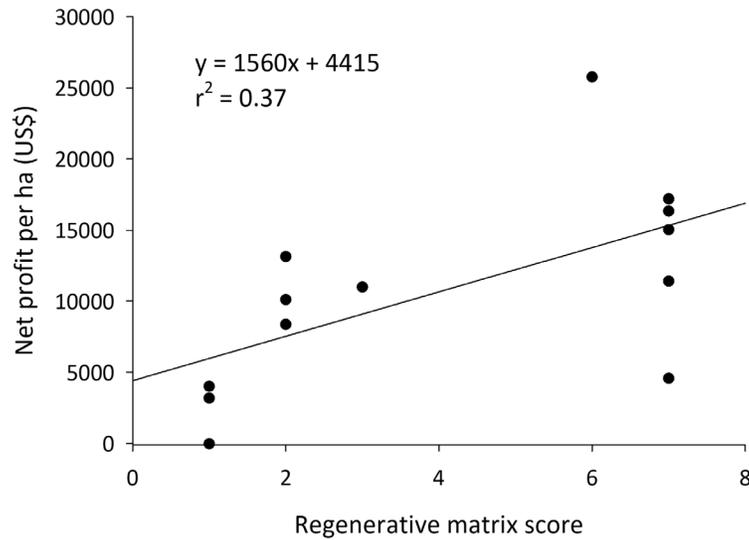


Figure 8. Net profit of California almonds as it relates to the number of regenerative practices on orchards.

continue to change and ground and surface waters are exhausted from conventional agricultural practices. Absorbing water into the soil when it becomes available is a crucial step in keeping these agricultural areas productive. Similar to previous studies (Pikul & Aase, 2003; Pikul *et al.*, 2009), tilled cornfields had lower water infiltration rates and lower bulk density. This data supports the argument that no-till practices are an indispensable core component of regenerative agriculture, and argues in favor of mandating that regenerative operations be no-till.

All organismal groups measured were enhanced as farms became more regenerative. There was greater bacterial biomass and Gram+ bacteria from the farms with higher regenerative scores. In general, bacterial dominated soils are undesirable, as they tend to drive faster decomposition and reduce soil organic matter (Hendrix *et al.*, 1986; Lehman *et al.*, 2015). Bacteria dominated soils are produced through tillage, and by definition, regenerative farms are not tilled. Only two of the almond orchards in this study had been tilled; and this must be considered when comparing bacterial communities in regenerative versus conventional almond systems. Despite increasing bacterial biomass in the regenerative orchard soils, the proportion of bacteria to other microbial life did not increase. Furthermore our soil health metric, the Haney Test that accounts for microbial function in a soil (Haney *et al.*, 2018), was positively associated with the regenerative score of a farm. This suggests that although soil bacterial biomass may be increasing on regenerative almond orchards, it is not at the expense of other microbial taxa and it enhances bacterial function. Invertebrate biomass, diversity, and abundance in the soil, on the soil surface, and in the vegetation were positively affected by regenerative practices in all three study systems. This was particularly true for the epigeal communities. We hypothesize that promotion of the invertebrate community on the soil surface is a product of

enhancing the plant community in this same stratum. Herbicides (Bohnenblust *et al.*, 2016; Morton *et al.*, 1972), fungicides (Johnson *et al.*, 2013), insecticides (Bredeson & Lundgren, 2018; Pecenka & Lundgren, 2019; Seagraves & Lundgren, 2012), and synthetic fertilizers (Cuesta *et al.*, 2008; Kromp, 1999) disrupt invertebrate communities. Regenerative farms that abandon these chemicals may experience enhanced invertebrate communities and biological diversity, which is important for farmers because it provides valuable services, including pest suppression.

Pest populations of rangelands and croplands were below economically damaging levels, but these populations were suppressed using very different means in regenerative and conventional systems. In conventional systems, pests are kept low through genetically modified crops and insecticide applications. Conventional almond orchards in this study experienced up to five insecticide applications annually. All of the conventional corn farms are planted with genetically modified, insect resistant (i.e., Bt) varieties that were treated with neonicotinoids. All of the most conventional ranches applied ivermectins in their animals to reduce fecal parasite loads. Our research shows that regenerative practices produce the same low pest populations as when cattle were treated with ivermectins, but with lower input costs. Other work in agricultural ecosystems has shown that invertebrate diversity is essential to mitigating pest populations (Crowder *et al.*, 2010; Letourneau *et al.*, 2009; Lundgren & Fausti, 2015; Lundgren & Fergen, 2014). This diversity within agricultural systems results in biological control performed by invertebrates and pathogens keeping pest populations at sub-economically damaging levels. We will test whether this same pattern was true in dung pats. Predation isn't the only mechanism at work, and we also hypothesize that host plants and livestock are healthier

in regenerative systems, which contributes to fewer pests (Barbosa *et al.*, 2009). Also, in almond orchards the more robust epigeal invertebrate community or livestock may be playing a key role in breaking down fallen mummy nuts in which navel orange worm larvae overwinter. There also are likely synergisms among organisms in regenerative systems that balance communities and resist outbreaks of specific pest populations. But in the end, the complexity of biological network interactions in complex ecosystems prevent us from understanding all of the mechanisms whereby regenerative systems consistently suppress pests without pesticide inputs.

Improved soil health and biodiversity on the studied farms was associated with increased profitability. In almonds, there was a clear relationship between more regenerative practices and the profitability of the orchards. In cornfields, the most regenerative farms were also the most profitable, but there was substantially more variance in this relationship. These higher net profits were the result of lowering seed costs, reducing chemical premiums, and value-added marketing of the most regenerative products. Yields did not follow the same pattern as the profitability with regards to matrix scores. Almond yield was not correlated with how regenerative a farm was, and corn yields were significantly reduced as regenerative practices were added to cornfields. It is important to interpret these results recognizing that regenerative agriculture is still in its infancy in many production systems. Three of the top 10 yielding corn fields were regenerative, meaning regenerative corn production methods do not necessarily lead to yield reductions. Given the low reliance on corn as a human food source, and the lack of yield reductions in almonds, concerns over regenerative food systems' ability to feed a growing human population are not supported by our research. All of this is to say that regenerative farming practices are correlated with increased profitability of a farm, thus supporting the notion that we can improve farm resilience while promoting the natural resource base of a habitat (Schipanski *et al.*, 2016).

In looking at the data on these established farm systems, there were consistently two distinct clusters of farms based on regenerative score that could be used to categorize regenerative and conventional farming operations. The natural groupings could be divided at scores of 5 and 3 for cropland and rangeland, respectively (in cornfields, this dividing line was around 3, since we did not ask all of the questions to corn farmers in that study). One explanation for this may be that farms have trouble staying in business when they only employ an intermediate number of regenerative practices. Farms in the “regenerative” and “conventional” categories are not all created equal, since changing the number of regenerative

or conventional practices can produce a variance in farm performance within each category. Also, the range of scores in the regenerative ranches were greater than that for in cropland, and could indicate a transition in the adoption of regenerative practices by ranchers of the Northern Plains. The duration that a farm is in its respective system also will likely affect the observed regenerative outcomes, and this temporal factor should be considered in further interpretations of regenerative scores. Uses for this matrix might include regulation based on carbon sequestration, water use efficiency, or pollution. Certification of regenerative operations and marketing of regenerative labeling might also employ the matrix approach as we have laid out. In the *Extended data*, we provide a survey for producers to generate their regenerative scores. We hope that this work will be built upon and tested as we confront the challenge of how to empirically define regenerative farming systems.

Data availability

Underlying data

Open Science Framework: Matrix paper, <https://doi.org/10.17605/OSF.IO/G697Y> (Lundgren, 2021) (registered 15th January 2021 <https://osf.io/7v4z2>).

This project contains the following underlying data:

- Corn
- Regenerative Rangeland 1
- Almond
- Regenerative Rangeland 2

Extended data

Open Science Framework: Matrix paper, <https://doi.org/10.17605/OSF.IO/G697Y> (Lundgren, 2021) (registered 15th January 2021 <https://osf.io/7v4z2>).

This project contains the following extended data:

- CROPLAND Regenerative Score Calculator
- RANGELAND Regenerative Score Calculator

Data are available under the terms of the [Creative Commons Zero “No rights reserved” data waiver](#) (CC0 1.0 Public domain dedication).

Acknowledgements

We thank the dozens of technicians who helped us to collect and enter the data from these projects. Mark Longfellow assisted in gathering survey information for the character matrix.

References

AGW: Certified Regenerative by A Greener World. Web-based document. 2020.
 Reference Source
 Alewell C, Ringeval B, Ballabio C, *et al.*: Global phosphorus shortage will be

aggravated by soil erosion. *Nat Commun.* 2020; 11(1): 4546.
 PubMed Abstract | Publisher Full Text | Free Full Text

Alfaro-Arguello R, Diemont SAW, Ferguson BG, *et al.*: Steps toward sustainable ranching: An emergy evaluation of conventional and holistic management

- in Chiapas, Mexico. *Agricultural Systems*. 2010; **103**(9): 639–646.
[Publisher Full Text](#)
- Barbosa P, Hines J, Kaplan I, et al.: **Associational Resistance and Associational Susceptibility: Having Right or Wrong Neighbors**. *Annu Rev Ecol Evol Syst*. 2009; **40**: 1–20.
[Publisher Full Text](#)
- Bentley WJ, Hendricks LC, Duncan RA, et al.: **BIOS and conventional almond orchard management compared**. *California Agriculture*. 2001; **55**(5): 12–19.
[Publisher Full Text](#)
- Bianchi FJA, Booij CJH, Tschamntke T: **Sustainable pest regulation in agricultural landscapes: a review on landscape composition, biodiversity and natural pest control**. *Proc Biol Sci*. 2006; **273**(1595): 1715–1727.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Bohnenblust EW, Vaudo AD, Egan JF, et al.: **Effects of the herbicide dicamba on nontarget plants and pollinator visitation**. *Environ Toxicol Chem*. 2016; **35**(1): 144–151.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Bredeson MM, Lundgren JG: **Thiamethoxam seed treatments reduce foliar predator and pollinator populations in sunflowers (*Helianthus annuus*), and extra-floral nectaries as a route of exposure for seed treatments to affect the predator, *Coleomegilla maculata***. (Coleoptera: Coccinellidae). *Crop Protection*. 2018; **106**: 86–92.
[Publisher Full Text](#)
- Burri NM, Weatherl R, Moeck C, et al.: **A review of threats to groundwater quality in the anthropocene**. *Sci Total Environ*. 2019; **684**: 136–154.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Carbon_Underground: **What is regenerative agriculture?** Web-based document. 2020.
[Reference Source](#)
- Colley TA, Olsen SI, Birkved M, et al.: **Delta life cycle assessment of regenerative agriculture in a sheep farming system**. *Integr Environ Assess Manag*. 2019; **16**(2): 282–290.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Cox DD, Todd AC: **Survey of gastrointestinal parasitism in Wisconsin dairy cattle**. *J Am Vet Med Assoc*. 1962; **141**: 706–709.
[PubMed Abstract](#)
- Crowder DW, Northfield TD, Strand MR, et al.: **Organic agriculture promotes evenness and natural pest control**. *Nature*. 2010; **466**(7302): 109–112.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Cuesta D, Taboada A, Calvo L, et al.: **Short- and medium-term effects of experimental nitrogen fertilization on arthropods associated with *Calluna vulgaris* heathlands**. *Environ Pollut*. 2008; **152**(2): 394–402.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Davies BE: **Loss-on-ignition as an estimate of soil organic matter**. *Soil Sci Soc Am J*. 1974; **38**(1): 150–151.
[Publisher Full Text](#)
- Delgado JA, Groffman PM, Nearing MA, et al.: **Conservation practices to mitigate and adapt to climate change**. *Journal of Soil and Water Conservation*. 2011; **66**: 118A–129A.
[Publisher Full Text](#)
- Doll D: **Taking a harvest sample for orchard IPM check-up**. The Almond Doctor, UCANR website. 2009.
[Reference Source](#)
- Drenovsky RE, Steenwerth KL, Jackson LE, et al.: **Land use and climatic factors structure regional patterns in soil microbial communities**. *Glob Ecol Biogeogr*. 2010; **19**(1): 27–39.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Elevitch CR, Mazaroli DN, Ragone D: **Agroforestry standards for regenerative agriculture**. *Sustainability*. 2018; **10**(9): 3337.
[Publisher Full Text](#)
- Fenster T, Oikawa PY, Lundgren JG: **A systems comparison between regenerative and conventional almond production**. *Frontiers in Sustainable Food Systems* submitted. 2021.
- Francis CA, Harwood RR, Parr JF: **The potential for regenerative agriculture in the developing world**. *American Journal of Alternative Agriculture* 1986; **1**(2): 64–74.
[Publisher Full Text](#)
- Frostegård Å, Tunlid A, Bååth E: **Use and misuse of PLFA measurements in soils**. *Soil Biology and Biochemistry*. 2011; **43**(8): 1621–1625.
[Publisher Full Text](#)
- Grant TA, Madden EM, Murphy RK, et al.: **Monitoring native prairie vegetation: the belt transect method**. *Ecological Restoration*. 2004; **22**(2): 106–111.
[Publisher Full Text](#)
- Gulick SH, Grimes DW, Munk DS, et al.: **Cover crop-enhanced water infiltration of a slowly permeable fine sandy loam**. *Soil Science Society of America Journal*. 1994; **58**(5): 1539–1546.
[Publisher Full Text](#)
- Gurda AM, Renz MJ, Brink GE: **Defining mob grazing in the Upper Midwestern United States**. *Journal of Extension*. 2018; **56**: 8.
[Reference Source](#)
- Haney RL, Haney EB, Smith DR, et al.: **The soil health tool-theory and initial broad-scale application**. *Applied Soil Ecology*. 2018; **125**: 162–168.
[Publisher Full Text](#)
- Hendrix PF, Parmalee RW, Crossley DAJ, et al.: **Detritus food webs in conventional and no-tillage agroecosystems**. *Bioscience*. 1986; **36**(6): 374–380.
[Publisher Full Text](#)
- Hillenbrand M, Thompson R, Wang F, et al.: **Impacts of holistic planned grazing with bison compared to continuous grazing with cattle in South Dakota shortgrass prairie**. *Agriculture Ecosystems and Environment*. 2019; **279**: 156–168.
[Publisher Full Text](#)
- HMI: **Holistic Management Institute: The regenerative solution**. Web-based document: 2020.
[Reference Source](#)
- Johnson RM, Dahlgren L, Siegfried BD, et al.: **Acaricide, Fungicide and drug interactions in honey bees (*Apis mellifera*)**. *PLoS One*. 2013; **8**(1): e54092.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Kassam A, Friedrich T, Derpsch R, et al.: **Conservation agriculture in the dry Mediterranean climate**. *Field Crops Res*. 2012; **132**: 7–17.
[Publisher Full Text](#)
- Kromp B: **Carabid beetles in sustainable agriculture: a review on pest control efficacy, cultivation impacts and enhancement**. *Agriculture Ecosystems and Environment*. 1999; **74**(1–3): 187–228.
[Publisher Full Text](#)
- LaCanne CE, Lundgren JG: **Regenerative agriculture: merging farming and natural resource conservation profitably**. *PeerJ*. 2018; **6**: e4428.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Lal R: **Regenerative agriculture for food and climate**. *J Soil Water Conserv*. in press. 2020.
[Publisher Full Text](#)
- Legner EF, Gordh G: **Lower navel orange worm (Lepidoptera: Phycitidae) population densities following establishment of *Goniozus legneri* (Hymenoptera: Bethyloidea) in California**. *J Econ Entomol*. 1992; **85**(6): 2153–5160.
[Publisher Full Text](#)
- Lehman RM, Cambardella CA, Stott DE, et al.: **Understanding and enhancing soil biological health: The solution for reversing soil degradation**. *Sustainability*. 2015; **7**(1): 988–1027.
[Publisher Full Text](#)
- Letourneau DK, Jedlicka JA, Bothwell SG, et al.: **Effects of Natural Enemy Biodiversity on the Suppression of Arthropod Herbivores in Terrestrial Ecosystems**. *Annu Rev Ecol Evol Syst*. 2009; **40**: 573–592.
[Publisher Full Text](#)
- Liu A, Ma BL, Bomke AA: **Effects of cover crops on soil aggregate stability, total organic carbon, and polysaccharides**. *Soil Sci Soc Am J*. 2005; **69**(6): 2041–2048.
[Publisher Full Text](#)
- Lucas Y: **The role of plants in controlling rates and products of weathering: importance of biological pumping**. *Annual Review of Earth and Planetary Science*. 2001; **29**: 135–163.
[Publisher Full Text](#)
- Lundgren J: **Matrix paper**. 2021.
<http://www.doi.org/10.17605/OSF.IO/G697Y>
- Lundgren JG, Fausti SW: **Trading biodiversity for pest problems**. *Sci Adv*. 2015; **1**(6): e1500558.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Lundgren JG, Fergen JK: **Predator community structure and trophic linkage strength to a focal prey**. *Mol Ecol*. 2014; **23**(15): 3790–3798.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Lundgren JG, Shaw JT, Zaborski ER, et al.: **The influence of organic transition systems on beneficial ground-dwelling arthropods and predation of insects and weed seeds**. *Renewable Agriculture and Food Systems*. 2006; **21**(4): 227–237.
[Publisher Full Text](#)
- Mall NK, Herman JD: **Water shortage risks from perennial crop expansion in California's Central Valley**. *Environ Res Lett*. 2019; **14**(10): 104014.
[Publisher Full Text](#)
- Morton HL, Moffett JO, MacDonald RH: **Toxicity of herbicides to newly emerged honey bees**. *Environ Entomol*. 1972; **1**(1): 102–104.
[Publisher Full Text](#)
- Newton P, Civita N, Frankel-Goldwater L, et al.: **What is regenerative agriculture? A review of scholar and practitioner definitions based on processes and outcomes**. *Front Sustain Food Syst*. 2020; **4**: 577723.
[Publisher Full Text](#)
- Niles MT, Garrett RD, Walsh D: **Ecological and economic benefits of integrating sheep into viticulture production**. *Agron Sustain Dev*. 2018; **38**: 1.
[Publisher Full Text](#)
- Patrignani A, Oschner TE: **Canopeo: A powerful new tool for measuring fractional green canopy cover**. *Agron J*. 2015; **107**(6): 2312–2320.
[Publisher Full Text](#)
- Pecenka JR, Lundgren JG: **Effect of cattle management systems on dung arthropod community structure**. *Basic and Applied Ecology*. 2019; **40**: 19–29.

Philippot L, Raaijmakers JM, Lemanceau P, *et al.*: **Going back to the roots: the microbial ecology of the rhizosphere.** *Nat Rev Microbiol.* 2013; **11**(11): 789–799.

[PubMed Abstract](#) | [Publisher Full Text](#)

Pikul JL Jr, Aase JK: **Water infiltration and storage affected by subsoiling and subsequent tillage.** *Soil Sci Soc Am J.* 2003; **67**(3): 859–866.

[Publisher Full Text](#)

Pikul JL Jr, Chilom G, Rice J, *et al.*: **Organic matter and water stability of field aggregates affected by tillage in South Dakota.** *Soil Sci Soc Am J.* 2009; **73**(1): 197–206.

[Publisher Full Text](#)

Rhodes CJ: **The imperative for regenerative agriculture.** *Sci Prog.* 2017; **100**(1): 80–129.

[PubMed Abstract](#) | [Publisher Full Text](#)

ROC Regenerative Organic Certified: **Farm like the world depends on it.** 2020. [Reference Source](#)

Rodale R: **Breaking new ground: the search for a sustainable agriculture.** *The Futurist.* 1983; **17**(1): 15–20.

[Reference Source](#)

Saunders ME, Luck GW, Mayfield MM: **Almond orchards with living ground cover host more wild insect pollinators.** *J Insect Conserv.* 2013; **17**: 1011–1025.

[Publisher Full Text](#)

Schipanski ME, MAdonald GK, Rosenzweig S, *et al.*: **Realizing resilient food systems.** *BioScience.* 2016; **66**(7): 600–610.

[Publisher Full Text](#)

Schmid RB, Lundgren JG: **Regenerative and conventional cattle management on dung beetle performance and parasite management.** *PeerJ submitted.* 2021.

Schreefel L, Schulte RPO, de Boer IJM, *et al.*: **Regenerative agriculture – the soil is the base.** *Glob Food Sec.* 2020; **26**: 100404.

[Publisher Full Text](#)

Seagraves MP, Lundgren JG: **Effects of neonicotinoid seed treatments on soybean aphid and its natural enemies.** *J Pest Sci.* 2012; **85**: 125–132.

[Publisher Full Text](#)

Sherwood S, Uphoff N: **Soil health: research, practice and policy for a more regenerative agriculture.** *Applied Soil Ecology.* 2000; **15**(1): 85–97.

[Publisher Full Text](#)

Soto RL, Martinez-Mena M, Padilla MC, *et al.*: **Restoring soil quality of woody agroecosystems in Mediterranean drylands through regenerative agriculture.** *Agric Ecosyst Environ.* 2021; **306**: 107191.

[Publisher Full Text](#)

Symmes E: **Harvest damage evaluation for almonds, UCANR website.** 2018. [Reference Source](#)

Teague R, Barnes M: **Grazing management that regenerates ecosystem function and grazingland livelihoods.** *Afr J Range Forage Sci.* in press. 2017; **34**(2): 1–10.

[Publisher Full Text](#)

Teague R, Kreuter U: **Managing grazing to restore soil health, ecosystem function, and ecosystem services.** *Front Sustain Food Syst.* 2020; **29**: 534187.

[Publisher Full Text](#)

Teague WR, Apfelbaum S, Lal R, *et al.*: **The role of ruminants in reducing agriculture's carbon footprint in North America.** *J Soil Water Conserv.* 2016; **71**(2): 156–164.

[Publisher Full Text](#)

Teague WR, Dowhower SL, Baker SA, *et al.*: **Grazing management impacts on vegetation, soil biota and soil chemical, physical and hydrological properties in tall grass prairie.** *Agric Ecosyst Environ.* 2011; **141**(3–4): 310–322.

[Publisher Full Text](#)

TerraGenesis: **TerraGenesis International: Regenerative agriculture definition.** 2020.

[Reference Source](#)

UnderstandingAG: **Fact sheet outlining the principles of regenerative agriculture.** 2020.

[Reference Source](#)

USDA-AMS: **USDA Agricultural Marketing Service Organic Standards: USDA Agricultural Marketing Service.** 2020.

[Reference Source](#)

USDA-NRCS: **Soil Health- The five principles of soil health.** 2020.

[Reference Source](#)

Wagner PM, Abagandura GO, Mamo M, *et al.*: **Abundance and diversity of dung beetles (Coleoptera: Scarabaeoidea) as affected by grazing management in the Nebraska sandhills ecosystem.** *Environ Entomol.* in press. 2020.

[Publisher Full Text](#)

Weil R, Brady N: **The Nature and Properties of Soils.** 15th edn. 2017.

[Reference Source](#)

Wendt JW, Hauser S: **An equivalent soil mass procedure for monitoring soil organic carbon in multiple soil layers.** *Eur J Soil Sci.* 2013; **64**(1): 58–65.

[Publisher Full Text](#)

Zak DR, Holmes WE, White DC, *et al.*: **Plant diversity, soil microbial communities, and ecosystem function: are there any links?** *Ecology.* 2003; **84**(8): 2042–2050.

[Publisher Full Text](#)

Open Peer Review

Current Peer Review Status:  

Version 1

Reviewer Report 10 March 2021

<https://doi.org/10.5256/f1000research.31489.r79612>

© 2021 Teague R. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Richard W. Teague

Texas A&M AgriLife Research, Vernon, TX, USA

This review of regenerative agriculture in many parts of US is the most comprehensive and thorough I have yet seen on the subject. It covers a wide array of biological disciplines to assess the various biodiversity elements required to assess the efficacy of regenerative agriculture relative to conventional agriculture.

The authors are to be particularly commended for the solid statistical analyses that provide an excellent foundation for the results and conclusions they present. This is a very welcome and solid contribution to the multiple co-benefits evident from those practicing regenerative agriculture as it is conducted by the agricultural community.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: I am a research scientist in the Texas A&M AgriLife Research organization working on documenting and testing causal mechanisms in comparing results from soil, vegetation, livestock, watershed health, and ecological economics. I have published widely on these aspects of regenerative agriculture with many co-authors for 5 decades.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 09 March 2021

<https://doi.org/10.5256/f1000research.31489.r79606>

© 2021 Holland J et al. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



John M. Holland 

Farmland Ecology Unit, Game and Wildlife Conservation Trust, Fordingbridge, UK

Jenny Bussell

The Allerton Project, Game and Wildlife Conservation Trust, Leicestershire, UK

A regenerative agriculture matrix scoring system is proposed that can be used to evaluate the extent to which a farming system has adopted regenerative practices. This is an important study as it defines a standard for regenerative systems in a similar way to those for other farming systems such as organic using simple scoring matrix that can be used by farmers. This has the advantage that they can monitor the progress of their system over time and adjust accordingly.

This paper represents a substantial amount of work, covering a large number of sites, fully replicated studies with a lot of samples and appropriate experimental designs. It includes three food production systems. The analysis was relevant and is described in good detail, and the statistics employed are robust. The paper uses a novel approach to overcome the problems of comparing the overall outcomes from the regenerative agriculture system between farms, with many different variables. The authors focus on methods that underly the principles of regenerative agriculture rather than defining it as a set of practices. This was achieved by defining a matrix scoring system compiled from sums of regenerative and conventional practices and looking for correlations between the matrix scores with detailed biological and physical measurements. This enabled them to compare a range of systems that exhibit few or many of the regenerative agriculture practices, and regress these with a large number of soil health; plant, insect and microbial diversity; pests and profitability. Much useful information is provided on the different methodologies and their use in farming systems research.

There are some limitations to the approach, especially with respect to farms that may be in transition to regenerative agriculture and are achieving this through slowly reducing reliance on agrochemicals. Therefore, with such farms their matrix scores may remain high despite achieving

substantial decreases in agrochemicals because 4 of the 9 points depending on not using inorganic fertilisers, herbicides, fungicides and insecticides. Nevertheless, the paper provides a contribution to scientific knowledge by allowing some broad comparisons to be made on the overall benefits of the regenerative agriculture approach.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Agroecologist, entomologist, soil scientist

We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research