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A search for quantitative trait loci for ovulation rate in cattle

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Summary

Seventy-seven polymorphic microsatellites were analysed in offspring of three elite sires that were part of the foundation of an experimental population selected for twinning rate at the US Meat Animal Research Center, Clay Center, Nebraska. All females were assessed for ovulation rate by rectal palpation of corpora lutea over 8–10 consecutive oestrous cycles from approximately 12 to 18 months of age, and associations between ovulation rate and sire allele were examined in each of the three sire groups. A preliminary analysis was performed using selectively genotyped daughters of each sire. Markers found significant or approaching significance were also genotyped in all daughters, sons and granddaughters of these sires. A test of marker associations limited to the granddaughter data provided an independent confirmation of marker effect and significance relative to the initial test with daughter data. Putative ovulation rate quantitative trait loci were detected on chromosomes 7 and 23. Marker UWCA20 on chromosome 7 was associated with an effect in excess of one phenotypic standard deviation and accounted for approximately 10% of phenotypic variation ovulation rate. Marker CYP21 (steroid 21-hydroxylase) on chromosome 23 was associated with an effect of slightly less than half a phenotypic standard deviation and accounted for approximately 4% of phenotypic variation.

Keywords: microsatellites, ovulation rate, quantitative trait loci, twinning

Introduction

Reproductive rate has a large impact on production costs of different animal species. For example, an average beef cow produces only 0.7 calves per annum (Bellows *et al.* 1979), a value that compares poorly to 130–150 offspring per annum in meat chickens (Andrews *et al.* 1988). Similarly, when considering cost

efficiency, 33% of cost input for producing a kilogram of protein is attributed to maintenance of the bovine breeding female, compared with approximately 11% in chickens (Dickerson 1978). Through results from experimentation and computer simulation of beef production systems, Dickerson *et al.* (1988) suggested that input costs per unit of beef output could be reduced by 20–30% for that proportion of the herd producing twins compared to single calves. As a consequence, profitability of beef production should improve with increased reproductive rate, provided additional management is available to overcome negative effects associated with twin births (Anderson *et al.* 1979).

The majority of economically important traits, such as reproductive rate in cattle, are thought to be controlled by the combined action of many genes, modified by various environmental factors. These genes are termed quantitative trait loci (QTL). However, for many economic traits there may be only a few genes that account for a large proportion of the genetic variation observed. Therefore, if it were possible to identify QTL responsible for a significant proportion of the genetic variation in reproductive rate in cattle, or at least detect a closely linked genetic marker that is coinherited with the QTL, then selection response could be enhanced by selecting desirable alleles at those loci.

The experimental population that provided data for this study was established in 1981 at the US Meat Animal Research Center (MARC) at Clay Centre, Nebraska (Gregory *et al.* 1990). Current twinning rate in the experimental population is approximately 30%. Twinning rate is approximately 1% for beef breeds and 4% for dairy breeds in which selection has not been practised (Rutledge 1975).

This paper reports results of experiments carried out to determine whether any genes closely linked to a series of molecular markers have a role in increased ovulation rate and twinning frequency within the MARC experimental population. Initially, a candidate gene study was undertaken; this study has now progressed to a more comprehensive genome search involving 77 relatively evenly spaced molecular markers, primarily microsatellites.

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Materials and methods

Animal resources

In this experimental population of approximately 750 calving females, selection has focused on increasing twinning rate by using ovulation rate in puberal heifers as a primary selection criterion. Progeny-proven sires were mated to approximately 25% of the females with highest estimated breeding values (EBV) for twinning, and approximately 30 young sires resulting from these matings were mated to the remaining 75% of the females to produce 6–10 daughter progeny on which ovulation rate was determined over 8–10 consecutive oestrous cycles (Gregory *et al.* 1990).

In terms of marker association work described in this paper, a single-trait animal model was used to estimate breeding value for ovulation rate. Starting at 12–13 months of age, ovulation rate was determined by rectal palpation of corpora lutea for 8–10 consecutive oestrous cycles (Gregory *et al.* 1990). The mean raw ovulation rate (averaged over 8–10 cycles) for the MARC experimental population was 1.145 ova per oestrous cycle with a standard deviation of 0.174. EBVs were calculated for average ovulation rate using best linear unbiased prediction (BLUP) statistical methodology within a complete animal model, fitting significant effects of heifer birth–year–season, age of heifer when ovulation rate was observed, and month of observation. The heritability of ovulation rate, averaged over eight consecutive oestrous cycles, was 0.38 (Echternkamp *et al.* 1990a). The computer program used to estimate breeding value for ovulation rate was the Prediction and Estimation package, PEST (Groeneveld *et al.* 1990).

Animals from three elite sire families (784403, 839802 & 839803, the latter being paternal half-sibs) were selectively genotyped (Darvasi & Soller 1992). These sires were chosen based on their relatively large numbers of daughters and their high EBV for ovulation rate. DNA was extracted from daughters whose EBV fell within the upper and lower 25% range of each sire family. To obtain an unbiased estimate of gene effect, those markers which appeared significant in daughter and granddaughter generations were subsequently genotyped in all daughters from the relevant sire family.

There were also a number of sons of each sire available, which in turn produced a significant number of granddaughters. In total, sire group 784403 consisted of 82 daughters, 15 sons and 116 paternal granddaughters; sire group 839802 consisted of 40 daughters, 12 sons and 158 paternal granddaughters and sire group 839803

consisted of 81 daughters, 24 sons and 236 paternal granddaughters.

DNA typing

DNA was extracted from white blood cells, semen or skin biopsies. Skin biopsy was preferred as a DNA source to eliminate chimerism associated with leucocyte populations from twins. DNA was extracted using the method reported by Mullenbach *et al.* (1989).

Polymerase chain reaction (PCR) was carried out in an MJ Research thermal controller (MJ Research Inc., MA, USA). Aliquots of approximately 50 ng of DNA were amplified ($V_t = 10 \mu\text{l}$) using 20 pmoles ml^{-1} primers, 120 μM dNTPs and 1.5 mM–3 mM MgCl_2 . Reactions were pre-heated to 95°C for 1.5 min, then cycled 4 times at 94°C for 1 min, 48°C–63°C (depending on microsatellite) for 1 min and 72°C for 1 min. The next 22–24 cycles consisted of 90°C for 25 s, 47–62°C for 40 s and 72°C for 45 s, and finally the reaction was maintained at 72°C for 5 min.

After amplification, samples were mixed with 3 μl of loading dye and loaded on 7% non-denaturing polyacrylamide gels and electrophoresed for 1400–3200 volt-hours. Gels were stained with ethidium bromide before being photographed under UV light.

Unless otherwise stated, all markers mentioned in this study have been previously described in mapping papers of Barendse *et al.* (1994), Bishop *et al.* (1994) and Sun *et al.* (1994).

Of the candidate genes examined, both IGF-1 and the putative bovine Fec^B (Montgomery 1993) analogue were identified by microsatellites. An AC dinucleotide repeat was resident within the 5' flanking region of IGF-1, whilst Fec^B was known to be closely linked in sheep (G. W. Montgomery, pers. commun.) to bovine microsatellite BM1329. Growth hormone and INHA were both identified as single-stranded conformational polymorphisms.

The recent development of the bovine genetic map (Barendse *et al.* 1994; Bishop *et al.* 1994) has resulted in a plethora of markers. As a consequence, the study was extended beyond a candidate gene analysis and markers were chosen in an effort to provide a broad genomic coverage across the three sires of at least two markers per chromosome. Once a marker was determined to be associated with ovulation rate, markers flanking that locus were also genotyped.

Statistical analysis

In general, statistical analysis performed in this study was two-stage in nature. Firstly, all infor-

mative markers were selectively genotyped in daughters of the three elite sires. Secondly, in an effort to validate any potential associations, those markers that appeared significant, or approached significance at the 5% level, plus all informative candidate genes, were genotyped in sons and, where informative, in granddaughters. Information from the granddaughter generation was also considered separately as an independent data set. In this case it was appropriate to evaluate significance relative to a one-sided hypothesis based on the results of the initial study.

The parameter that best describes the association between marker genotype and ovulation rate was chromosome substitution effect, difference between ovulation rate of offspring inheriting marker allele A_1 versus those receiving marker allele A_2 from a heterozygous sire (A_1A_2). However, for offspring heterozygous for sire alleles (A_1A_2) it was impossible to determine whether they inherited marker allele A_1 or A_2 . In order to utilize such offspring, genotypic probability was calculated in a manner similar to that described by Dentine & Cowan (1990). For ambiguous offspring, the expected segregation of the sire allele was dependent upon frequency of that allele in the dam population. Allele frequencies were estimated using genotypes of daughters, sons and a sample of 25 individuals from the experimental population (dam contemporaries).

The model used to test for effects of sire alleles was:

$$Y_{ij} = \alpha + b_j(x) + e_{ij}$$

where Y_{ij} = EBV of j^{th} individual for average ovulation rate; α = regression intercept; b_j = chromosome substitution effect sire allele A_1 vs sire allele A_2 ; x = probability of inheriting sire allele A_1 ; e_{ij} = residual deviation not accounted for in model.

Since sire allele inheritance is treated as a probability and incorporated in the model as a covariate, this method permits the inclusion of data from all animals. Given the limited number of observations available for this study, this approach is preferred to the alternative of dropping ambiguous genotypes from the analysis.

Results

In all, 77 markers were genotyped in this study, of which 43 were informative in sire 784403; 45 were informative in sire 839802; and 44 were informative in sire 839803. In addition, eight markers were homozygous in all three sires. Hence, approximately 57% of markers were

informative (sire heterozygous) in any one sire family. An overall genomic distribution of approximately two markers per chromosome across the three sires was achieved for all chromosomes, except chromosomes 24, 26 and 29, which had one marker genotyped on each.

Of the four candidate genes examined, three proved informative in at least one sire. Unfortunately, *INHA* could not be examined because it was homozygous in all three sires. Both *BM1329* and *IGF-1* approached significance in the daughter generation of 839802, whereas growth hormone, informative only in 784403, was not significant ($P = 0.53$).

The ten markers significant in the daughter generation along with two of the three candidate genes, *IGF-1* and *BM1329*, were subsequently genotyped in sons and informative granddaughters (daughters of heterozygous sons). Genotype data for daughters, sons and granddaughters were combined in an analysis, which contrasts the effects of alternative grandsire alleles (Table 1). Only *UWCA4* (-0.042 ± 0.029 ; $P = 0.08$), in 784403 sire group; *UWCA20* (0.12 ± 0.023 ; $P = 0.001$) in 839802 sire group; and *CYP21* (-0.025 ± 0.011 ; $P = 0.05$) in 839803 sire group remained significantly associated with ovulation rate, or approached significance at the 5% level.

Separate analysis of granddaughter data provided confirmatory results in two of three cases. *UWCA20* sire allele 1 was significantly different from sire allele 2 when analysed solely within granddaughters (0.17 ± 0.031 ; $P < 0.0001$) and was also significantly different when analysed within the granddaughter generation against all other dam alleles grouped into a third allele (0.15 ± 0.028 ; $P < 0.0001$). Likewise, *CYP21* remained significant when contrasting sire alleles 1 and 2 in the granddaughter generation (-0.03 ± 0.014 ; $P = 0.05$). However, *UWCA4* (-0.005 ± 0.032 ; $P = 0.83$) was not significant when the analysis was restricted to granddaughter information.

Unfortunately, polymorphism of markers *IGF-1*, *ILSTS001*, *HEL10* and growth hormone was so low that many sons were ambiguous in terms of the sire allele inherited. Thus few granddaughters were informative. In the case of growth hormone, several other linked markers were examined, including *MAP2C*, *CSSM65* and *GFAP*, but in all cases these were uninformative in all three sires.

To further validate the significance of *UWCA20* and *CYP21*, while resolving the uncertainty surrounding *UWCA4*, markers were genotyped that flanked these loci (Table 1). Informative markers flanking *UWCA4* were

ILSTS103 (Kemp *et al.* 1995) and TGLA337; both were non-significant in sire group 784403. Similarly, TGLA122 also mapped to chromosome 21, and although initially significant in daughter analysis ($P = 0.03$) was not significant upon addition of son and granddaughter information ($P = 0.26$). The fact that ILSTS103 in particular was very tightly linked to UWCA4 (W. Barendse, pers. commun.), and, yet not significant, plus the non-significance of this marker in the granddaughter generation, casts considerable doubt on whether UWCA4 was indeed associated with ovulation rate. The discrepancy between significance level for closely linked markers is in part interpretable as a sampling affect related to varying assignment of daughters to ambiguous versus unambiguous sire allele classes and inclusion of different granddaughters in the respective marker analyses.

Table 1. Marker-QTL association for putatively significant markers (a)[†], candidate genes (b) and flanking markers (c)

Marker	Chromosome [‡]	Animals (n)	Estimate (±SEM) [§]	Probability
Sire 784403				
ILSTS103 ^c	21	48	-0.050 ± 0.035	0.15
UWCA4 ^a	21	84	-0.042 ± 0.029	0.08
TGLA337 ^c	21	53	-0.013 ± 0.02	0.50
TGLA122 ^a	21	73	0.024 ± 0.02	0.26
TGLA431 ^a	2	63	0.017 ± 0.02	0.40
IGF-1 ^b	5	57	0.01 ± 0.03	0.66
Sire 839802				
CSSM29 ^c	7	28	0.061 ± 0.04	0.18
UWCA20 ^a	7	78	0.120 ± 0.023	0.0001**
RM006 ^c	7	42	0.077 ± 0.036	0.04*
ILSTS001 ^a	7	33	0.07 ± 0.04	0.10
BM7160 ^c	7	36	0.17 ± 0.05	0.0005***
IGF-1 ^b	5	34	0.03 ± 0.05	0.53
BM1329 ^b	6	58	0.018 ± 0.04	0.62
MAF50 ^a	4	46	-0.014 ± 0.04	0.68
HEL10 ^a	19	35	0.014 ± 0.06	0.80
Sire 839803				
RM002 ^c	23	100	-0.048 ± 0.018	0.006**
CYP21 ^a	23	136	-0.025 ± 0.011	0.05*
RM185 ^c	23	63	-0.046 ± 0.03	0.13
CSSM24 ^c	23	101	0.032 ± 0.026	0.26
TGLA53 ^a	16	61	-0.022 ± 0.032	0.65
BM6108 ^a	12	81	0.037 ± 0.028	0.15
IGF-1 ^b	5	63	0.06 ± 0.054	0.21
BM1329 ^b	6	111	0.004 ± 0.02	0.83

[†] Denotes markers associated with ovulation rate in daughters ($P < 0.10$).

[‡] Based on data generated in this study chromosome 7 and 23 marker maps are (CSSM29 – 8 cM – UWCA20 – 24 cM – RM006 – 8 cM – ILSTS001 – 16 cM – BM7160) and (RM002 – 1 cM – CYP21 – 6 cM – RM185 – 16 cM – CSSM24), respectively.

[§] Sire allele 1 vs sire allele 2.

In sire group 839802, markers flanking UWCA20 on chromosome 7 included CSSM29 (Moore *et al.* 1994), RM006 and BM7160 (C. Beattie, unpubl. obs.). Interestingly, CSSM29 was not significant, RM006 was significant and BM7160 was highly significant. Owing to the large distance (>40 cM, C. Beattie, pers. comm.) between the two highly significant loci, UWCA20 and BM7160, it could be hypothesized there were two separate ovulation rate QTL present on chromosome 7. However, the data presented here are insufficient to resolve the hypothesis of two QTL at a distance on chromosome 7 versus the alternative of a single, intermediately located QTL.

The three markers flanking CYP21 on chromosome 23 in sire group 839803 were RM002, RM185 and CSSM24. RM002 was significantly associated with ovulation rate, whilst RM185 and CSSM24 were not significant. Similarly, UWCA1 investigated in the daughter analysis was also non-significant (-0.044 ± 0.03 ; $P = 0.14$).

Discussion

In two of three elite sire groups there was good evidence for a molecular marker linked to a QTL that accounted for a significant proportion of the genetic variation in ovulation rate. UWCA20 in particular seemed a very robust marker for ovulation rate within the 839802 family. The UWCA20-linked QTL had an effect of greater than one phenotypic standard deviation and accounted for approximately 10% of the phenotypic variation when analysed using raw ovulation rate data. The magnitude of this effect was approximately the same, regardless of whether it was measured in daughters, daughters and granddaughters, granddaughters alone, or indeed against all other dam alleles collectively. The fact that the QTL effect remained consistent in size and significance across generations provides very strong evidence that UWCA20 was linked to a true QTL for ovulation rate within the 839802 sire family.

We hypothesize that two QTLs for ovulation rate may be present on chromosome 7, segregating within sire group 839802. The reasoning behind this is two-fold. Firstly, UWCA20 and BM7160 were separated by greater than 40 cM and as a consequence would be considered essentially unlinked. Secondly, if only one intervening QTL was involved, the intervening markers RM006 ($P = 0.04$) and ILSTS001 ($P = 0.1$) would be expected to have estimates of effect greater than those found with UWCA20 and BM7160. Unfortunately, the lack of informative granddaughters for BM7160 meant that

there was no opportunity to independently validate the putative association with ovulation rate. Given the limited sample size in this study and the varying marker informativeness, it is conceivable that the lesser significance of the intervening markers is a sampling effect. Consequently the results are also compatible with a single QTL centrally located between UWCA20 and BM7160. Analysis of additional data will be required to draw conclusions regarding the presence of one versus two QTL on chromosome 7.

As described in Table 3, CYP21 was significantly associated with ovulation rate, and a very closely linked marker, RM002 (W. Barendse, pers. comm.), was also significant. The CYP21-linked QTL had an effect of less than one half of one phenotypic standard deviation and accounted for approximately 4% of the phenotypic variation when analysed using raw ovulation rate data. Because of the relatively smaller size of this QTL and the relatively small number of animals genotyped, it is not surprising that the somewhat distant markers UWCA1, RM185 and CSSM24 were not significantly associated with ovulation rate. Unlike UWCA20, CYP21 was only significant at the 5% level when analysed in granddaughter data. A possible explanation for this may involve the scenario where linkage between the putative ovulation rate QTL and CYP21 was weakened by recombination evidence in the granddaughter generation.

In an effort to ensure that any significant associations were not merely functions of EBV, analyses employing raw ovulation rate from daughters and granddaughters were also performed. Marker associations were analysed either as described above (substituting raw ovulation rate for EBV, with paternal allele inheritance treated as a covariate) or by analysis of ovulation rate in a mixed effects model that accounted for genetic relationships and treated marker inheritance as a fixed effect. Reassuringly, results from these analyses were consistent with the results observed when using EBVs as data.

In the work described here, a total of 77 genetic markers were used in searching for linkage with ovulation rate, with each marker analysed separately. Because so many multiple comparisons were made, a number of significant associations found in the original daughter analysis may have occurred purely by chance. This problem has been recognized for decades (Niemann-Sorenson & Robertson 1961) and numerous methods have attempted to address it (Cooper 1968; Prentice *et al.* 1984). In general, these methods are reliant upon increasing significance level and although this decreases

incidence of type-I error, it also increases type-II error. Ultimately, the true test of an association is to test the marker in another independent sample, or to select animals on basis of marker genotype and measure them for the trait of interest. In this study, the use of granddaughters as a confirmation tool, plus genotyping flanking markers, were important safeguards in minimizing spurious associations.

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