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Development of a Chimeric *Plasmodium berghei* Strain Expressing the Repeat Region of the *P. vivax* Circumsporozoite Protein for *In Vivo* Evaluation of Vaccine Efficacy

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The development of vaccine candidates against *Plasmodium vivax*—the most geographically widespread human malaria species—is challenged by technical difficulties, such as the lack of *in vitro* culture systems and availability of animal models. Chimeric rodent *Plasmodium* parasites are safe and useful tools for the preclinical evaluation of new vaccine formulations. We report the successful development and characterization of chimeric *Plasmodium berghei* parasites bearing the type I repeat region of *P. vivax* circumsporozoite protein (CSP). The *P. berghei*-*P. vivax* chimeric strain develops normally in mosquitoes and produces highly infectious sporozoites that produce patent infection in mice that are exposed to the bites of as few as 3 *P. berghei*-*P. vivax*-infected mosquitoes. Using this transgenic parasite, we demonstrate that monoclonal and polyclonal antibodies against *P. vivax* CSP strongly inhibit parasite infection and thus support the notion that these antibodies play an important role in protective immunity. The chimeric parasites we developed represent a robust model for evaluating protective immune responses against *P. vivax* vaccines based on CSP.

The global burden of malaria due to *Plasmodium vivax* is approximately 70 to 390 million cases annually (1), with around 2.5 billion people living at risk of infection (2). Even though *Plasmodium falciparum* remains the leading cause of malaria in Africa, *P. vivax* malaria is the most geographically widespread of the human malarias (2, 3). It is extensively distributed throughout the tropics, in the Middle East, Asia, the western Pacific, and Latin America (1–4).

P. vivax malaria was long considered a benign and non-life-threatening disease; however, recent publications report an increasing number of severe, complicated cases due to *P. vivax* infections (5, 6). Efforts toward the development of antimalaria vaccines are still mainly focused on *P. falciparum*, although *P. vivax* malaria is “harder to prevent, diagnose, and treat, and both species are coendemic” (7). While the importance of a vaccine targeting *P. vivax* is recognized, the development of *P. vivax* malaria vaccines is hampered by insufficient funding and technical difficulties (2, 8, 9). The lack of *in vitro* culture systems and suitable animal models, for example, imposes a significant limitation on testing novel vaccine candidates (7, 10).

The circumsporozoite protein (CSP) is the most abundant protein on the surfaces of sporozoites and is responsible for eliciting both T-cell and antibody responses. The plasmodial CSP is one of the best studied antigens and is considered to be a major malaria vaccine candidate. RTS,S, the most effective malaria vaccine candidate to date, is based on *P. falciparum* CSP, thus underscoring CSP’s immunogenic properties. CSP is comprised of an immunodominant central repeat region, diverse among *Plasmodium* species, flanked by two conserved domains: region I at the N terminus of the repeats and a type I thrombospondin repeat (TSR) motif C terminal to the repeat region. Early studies in animals have indicated that antibodies against the repeat region neutralize the infectivity of sporozoites and confer sterile immunity (11, 12). Importantly, recent studies in humans vaccinated with RTS,S in-

dicated that protection among vaccinees correlates with the titer of anti-CSP repeat antibodies (13).

Plasmodium berghei parasites bearing the repeat region of *P. falciparum* CSP have been developed and characterized by Persson et al. (14). These parasites, “antigenically *P. falciparum* but functionally rodent malaria” (14), are a practical tool for evaluating the efficacy of human preerythrocytic *P. falciparum* CSP-based vaccine candidates. On the basis of the study by Persson et al., we developed and characterized a chimeric *P. berghei* parasite strain expressing the repeat region of *P. vivax* (VK210) CSP as a first step in evaluating vaccine candidates based on the CSP of *P. vivax*. We demonstrate that these chimeric parasites are recognized by a monoclonal antibody (MAb), 2F2, specific for the *P. vivax* VK210 CSP repeats (15) and that the *in vivo* infectivity of chimeric sporozoites is strongly inhibited by this monoclonal antibody, as well as by polyclonal antibodies raised against a recombinant *P. vivax* CSP construct (16).

MATERIALS AND METHODS

Plasmid construction and parasite transfection. The transfection plasmid, pR-CSPv, carrying the repeat region of *P. vivax*, was derived from plasmid pR-CSwt (17), which carries the CS gene of *P. berghei* (wild type [WT]) and results from the combination of plasmids pIC-CSwt (18) and pPv (Retrogen Inc., CA) encoding the VK210 repeat motif based on the Salvador I (SalI) strain of *P. vivax*. Briefly, a PmlI-SexAI 567-bp fragment was excised from pIC-CSwt and replaced with a PmlI-SexAI 675-bp frag-

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ment comprising the *P. vivax* VK210 CSP repeat region, which was codon harmonized to optimize protein synthesis and folding in the *P. berghei* transgenic host (19) and was excised from the engineered pPv. From the resulting intermediate plasmid, pIC-CSPv, the entire CSP gene was excised as a KpnI-XhoI fragment and inserted into the transfection plasmid, pR-CSPv. KpnI and SacI were used to linearize pR-CSPv prior to transfection; WT *P. berghei* (strain ANKA) parasites were transfected as previously described (20).

Chimeric parasites were selected in Swiss Webster mice (NCI, Frederick, MD) by treatment with pyrimethamine in drinking water (0.07 mg/ml). Parasites surviving drug treatment were then cloned by means of a limiting-dilution assay (21). Recombination at the 5' and 3' ends of the locus was verified by PCR. The primers used to verify integration at the 5' end of the construct are 5'F (TCACCCTCAAGTTGGGTA AAA) and 5'R (CCTGTCCCCTGGTTGCTTA); the primers to verify integration at the 3' end are 3'F (TGTA AAAATGTGTATGTTGTGTGC) and 3'R (GTGCCATTACGACTTTGCT). To verify that the isolated clone did not have contaminating WT *P. berghei* parasites, a PCR product flanking the SexAI restriction site was digested with SexAI. The primers used for this PCR are PbCS-F (AAGAAAGCAGAAGATTTGACCTT) and PbCS-R (AAGGTC AAATCTTCTGCTTTCTT). Finally, DNA isolated from the cloned transgenic parasites was sequenced (Macrogen Inc., USA) to confirm the replacement of the *P. berghei* repeat region with the *P. vivax* repeat region for the generation of *P. berghei*-*P. vivax* chimeric parasites.

Mosquito infection and parasite development. *Anopheles stephensi* mosquitoes were fed on Swiss Webster mice (NCI, Frederick, MD) infected with blood stages of cloned *P. berghei*-*P. vivax* CS_{Repeat} (*P. berghei*-*P. vivax*) parasites. Development of viable male gametocytes in blood was qualitatively assessed by means of an exflagellation assay. Briefly, 1.5 µl of blood from a mouse with patent blood parasitemia was incubated at room temperature for 20 min with 1.5 µl of heparin (40 U/ml in 1× phosphate-buffered saline [PBS]) and 7 µl of complete ookinete medium (RPMI 1640 medium with 10% fetal calf serum, 25 mM HEPES, 2 mM glutamine, 0.2 mM hypoxanthine, 12 mM sodium bicarbonate, 1 µM xanthurenic acid). After incubation, the mixture was placed on a slide and observed under a light microscope (×100 magnification).

Development of oocysts in mosquito midguts was assessed 14 days post-blood meal. The midguts were dissected in sterile PBS and stained in a 0.1% mercury chromate solution.

Infectivity of *P. berghei*-*P. vivax* sporozoites. The *in vivo* infectivity of chimeric *P. berghei*-*P. vivax* sporozoites was assessed in 5- to 8-week-old C57BL/6 mice both by assessing the liver parasite load, measured by quantitative PCR (qPCR) (22), in mice that were challenged intravenously (i.v.) with 1×10^4 *P. berghei*-*P. vivax* sporozoites and by the development of blood-stage parasites after feeding with 3 *P. berghei*-*P. vivax*-infected *A. stephensi* mosquitoes for 3 min.

Antibodies. Development of 3D11, a monoclonal antibody directed against the repeat region of CSP of *P. berghei* (23), and 2F2, a monoclonal antibody directed against the repeat region of *P. vivax* CSP (24), has been previously described.

Polyclonal antibodies against *P. vivax* CSP were generated by immunizing New Zealand White rabbits with 25 µg VMP001 (16) in complete Freund's adjuvant, followed by 2 booster immunizations with 25 µg VMP001 in incomplete Freund's adjuvant. Serum obtained post-3rd immunization was pooled and used in the present study.

IFA. An indirect immunofluorescence assay (IFA) was used to characterize *P. berghei*-*P. vivax* sporozoites using monoclonal antibodies. Briefly, 10 µl of a sporozoite suspension (4×10^5 to 6×10^5 sporozoites/ml) was air dried on poly-L-lysine-covered slides (Tekdon Inc., Myakka City, FL) and incubated for 30 min at room temperature with 1 µg/ml of 3D11 or 2F2 monoclonal antibody against the *P. berghei* (23) or the *P. vivax* (15, 24, 25) CSP repeat region, respectively. The slides were then washed with PBS-1% bovine serum albumin (BSA) and incubated for 30 min at room temperature with a secondary-antibody solution [Alexa Fluor 488 F(ab')₂ fragment of goat anti-mouse IgG(H+L); 2 mg/ml; In-

vitrogen]. Green-fluorescent sporozoites were visualized under an upright fluorescence microscope (Nikon Eclipse 90i).

Inhibition of sporozoite infectivity by passive transfer of monoclonal antibodies or polyclonal rabbit sera directed against *P. vivax* CSP. Three hundred micrograms of 2F2 antibody specific for the repeat region of *P. vivax* (15) or 0.5 ml pooled sera from rabbits immunized with VMP001, a recombinant *P. vivax* CSP (16), were passively transferred into 5- to 8-week-old C57BL/6 mice i.v. immediately prior to sporozoite challenge. Following antibody transfer, the mice were challenged with 1×10^4 sporozoites delivered by tail vein injection into each mouse. Forty-two hours after challenge, livers were harvested to assess the parasite burden by qPCR. In some experiments, mice receiving monoclonal antibody were challenged by mosquito bites. For this purpose, 5- to 8-week-old C57BL/6 mice were anesthetized by intraperitoneal injection of 250 µl of 2% avertin, and then 400 µg of 2F2 antibody was passively transferred i.v. prior to feeding *P. berghei*-*P. vivax*-infected *A. stephensi* mosquitoes. For increased stringency, we allowed 4 mosquitoes to feed for 5 min and then visually examined them for a blood feed. The presence of sporozoites was then assessed in those mosquitoes that fed to ensure that mice were exposed to the bite of at least 1 infected mosquito.

Data analysis. Most of the data were plotted using Prism 4 software (GraphPad). Unless otherwise stated, data were compared for significance using a Mann-Whitney test.

RESULTS

Generation and biological characterization of *P. berghei*-*P. vivax* chimeric parasites. Using a plasmid containing the sequences of the *P. berghei* CSP gene and a modified version of the dihydrofolate reductase (DHFR) gene, which confers resistance to pyrimethamine on transfected parasites, we generated *P. berghei*-*P. vivax* chimeric parasites. Briefly, we replaced the repeat region of *P. berghei* CSP with the repeat domain of *P. vivax* CSP. The *P. vivax* replacement represents the VK210 CSP sequence of the *P. vivax* Sall strain, from nucleotides 268 to 825 (amino acids 90 to 275). *P. berghei* ANKA blood stages were transfected *in vitro* with a plasmid linearized using KpnI and XhoI restriction enzymes, as described previously (21) (Fig. 1A). The transfected parasites were injected intravenously into Swiss Webster mice, and transgenic parasites were selected under pyrimethamine treatment. Ten days later, drug-resistant parasites were recovered and cloned by limiting dilution.

PCR analysis was used to confirm recombination at the 5' and 3' ends for gene replacement (Fig. 1B), and the cloned parasites were verified by PCR-restriction fragment length polymorphism (RFLP) (Fig. 1C). The cloned parasites were sequenced to further confirm that the *P. berghei*-*P. vivax* parasites have the correct insertion in the CSP gene (Fig. 1D).

Mosquito infection. A cloned *P. berghei*-*P. vivax* parasite isolate was used to infect mice, which were then used to feed *A. stephensi* mosquitoes. Transgenic parasites were evaluated by using standard methods, such as exflagellation and oocyst and sporozoite counts, to compare them to the wild-type *P. berghei*.

Exflagellation of male *P. berghei*-*P. vivax* gametocytes, assessed prior to feeding female *A. stephensi* mosquitoes on infected mice, was normal and comparable to that of WT *P. berghei* ANKA parasites (data not shown). Twelve to 14 days after mosquito feeding, *P. berghei*-*P. vivax*-infected mosquito midguts had approximately 100 to 150 healthy-looking oocysts per midgut, which was also comparable to WT *P. berghei* ANKA parasites. Finally, on day 21, mosquito salivary glands were dissected, and the parasite load was found to be comparable to that of WT *P. berghei* ANKA (Table 1).

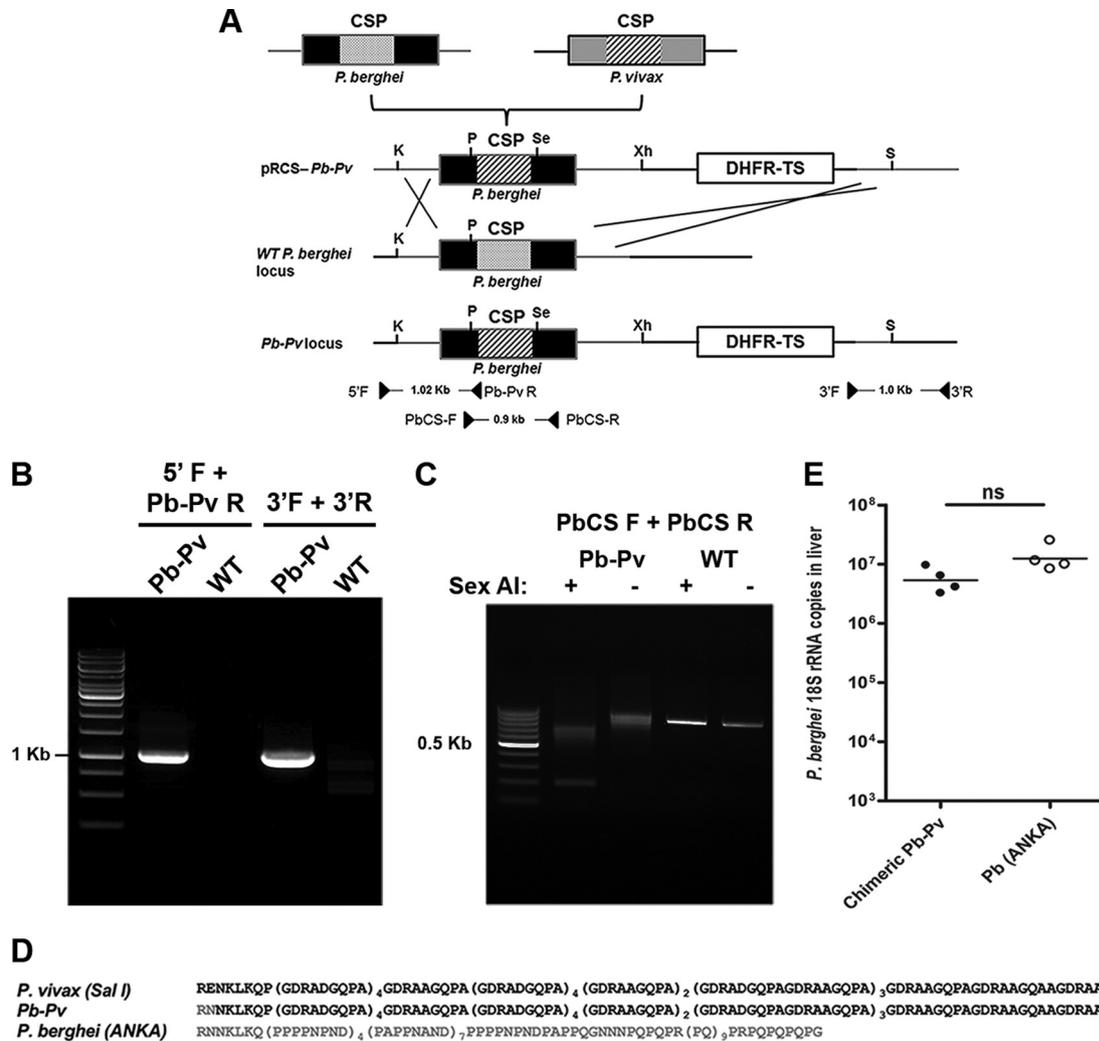


FIG 1 Generation of *P. berghei-P. vivax* (Pb-Pv) chimeric parasites. (A) Schematic representation of the approach used for replacing the CSP gene of *P. berghei* ANKA with the *P. vivax* SalI repeat region. The locations of the primers used to verify recombination by PCR are indicated at the bottom. The restriction sites shown are KpnI (K), PmlI (P), SexAI (Se), XhoI (Xh), and SacI (S). (B) Integration at the 5' and 3' ends of the replacement fragment was verified by PCR using genomic DNA from *P. berghei-P. vivax* parasites. Primers 5'F and Pb-Pv R yield a 1,020-bp product; primers 3'F and 3'R yield a 1,000-bp product. (C) To demonstrate that the parasite population was clonal, a 908-bp PCR fragment was amplified from the CSP gene using primers PbCS-F and PbCS-R. The PCR product was then digested with SexAI, which does not cut the WT CSP gene. Genomic DNA from *P. berghei* ANKA was used as a control. (D) Comparison of CSP amino acid sequences of *P. berghei* ANKA, *P. berghei-P. vivax*, and *P. vivax* strain Salvador I; the *P. berghei-P. vivax* amino acid sequence was derived from DNA sequencing of clonal transgenic parasites. (E) *In vivo* infectivity of *P. berghei-P. vivax* sporozoites compared to WT *P. berghei* ANKA sporozoites, both injected intravenously. ns, not significant; horizontal bars, geometric means.

TABLE 1 Developmental characteristics and infectivity of *P. berghei-P. vivax* parasites in *A. stephensi* mosquitoes^a

Parasite	% of midgut infected	No. of oocysts/midgut	% of salivary gland infected	No. of sporozoites/salivary gland (10 ³)	% of mice infected after mosquito bites ^b
<i>P. berghei-P. vivax</i>	80.72	54.76	75	8.5	100
<i>P. berghei</i> ANKA	75.14	42.25	66	10	100

^a Mean of 3 experiments, with at least 20 mosquitoes examined in each experiment.
^b Three infected mosquitoes were allowed to feed on C57BL/6 mice for 3 min. Blood-stage parasitemia was assessed 5 days later.

Characterization of *P. berghei-P. vivax* sporozoites with monoclonal antibodies. Sporozoites obtained from salivary glands of *P. berghei-P. vivax*-infected mosquitoes were used for IFA with antibodies specific for the repeat domain of *P. vivax* (2F2) CSP. An antibody specific for the repeat region of *P. berghei* (3D11) served as a control. The IFA results clearly demonstrated that the chimeric *P. berghei-P. vivax* sporozoites were all uniformly positive for the anti-*P. vivax* repeat antibodies and negative for the antibody against *P. berghei* repeats (Fig. 2).

Infectivity of *P. berghei-P. vivax* chimeric parasites in mice. To evaluate the *in vivo* fitness of the transgenic parasites, we challenged C57BL/6 mice with 1 × 10⁴ sporozoites, delivered by i.v. injection, from either WT *P. berghei* ANKA or *P. berghei-P. vivax* parasites. Forty-two hours after infection, the parasite load in the liver was measured by qPCR specific for the *P. berghei* 18S rRNA.

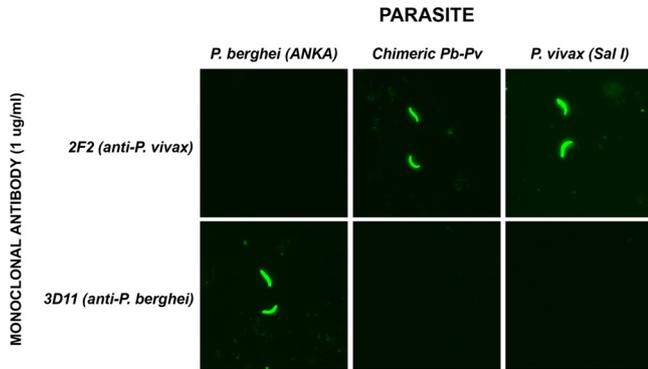


FIG 2 Characterization of *P. berghei-P. vivax* sporozoites using anti-repeat monoclonal antibodies. Monoclonal antibody 2F2, targeting the repeat region of *P. vivax* CSP VK210, specifically recognizes *P. berghei-P. vivax* chimeric sporozoites and *P. vivax* SalI sporozoites. No cross-reactivity was observed against WT *P. berghei* ANKA sporozoites. Monoclonal antibody 3D11, specific for the repeat region of *P. berghei*, reacts to wild-type *P. berghei*, but not to *P. vivax* or *P. berghei-P. vivax* chimeric sporozoites.

The data indicated that the infectivities of WT *P. berghei* ANKA and *P. berghei-P. vivax* sporozoites were comparable, and there was no statistically significant difference between the parasites.

The infectivity of the transgenic parasites was further confirmed by delivering the parasites via the natural route, i.e., using mosquito bites. Mice exposed to the bites of 3 infected mosquitoes demonstrated patent blood-stage infection by day 5 postchallenge.

Anti-*P. vivax* antibodies inhibit the development of parasite liver stages. In order to evaluate the utility of the transgenic par-

asites as a tool to assess the efficacy of future vaccines, we tested the neutralization capabilities of anti-*P. vivax* antibodies *in vivo*. The anti-repeat monoclonal antibody 2F2 was passively transferred into mice immediately prior to an intravenous challenge with 1×10^4 chimeric *P. berghei-P. vivax* sporozoites. As seen in Fig. 3, mice that received 2F2 antibody showed a significant reduction in parasite load in the liver (99% reduction compared to naive controls). In contrast, the group that received the irrelevant antibody had a parasite load comparable to that of naive controls, confirming the specificity of the results observed with the anti-*P. vivax* MAb 2F2. To determine the effects of polyclonal antibodies that contain a mixture of specificities (as opposed to the repeat-specific monoclonal antibody 2F2), we evaluated the neutralization capability of rabbit polyclonal serum that was generated against a recombinant *P. vivax* CSP protein, VMP001. Rabbit anti-VMP001 antibodies recognized CSP on the surfaces of *P. vivax* sporozoites and reacted to the whole protein (VMP001), in addition to the N-terminal and C-terminal proteins, as well as a synthetic type 1 peptide. As shown in Fig. 3B, similar to MAb 2F2, polyclonal rabbit sera also reduced the liver parasite burden in comparison to mice that either were naive or received preimmune rabbit serum. When mice receiving 400 μ g of monoclonal antibody 2F2 were exposed to the bites of 4 *P. berghei-P. vivax*-infected mosquitoes for 5 min, we did not observe sterile protection. However, control mice ($n = 5$) receiving an irrelevant monoclonal antibody developed parasitemia at day 4, while mice receiving anti-parasite monoclonal antibody ($n = 5$) became patent at day 5. Taken together, these results demonstrate that *P. berghei-P. vivax* sporozoites are highly infectious and a stringent model to evaluate immune responses

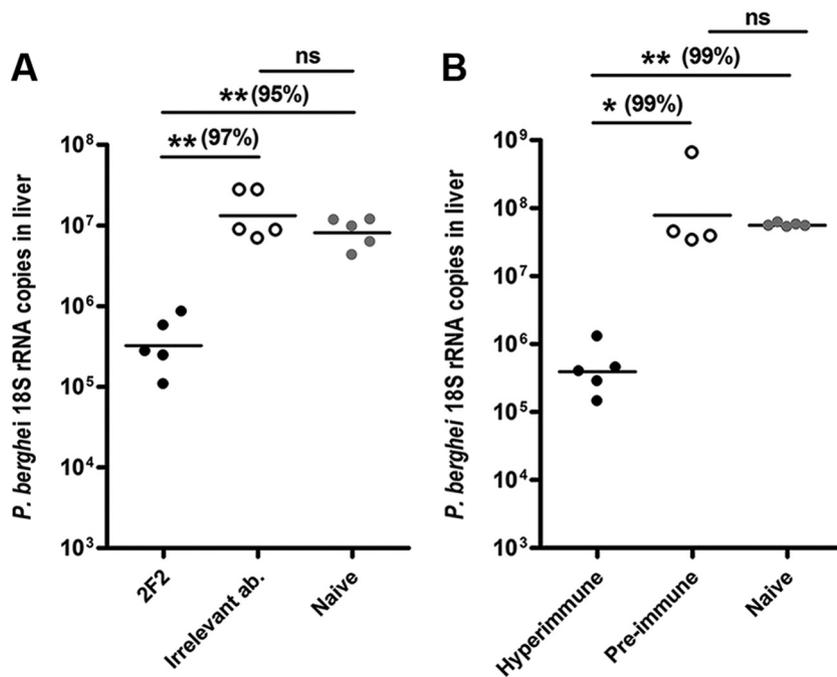


FIG 3 Protection against *P. berghei-P. vivax* sporozoite challenge by passive transfer of monoclonal antibodies or hyperimmune sera. Following passive transfer of monoclonal antibodies or sera, mice were challenged intravenously with 1×10^4 *P. berghei-P. vivax* sporozoites. Forty hours after challenge, livers were harvested to determine parasite burdens by qPCR. Passive transfer of monoclonal antibody 2F2 (A) or rabbit polyclonal sera (B) had a significant protective effect against *P. berghei-P. vivax* sporozoites compared to mice that received irrelevant antibody (ab.) or preimmune sera or to naive controls. The horizontal bars show the geometrical means. *, $P < 0.05$; **, $P < 0.01$. The percent parasite burden inhibition in the liver is shown in parentheses. The results are representative of at least two independent experiments. Each group consisted of 4 or 5 mice.

that inhibit parasite development in the liver, as well as the acquisition of sterile immunity after immunization with vaccine candidates.

DISCUSSION

Studies on assessing protective immune responses against malaria in general, and *P. vivax* in particular, are difficult due to the inability to culture the parasite and to the lack of a suitable animal model. Human studies are expensive, and therefore, this is not a practical first step to evaluate vaccine candidates. Over the last decade or so, transfection of *Plasmodium* has revolutionized the ways in which protective immune responses can be evaluated using transgenic rodent parasites that have been transfected with the gene encoding a specific human malaria antigen (14, 26–29). Active- and passive-immunization studies directed toward these antigens are more indicative of a biologically relevant immune response than the evaluation of immunogenicity alone. Transgenic parasites carrying the CSP, MSP, and P25 genes of *P. falciparum* have been developed and are being used to evaluate vaccine candidates and/or platforms (14, 26–29). To date, only one transgenic *P. berghei* sporozoite expressing P25 from *P. vivax* has been developed as a tool for evaluating the efficacy of vaccine candidates (29).

Here, we describe the successful development, characterization, and applicability of chimeric *P. berghei* parasites expressing the repeat region of *P. vivax* CSP. We successfully replaced the *P. berghei* repeat region with the VK210 repeat region of *P. vivax*. The resulting parasites produce viable gametocytes and successfully infect *A. stephensi* mosquitoes, in which their development is comparable to that of WT *P. berghei*. The *in vivo* infectivity of *P. berghei-P. vivax* sporozoites is comparable to that of WT *P. berghei* in C57BL/6 mice. Moreover, feeding of *P. berghei-P. vivax*-infected mosquitoes on naive C57BL/6 mice results in the development of patent blood-stage parasitemia. Following immunization, the true potential of a vaccine needs to be evaluated using a natural route of sporozoite inoculation, which allows the participation of all components of the immune response in protection.

We used the transgenic parasites to evaluate their applicability as a tool for assessing the efficacies of antibodies in protection. Passive transfer of monoclonal antibodies or hyperimmune sera into naive recipients conferred significant protection against *P. berghei-P. vivax* sporozoite challenge. Mice that received 2F2 antibody or hyperimmune rabbit sera both showed above 95% inhibition of the parasite burden in the liver compared to the naive controls, thus proving that the transgenic parasites can serve as an important tool in evaluating protective anti-repeat immune response. Given that sporozoites rapidly migrate through the circulatory system (30), the availability and affinity of circulating anti-sporozoite antibodies determine their protective capacity. While the monoclonal antibody 2F2 was able to decrease liver infection by 90% of the sporozoites—as shown by measurement of the parasite load in the liver—and delayed the prepatent period by 1 day, it does not appear to be efficient enough to confer sterile immunity. Further comparative studies will be necessary to define the dosage, avidity, and fine specificity of different monoclonal and polyclonal antibodies capable of conferring sterile immunity. The possibility that sterile immunity was not achieved due to generation of mutants or revertant parasites is unlikely; however, this is an issue worth considering for future studies. Taken together, these results demonstrate that the *P. berghei-P. vivax* parasites are

robust and that a natural mosquito bite challenge can be used as a stringent test to assess the full potential of vaccine candidates. Passive-transfer experiments in mice using serum samples from volunteers immunized with novel vaccine candidates could potentially be used to assess humoral protective capacity against *P. berghei-P. vivax* sporozoites. Such studies should also be useful for determining more accurate correlates between antibody titers and protective efficacy against *P. vivax* infection.

Chimeric rodent malaria parasites are a simple, robust, and practical tool for the *in vivo* evaluation of vaccine candidates against malaria. This is the first report of the development of a rodent parasite that can be used to evaluate the efficacy of the circumsporozoite protein central repeat region of *P. vivax*. As the replacement of the entire CSP gene could result in parasites with reduced infectivity in both mice and mosquitoes (31), we chose to first develop chimeric parasites in which we replaced only the *P. berghei* CSP repeat domain with that of *P. vivax*, which is a well-defined target of protective antibody responses. The successful generation of parasites that are able to reproducibly infect mice opens up the road to develop additional transgenic parasites expressing the N-terminal and C-terminal regions and, if possible, the full-length CSP to evaluate T-cell-mediated immune-protective mechanisms.

Such parasites will help select future vaccine candidates for clinical development. Over the last decade, the utility of chimeric rodent parasites has been highlighted by an increasing number of studies that report their use for the assessment of novel protein constructs, recombinant viral vectors, and synthetic adjuvant formulations (32–37). The *P. berghei-P. vivax* parasites we have generated should also facilitate the appraisal of new vaccine candidates and represent a practical platform for evaluating CSP-based protective immune responses against *P. vivax*.

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REFERENCES

- Price RN, Tjitra E, Guerra CA, Yeung S, White NJ, Anstey NM. 2007. Vivax malaria: neglected and not benign. *Am. J. Trop. Med. Hyg.* 77:79–87.
- Gething PW, Elyazar IRF, Moyes CM, Smith DL, Battle KE, Guerra CA, Patil AP, Tatem AJ, Howes RE, Myers MF, George DB, Horby P, Wertheim HFL, Price RN, Mueller I, Baird JK, Hay SI. 2012. A long neglected world malaria map: Plasmodium vivax endemicity in 2010. *PLoS Negl. Trop. Dis.* 6:e1814. doi:10.1371/journal.pntd.0001814.
- Arévalo-Herrera M, Chitnis C, Herrera S. 2010. Current status of Plasmodium vivax vaccine. *Hum. Vaccin.* 6:124–132.
- Mendis K, Sina BJ, Marchesini P, Carter R. 2001. The neglected burden of Plasmodium vivax malaria. *Am. J. Trop. Med. Hyg.* 64:97–106.
- Baird JK. 2007. Neglect of Plasmodium vivax. *Trends Parasitol.* 23:533–539.
- Baird JK. 2013. Evidence and implications of mortality associated with acute Plasmodium vivax malaria. *Clin. Microbiol. Rev.* 26:36–57.
- Carlton JM, Sina BJ, Adams JH. 2011. Why is Plasmodium vivax a neglected tropical disease? *PLoS Negl. Trop. Dis.* 5:e1160. doi:10.1371/journal.pntd.000160.
- Pigott DM, Atun R, Moyes CL, Hay SI, Gething PW. 2012. Funding for malaria control 2006–2010: a comprehensive global assessment. *Malar. J.* 11:246.
- Battle KE, Gething PW, Elyazar IR, Moyes CL, Sinka ME, Howes RE, Guerra CA, Price RN, Baird JK, Hay SI. 2012. The global public health significance of Plasmodium vivax. *Adv. Parasitol.* 80:1–111.

10. Herrera S, Corradin G, Arévalo-Herrera M. 2007. An update on the search for a *Plasmodium vivax* vaccine. *Trends Parasitol.* 23:122–128.
11. Tam JP, Clavijo P, Lu YA, Nussenzweig V, Nussenzweig R, Zavala F. 1990. Incorporation of T and B epitopes of the circumsporozoite protein in a chemically defined synthetic vaccine against malaria. *J. Exp. Med.* 171:299–306.
12. Zavala F, Tam JP, Romero PJ, Ley V, Nussenzweig RS, Nussenzweig V. 1987. Synthetic peptide vaccine confers protection against murine malaria. *J. Exp. Med.* 66:1591–1596.
13. Kester KE, Cummings JF, Ofori-Anyiam O, Ockenhouse CF, Krzych U, Moris P, Schwenk R, Nielsen RA, Debebe Z, Pinelis E, Juompan L, Williams J, Dowler M, Stewart VA, Wirtz RA, Dubois MC, Lievens M, Cohen J, Ballou WR, Heppner DG, Jr, RTS,S. Vaccine Evaluation Group. 2009. Randomized, double-blind, phase 2a trial of falciparum malaria vaccines RTS,S/AS01B and RTS,S/AS02A in malaria-naïve adults: safety, efficacy, and immunologic associates of protection. *J. Infect. Dis.* 200:337–346.
14. Persson C, Oliveira GA, Sultan AA, Bhanot P, Nussenzweig V, Nardin EH. 2002. Cutting edge: a new tool to evaluate human pre-erythrocytic malaria vaccines: rodent parasites bearing a hybrid *Plasmodium falciparum* circumsporozoite protein. *J. Immunol.* 169:6681–6685.
15. Romero P, Heimer EP, Herrera S, Felix AM, Nussenzweig RS, Zavala F. 1987. Antigenic analysis of the repeat domain of the circumsporozoite protein of *Plasmodium vivax*. *J. Immunol.* 139:1679–1682.
16. Yadava A, Sattabongkot J, Washington MA, Ware LA, Majam V, Zheng H, Kumar S, Ockenhouse CF. 2007. A novel chimeric *Plasmodium vivax* circumsporozoite protein induces biologically functional antibodies that recognize both VK210 and VK247 sporozoites. *Infect. Immun.* 75:1177–1185.
17. Wang Q, Fujioka H, Nussenzweig V. 2005. Exit of *Plasmodium* sporozoites from oocysts is an active process that involves the circumsporozoite protein. *PLoS Pathog.* 1:e9. doi:10.1371/journal.ppat.0010009.
18. Cockburn IA, Tse S-W, Radtke AJ, Srinivasan P, Chen Sinnis Y-CP, Zavala F. 2011. Dendritic cells and hepatocytes use distinct pathways to process protective antigen from *Plasmodium* in vivo. *PLoS Pathog.* 7:e1001318. doi:10.1371/journal.ppat.1001318.
19. Angov E, Hillier CJ, Kincaid RL, Lyon JA. 2008. Heterologous protein expression is enhanced by harmonizing the codon usage frequencies of the target gene with those of the expression host. *PLoS One* 3:e2189. doi:10.1371/journal.pone.0002189.
20. Janse CJ, Ramesar J, Waters AP. 2006. High-efficiency transfection and drug selection of genetically transformed blood stages of the rodent malaria parasite *Plasmodium berghei*. *Nat. Protoc.* 1:346–356.
21. Menard R, Janse CJ. 1997. Gene targeting in malaria parasites. *Methods* 13:148–157.
22. Bruña-Romero O, Hafalla JC, González-Aseguinolaza G, Sano G, Tsuji M, Zavala F. 2001. Detection of malaria liver-stages in mice infected through the bite of a single *Anopheles* mosquito using a highly sensitive real-time PCR. *Int. J. Parasitol.* 31:1499–1502.
23. Yoshida N, Nussenzweig RS, Potocnjak P, Nussenzweig V, Aikawa M. 1980. Hybridoma produces protective antibodies directed against the sporozoite stage of malaria parasite. *Science* 207:71–73.
24. Nardin EH, Nussenzweig V, Nussenzweig RS, Collins WE, Harinasuta KT, Tapchaisri P, Chomcharn Y. 1982. Circumsporozoite protein of human malaria parasites *Plasmodium falciparum* and *Plasmodium vivax*. *J. Exp. Med.* 156:20–30.
25. Zavala F, Cochrane AH, Nardin EH, Nussenzweig RS, Nussenzweig V. 1983. Circumsporozoite proteins of malaria parasites contain a single immunodominant region with two or more identical epitopes. *J. Exp. Med.* 157:1947–1957.
26. Cao Y, Zhang D, Pan W. 2009. Construction of transgenic *Plasmodium berghei* as a model for evaluation of blood-stage vaccine candidate of *Plasmodium falciparum* chimeric protein 2.9. *PLoS One* 4:e6894. doi:10.1371/journal.pone.0006894.
27. de Koning-Ward TF, O'Donnell RA, Drew DR, Thomson R, Speed TP, Crabb BS. 2003. A new rodent model to assess blood stage immunity to the *Plasmodium falciparum* antigen merozoite surface protein 119 reveals a protective role for invasion inhibitory antibodies. *J. Exp. Med.* 198:869–875.
28. Mlambo G, Maciel J, Kumar N. 2008. Murine model for assessment of *Plasmodium falciparum* transmission-blocking vaccine using transgenic *Plasmodium berghei* parasites expressing the target antigen Pfs25. *Infect. Immun.* 76:2018–2024.
29. Ramjane S, Robertson JS, Franke-Fayard B, Sinha R, Waters AP, Janse CJ, Wu Y, Blagborough AM, Saul A, Sinden RE. 2007. The use of transgenic *Plasmodium berghei* expressing the *Plasmodium vivax* antigen P25 to determine the transmission-blocking activity of sera from malaria vaccine trials. *Vaccine* 25:886–894.
30. Amino R, Thiberge S, Martin B, Celli S, Shorte S, Frischknecht F, Menard R. 2006. Quantitative imaging of *Plasmodium* transmission from mosquito to mammal. *Nat. Med.* 12:220–224.
31. Tewari K, Spaccapelo R, Bistoni F, Holder AA, Crisanti A. 2002. Function of region I and II adhesive motifs of *Plasmodium falciparum* circumsporozoite protein in sporozoite motility and infectivity. *J. Biol. Chem.* 277:47613–47618.
32. Nardin EH, Oliveira GA, Calvo-Calle JM, Wetzel K, Maier C, Birkett AJ, Sarpotdar P, Corado ML, Thornton GB, Schmidt A. 2004. Phase I testing of a malaria vaccine composed of hepatitis B virus core particles expressing *Plasmodium falciparum* circumsporozoite epitopes. *Infect. Immun.* 72:6519–6527.
33. Calvo-Calle JM, Oliveira GA, Watta CO, Soverow J, Parra-Lopez C, Nardin EH. 2006. A linear peptide containing minimal T- and B-cell epitopes of *Plasmodium falciparum* circumsporozoite protein elicits protection against transgenic sporozoite challenge. *Infect. Immun.* 74:6929–6939.
34. Oliveira GA, Wetzel K, Calvo-Calle JM, Nussenzweig R, Schmidt A, Birkett A, Dubovsky F, Tierney E, Gleiter CH, Boehmer G, Luty AJ, Ramharter M, Thornton GB, Kreamsner PG, Nardin EH. 2005. Safety and enhanced immunogenicity of a hepatitis B core particle *Plasmodium falciparum* malaria vaccine formulated in adjuvant Montanide ISA 720 in a phase I trial. *Infect. Immun.* 73:3587–3597.
35. Kumar KA, Oliveira GA, Edelman R, Nardin EH, Nussenzweig V. 2004. Quantitative *Plasmodium* sporozoite neutralization assay (TSNA). *J. Immunol. Methods* 292:157–164.
36. Palma C, Overstreet MG, Guedon JM, Hoiczky E, Ward C, Karen KA, Zavala F, Ketner G. 2011. Adenovirus particles that display the *Plasmodium falciparum* circumsporozoite protein NANP repeat induce sporozoite-neutralizing antibodies in mice. *Vaccine* 29:1683–1689.
37. Kastenmüller K, Espinosa DA, Trager L, Stoyanov C, Salazar AM, Pokalwar S, Singh S, Dutta S, Ockenhouse CF, Zavala F, Seder RA. 2013. Full-length *Plasmodium falciparum* circumsporozoite protein administered with long-chain poly(I-C) or the Toll-like receptor 4 agonist glucopyranosyl lipid adjuvant-stable emulsion elicits potent antibody and CD4+ T cell immunity and protection in mice. *Infect. Immun.* 81:789–800.