

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Roman L. Hruska U.S. Meat Animal Research
Center

U.S. Department of Agriculture: Agricultural
Research Service, Lincoln, Nebraska

2016

Effect of Direct-Fed Microbial Dosage on the Fecal Concentrations of Enterohemorrhagic *Escherichia coli* in Feedlot Cattle

Brandon E. Luedtke
USDA-ARS, luedtkebe@unk.edu

Joseph M. Bosilevac
USDA Meat Animal Research Center, mick.bosilevac@ars.usda.gov

Dayna M. Harhay
USDA-ARS, dayna.harhay@ars.usda.gov

Terrance M. Arthur
USDA Meat Animal Research Center, terrance.arthur@ars.usda.gov

Follow this and additional works at: <http://digitalcommons.unl.edu/hruskareports>

Luedtke, Brandon E.; Bosilevac, Joseph M.; Harhay, Dayna M.; and Arthur, Terrance M., "Effect of Direct-Fed Microbial Dosage on the Fecal Concentrations of Enterohemorrhagic *Escherichia coli* in Feedlot Cattle" (2016). Roman L. Hruska U.S. Meat Animal Research Center. 409.

<http://digitalcommons.unl.edu/hruskareports/409>

This Article is brought to you for free and open access by the U.S. Department of Agriculture: Agricultural Research Service, Lincoln, Nebraska at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Roman L. Hruska U.S. Meat Animal Research Center by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Effect of Direct-Fed Microbial Dosage on the Fecal Concentrations of Enterohemorrhagic *Escherichia coli* in Feedlot Cattle

Brandon E. Luedtke,^{1,2} Joseph M. Bosilevac,¹ Dayna M. Harhay,¹ and Terrance M. Arthur¹

Abstract

Contamination of beef products by Shiga toxin-producing *Escherichia coli* is a concern for food safety with a particular subset, the enterohemorrhagic *E. coli* (EHEC), being the most relevant to human disease. To mitigate food safety risks, preharvest intervention strategies have been implemented with the aim to reduce EHEC in cattle. One class of interventions that has been widely used in feedlots is direct-fed microbials (DFMs), which can contain various dosing rates of probiotic bacteria. Here we compare the use of two different doses of a commercially available DFM on total EHEC load in a commercial feedlot setting. The DFMs used were the standard 10^9 *Propionibacterium freudenreichii* and 10^6 *Lactobacillus acidophilus* colony forming units (CFUs)/head/day dose of Bovamine[®] (Nutrition Physiology Company, Guymon, OK) and the higher dose, Bovamine Defend[™] (Nutrition Physiology Company), which is dosed at 10^9 *P. freudenreichii* and 10^9 *Lactobacillus acidophilus* CFUs/head/day. To analyze the total EHEC fecal concentration, 2200 head of cattle were assigned a DFM feed regimen lasting approximately 5 months. At harvest, 480 head of cattle were sampled using rectoanal mucosal swabs. A quantitative polymerase chain reaction assay targeting *ecfI* was used to enumerate the total EHEC fecal concentration for 240 head fed the low-dose DFM and 240 head fed the high-dose DFM. No significant difference ($p > 0.05$) in the fecal concentration of total EHEC was observed between the two doses. This suggests that using an increased dosage provides no additional reduction in the total EHEC fecal concentration of feedlot cattle compared to the standard dosage.

Introduction

THERE ARE ESTIMATED to be hundreds of serotypes of Shiga toxin-producing *Escherichia coli* (STEC) associated with human disease. However, those of the greatest concern for food safety are the enterohemorrhagic *E. coli* (EHEC) that cause the most severe forms of disease. While the majority of EHEC disease is caused by *E. coli* O157:H7, six additional serogroups (O26, O45, O103, O111, O121, and O145) are collectively referred to as the “top 6” and are responsible for 70–80% of non-O157 EHEC-disease. Thus, there remain ~20% of human non-O157 EHEC disease cases caused by other EHEC serogroups (Brooks *et al.*, 2005). These serogroups potentially pose a food contaminant concern (Bosilevac and Koohmaraie, 2011). In addition, atypical enteropathogenic *E. coli* (aEPEC) of multiple serogroups, which can

transiently possess Shiga toxin, are an emerging threat to contaminant detection and clinical diagnosis (Trabulsi *et al.*, 2002; Bielaszewska *et al.*, 2007).

Current molecular-based EHEC screening methods target *E. coli* O157:H7 and the other top 6 serotypes (referred to as top 7, here forward) and detect a combination of the genes associated with serotype differentiation, intimin (*eae*), Shiga toxin variants (*stx1/2*) (Perelle *et al.*, 2007; Jacob *et al.*, 2012; Bibbal *et al.*, 2014), and a set of genes with a propensity to be associated with EHEC (Delannoy *et al.*, 2013). Using the current methodologies can result in false-negative or false-positive results when a polymicrobial sample, like feces, is analyzed (Verstraete *et al.*, 2014). This is due to background bacteria independently possessing a target gene(s) within the sample (Feng, 2014). However, the effacing gene-positive conserved fragment 1 (*ecfI*) has been shown to be specific to

¹U.S. Department of Agriculture*, Agricultural Research Service, Roman L. Hruska U.S. Meat Animal Research Center, Clay Center, Nebraska.

²Department of Biology, University of Nebraska at Kearney, Kearney, Nebraska.

*USDA is an equal opportunity provider and employer. Trade names are necessary to report factually on available data; however, the USDA neither guarantees nor warrants the standard of the product, and the use of the name by USDA implies no approval of the product to the exclusion of others that may also be suitable.

pathogenic *E. coli* jointly possessing *eae* and *stx1* and/or *stx2* (Boerlin *et al.*, 1998) and in some instances aEPEC (Livezey *et al.*, 2015). By targeting *ecf1*, the total EHEC load associated with a sample can be assessed (Luedtke *et al.*, 2014).

Cattle and other ruminants act as natural reservoirs of EHEC, transmitting the bacteria through fecal shedding. Super shedding cattle, which have been defined as releasing greater than 10⁷ EHEC colony forming units (CFUs)/g of feces, (Matthews *et al.*, 2006; Menrath *et al.*, 2010), pose a concern for spreading EHEC among a herd and during the animal harvesting process (Arthur *et al.*, 2010). The use of various preharvest intervention strategies has been implemented with the aim of controlling the transmission of EHEC (LeJeune and Wetzel, 2007). An intervention strategy that has been adopted among many feedlots is the use of probiotics termed direct-fed microbials (DFMs) that are composed of *Lactobacillus* strains and other genera (Wheeler *et al.*, 2014).

The use of the standard dosage DFM marketed as Bovamine[®], which is dosed at 10⁶ *Lactobacillus acidophilus* CFUs/head/day, has been suggested to decrease the fecal shedding of O157:H7 (Brashears *et al.*, 2003; McAllister *et al.*, 2011). An additional formulation of Bovamine marketed as Bovamine Defend[™], which has an increased dosage of 10⁹ *Lactobacillus acidophilus* CFUs/head/day, is also available to cattle producers. Previous laboratory studies suggest that DFM reduce *E. coli* O157:H7 in a dose-dependent manner (Younts-Dahl *et al.*, 2005). Little data exist, however, on the ability of a high-dose DFM to reduce the concentration of total EHEC in commercial feedlot cattle to an extent greater than the standard dose.

In this study, we sought to (1) compare the total EHEC load in feces from commercial feedlot cattle fed either a standard-dose or high-dose DFM and (2) provide an alternative method to analyze the efficacy of preharvest interventions in the mitigation of a broad range of EHEC serotypes.

Materials and Methods

Cattle and treatment assignment

Approximately 2200 steers were identified that arrived at a feedlot located in the Beef Industry Food Safety Council (BIFSCo) monitoring region 3 in March and early April of 2014 and scheduled for harvest in September. Incoming cattle from the same source were randomly split using a two-way sort at initial processing, resulting in paired pens across treatments. The majority of cattle came from BIFSCo regions 6 and 7 and were analyzed using this region classification.

Pens assigned the numbers 1, 2, 6, 9, and 10 were considered region 6 sourced cattle. Pens numbered 1, 6, 9, and 10 each contained cattle from a different supplier within that region, while pen number 2 had a mixture of cattle from one supplier in region 6 and one supplier in 7, but nevertheless was considered as a region 6 pen for this study. Pens assigned numbers 3, 4, 5, 7, and 8 were considered region 7 sourced cattle. Pens numbered 3 and 7 each contained cattle from a single different supplier within that region. Pens 4 and 8 contained cattle from two different sources within region 7 that were grouped before sorting, while pen number 5 had a mixture of cattle from one supplier in region 7 and one supplier in region 3 but was considered as region 7 pen for this study (Fig. 1). DFM treatments were assigned, such that

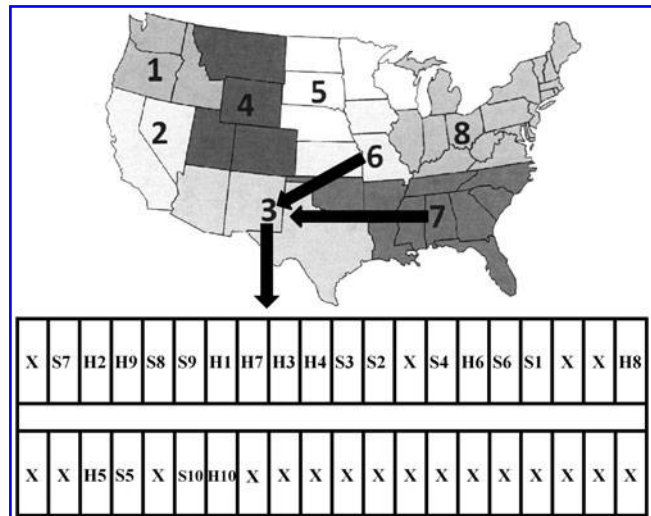


FIG. 1. Map of the cattle origins using the BIFSCo monitoring regions and pen identification. Two hundred forty head of cattle from region 6 and 240 head from region 7 were placed on a feedlot in region 3. Pens were grouped based on origin with 10 paired pens from region 6 and 10 paired pens from region 7. Samples were collected from 24 head per pen. Each paired pen was assigned a number and a feed regimen of either the standard-dose (S) or high-dose (H) direct-fed microbials (DFM), for example, S1 or H1. Each row consisted of 20 pens. X, Pens not part of study.

five paired pens of cattle from each region were fed the standard-dose DFM (Bovamine; Nutrition Physiology Company, Guymon, OK) and five paired pens of cattle from each region were fed the high-dose DFM (Bovamine Defend; Nutrition Physiology Company). Pens were as close to each other as possible to minimize differences due to location (moisture, flies, traffic, other cattle).

Cattle feeding and rations

The standard-dose DFMs (10⁹ *Propionibacterium freudenreichii* and 10⁶ *Lactobacillus acidophilus*) and high-dose DFMs (10⁹ *P. freudenreichii* and 10⁹ *Lactobacillus acidophilus*) were fed according to the manufacturer’s instructions through supplementation of the feedyard transition and finishing diets that included (dry matter basis) 31% flaked corn, 18% high-moisture corn, 12% corn gluten feed, 10% silage, 24% Sweet Bran, 3% tallow, and 2.0% vitamin/trace mineral mix.

Fecal swab sampling and DNA preparation

Fecal swabs were collected from 24 head per pen by swabbing the rectoanal junction of cattle at harvest in a commercial beef processing plant. Swabs were collected immediately following stunning and exsanguination. After collection, swabs were placed into 15-mL conical tubes containing 5 mL of tryptic soy broth (TSB, Difco; Becton Dickinson, Franklin Lakes, NJ) with phosphate buffer (TSB+PO4; 30 g of TSB, 2.31 g of KH2PO4, and 12.54 g of K2HPO4) per liter of solution and stored on ice for transport to the U.S. Meat Animal Research Center. Upon arrival at the laboratory, all tubes were vigorously mixed by vortex for 15 s and then a DNA preparation was made using a 20 µL aliquot

of each fecal suspension, which was added to a respective tube containing 180 μ L of the BAX[®] system lysis buffer supplemented with the BAX system protease (DuPont, Wilmington, DE). DNA from the samples was then prepared according to the manufacturer's instructions (DuPont) and stored at -20°C .

Quantitative polymerase chain reaction

The quantitative polymerase chain reaction (qPCR) was based on the detection of the *ecf-1* and *eae* genes, in which a 5-log standard curve was developed using the *E. coli* O157:H7 reference strain EDL 932 (ATCC 43894) as previously described (Luedtke *et al.*, 2014). The standard curve was run on each of the qPCR plates and the resulting Cq values for each dilution from across the plates were averaged to create a pooled standard curve for absolute enumeration as previously described (Sivaganesan *et al.*, 2010). The DNA preparations were thawed completely and briefly mixed by vortex before qPCR analysis. All samples, no template controls, and the standard curve were run in duplicate reactions on an ABI 7500 Fast Real Time PCR machine (Applied Biosystems[®] by Life Technologies, Carlsbad, CA) using the previously described duplex qPCR assay targeting *eae* and *ecf1* (Luedtke *et al.*, 2014).

Statistical analyses

All statistical analyses were performed using GraphPad Prism 6 (GraphPad Software, La Jolla, CA). The pen was considered the experimental unit. To determine significant differences between more than two groups of cattle, a one-way ANOVA and the Tukey's *post hoc* test were used, while a paired t-test was used to compare two groups of cattle. A $p < 0.05$ was considered significant for all tests.

Results and Discussion

The standard curve was reproducible for each dilution across the reaction plate (Table 1). Using the pooled standard curve approach, the curve was consistent and the qPCR efficiency for both *eae* and *ecf1* was 96% (Fig. 2). The no template controls were consistently negative for all reactions. These results are consistent with a previous report utilizing this assay to enumerate total EHEC directly from cattle feces obtained by rectoanal mucosal swab (RAMS) and provide an

TABLE 1. AVERAGE CQ VALUES AND THE CALCULATED COEFFICIENTS OF VARIATION FROM POOLED STANDARD CURVES RETURNED DURING THE ENUMERATION OF TOTAL EHEC FROM CATTLE FECES

\log_{10} CFUs/mL	<i>eae</i>		<i>ecf1</i>	
	Cq \pm SD ^a	CV (%)	Cq \pm SD	CV (%)
7.19	20.28 \pm 0.23	9.31	18.05 \pm 0.23	9.23
6.19	23.76 \pm 0.12	4.75	21.49 \pm 0.10	3.99
5.19	27.11 \pm 0.14	5.77	24.82 \pm 0.12	4.88
4.19	30.51 \pm 0.18	7.35	28.30 \pm 0.18	7.14
3.19	34.03 \pm 0.37	14.87	31.74 \pm 0.39	15.78

^aThe average Cq value \pm the standard deviation.

CFU, colony forming units; EHEC, enterohemorrhagic *Escherichia coli*; CV, coefficient of variation.

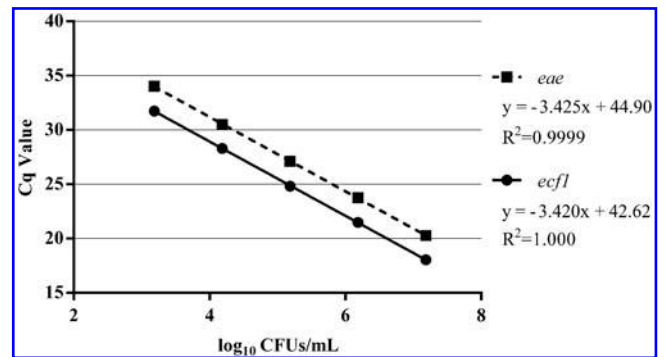


FIG. 2. Pooled standard curve used for the enumeration of the total enterohemorrhagic *Escherichia coli* (EHEC) fecal concentration. A 5 log standard curve was run in duplicate for each reaction plate and the Cq values for the targets *eae* and *ecf1* were pooled as described.

expected reliable limit of quantification of 1250 CFUs/mL (Luedtke *et al.*, 2014).

The use of RAMS and feces to evaluate the colonization and shedding level of EHEC by cattle has been documented in the literature (Rice *et al.*, 2003; Greenquist *et al.*, 2005; Niu *et al.*, 2008; Cernicchiaro *et al.*, 2011). While one study reported fair to moderate agreement (Kappa values of 0.36–0.45) between RAMS and feces results, with feces samples found more often positive for EHEC (Niu *et al.*, 2008).

Other studies found RAMS samples were more sensitive than fecal samples for determining the prevalence of *E. coli* O157 in feedlot cattle (Greenquist *et al.*, 2005) and concluded that RAMS more directly measured the relationship between cattle and EHEC, distinguishing between colonized cattle and those transiently shedding EHEC (Rice *et al.*, 2003). When differences in the reported impact of diet and probiotics on the shedding of *E. coli* O157:H7 were found using RAMS and feces (Cernicchiaro, 2010), subsequent studies later showed that the differences between the two methods were not statistically significant (Cernicchiaro, 2011). The use of RAMS, however, provides results as \log_{10} CFU/mL, and therefore, care must be taken when interpreting and comparing results with other results in the literature that may have directly tested feces and present units as \log_{10} CFU/g (Omisakin *et al.*, 2003).

As this study was performed in a commercial feedyard, animal handling was kept to a minimum to avoid production losses. While potentially providing additional information, repeated sampling over the course of the feeding would have required the animals to be handled more than what is typically done under commercial conditions and may have incurred production losses. To minimize pen-to-pen variation at the start of the study, animals from each incoming cattle lot were randomly sorted into the two treatments. The effects of DFM described in the literature have cumulative endpoint reductions in EHEC carriage and shedding (Brashears *et al.*, 2003; McAllister *et al.*, 2011); thus, our endpoint sample collection is a valid approach to evaluate the efficacy of the two probiotic dosages.

The concentration of total EHEC ranged from 0 to 10^7 CFUs/mL for cattle fed either the standard-dose or high-dose DFM. Fecal samples with total EHEC concentrations below

TABLE 2. DISTRIBUTION OF TOTAL EHEC ENUMERATED DIRECTLY FROM FECES OF CATTLE FED EITHER A STANDARD-DOSE OR HIGH-DOSE DFM

log_{10} CFUs/mL	Standard dose ^a		High dose ^a	
	Incidence ^b	Average ^c	Incidence ^b	Average ^c
7	1	7.23 ± 0.00	1	7.07 ± 0.00
6	7	6.41 ± 0.19	21	6.33 ± 0.26
5	35	5.42 ± 0.30	26	5.48 ± 0.28
4	44	4.47 ± 0.31	51	4.49 ± 0.29
3	79	3.48 ± 0.28	79	3.47 ± 0.30
2	44	2.52 ± 0.26	40	2.58 ± 0.21
1	24	1.68 ± 0.23	14	1.73 ± 0.20
0	6	N/A ^d	8	N/A

^an = 240 head of cattle.

^bNumber of cattle with total EHEC concentration at each order of magnitude.

^cAverage concentration of total EHEC ± the standard deviation within each order of magnitude.

^dN/A, Not applicable, the 0 log_{10} CFU/mL samples had no detectable amount of *ecfI* and could not be enumerated.

CFU, colony forming units; DFM, direct-fed microbials; EHEC, enterohemorrhagic *Escherichia coli*.

10³ CFUs/mL were enumerable based on the standard curve but likely were influenced by a stochastic effect (data not shown), and subsequently, all samples were grouped based on the whole log_{10} CFUs/mL value. The highest frequency of 79 (33%) head in the 10³ CFUs/mL range was observed for both the DFMs (Table 2). Cattle fed the standard dose had 74 (31%) head below 10³ CFUs/mL, while cattle fed the high dose had 60 (25%) head below this level (Table 2). When looking at potential super shedding cattle, the cattle fed the standard dose had 87 (36%) head with a total EHEC load at a concentration greater than or equal to 10⁴ CFUs/mL, while cattle fed the high dose had 99 (41%) head with a total EHEC load over the same range (Table 2). Since this qPCR assay does not differentiate between the different serotype concentrations that could be contained in a respective sample and the absence of research investigating if the total EHEC load influences downstream food contamination and human disease, labeling these cattle as super shedders is not definitive. However, the low infectious dose of 10–100 cells for O157:H7 and a speculated similar infectious dose for the non-O157:H7 (Pihkala *et al.*, 2012) serotypes warrant future research to determine if cattle with increased loads of total EHEC are a significant contributor to environmental transmission.

The average total EHEC from feces of cattle fed either a standard dose or high dose was 3.61 ± 1.38 and 3.80 ± 1.45 log_{10} CFUs/mL, respectively. No significant difference ($p > 0.05$) was observed for the average total EHEC concentration determined for cattle fed either DFM. While there was no difference in the fecal concentrations between the two DFM treatments, differences were observed between cattle lots that arrived from different origins. Total EHEC concentration based on the location of origin for each pen of cattle was analyzed, to determine if a regional difference existed. Only the region 7 lots fed the high dose were significantly different ($p < 0.05$) from the region 6 lots fed the standard dose, with the region 7 lots having the higher concentration of total EHEC. When the feed regimen was not taken into account, the region 6 lots had an average total EHEC concentration of 3.49 ± 1.47

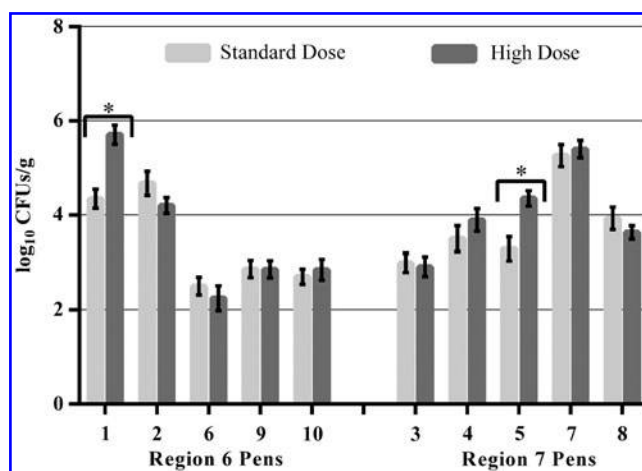


FIG. 3. The average total enterohemorrhagic *Escherichia coli* (EHEC) concentration from cattle fed either a standard-dose or high-dose direct-fed microbials (DFM) based on paired pens. From each pen, 24 head of cattle were sampled. The results represent the average total EHEC concentration from the 24 head sampled in each pen. The error bars indicate the standard error of the mean. *Indicates a significant difference ($p < 0.05$) in total EHEC concentration between feed regimens for the pens.

log_{10} CFUs/mL, while the region 7 lots had an average total EHEC concentration of 3.92 ± 1.33 log_{10} CFUs/mL. The difference in total EHEC concentrations between the two regions was significant ($p < 0.05$), however, the biological impact of these observed differences requires additional studies. At the pen level, significant differences ($p < 0.05$) in the total EHEC concentration were observed in paired pens numbered 1 and 5, with cattle fed the high dose having the higher concentration (Fig. 3). Across all 20 pens and the DFM used, multiple pens and DFM treatments were significantly different from each

TABLE 3. PREVALENCE OF CATTLE WITH TOTAL EHEC ABOVE 10³ CFUs/mL AND AVERAGE TOTAL EHEC CONCENTRATION OF PENS FED EITHER A STANDARD-DOSE OR HIGH-DOSE DFM

Assigned number	Standard dose ^a		High dose ^a	
	Head of cattle (%) ^b	Average ^c	Head of cattle (%) ^b	Average ^c
1	13 (54)	4.35 ± 0.98	23 (96)	5.71 ± 1.00
2	19 (79)	4.67 ± 1.25	17 (71)	4.21 ± 0.82
3	2 (8)	2.99 ± 0.99	2 (8)	2.91 ± 1.03
4	9 (38)	3.50 ± 1.34	10 (42)	3.90 ± 1.17
5	7 (29)	3.29 ± 1.24	15 (63)	4.35 ± 0.81
6	1 (4)	2.50 ± 0.92	2 (8)	2.25 ± 1.29
7	21 (88)	5.26 ± 1.14	22 (92)	5.40 ± 0.89
8	13 (54)	3.93 ± 1.15	5 (21)	3.63 ± 0.71
9	1 (4)	2.87 ± 0.90	1 (4)	2.86 ± 0.90
10	1 (4)	2.70 ± 0.80	2 (8)	2.85 ± 1.07

^an = 24 head of cattle per pen.

^bNumber of cattle with a total EHEC concentration above 10³ CFUs/mL.

^cAverage concentration of total EHEC ± the standard deviation for the entire pen.

CFU, colony forming units; DFM, direct-fed microbials; EHEC, enterohemorrhagic *Escherichia coli*.

other (date not shown). As expected, cohorts with a higher prevalence of cattle with a total EHEC concentration above 10^3 CFUs/mL also had a higher average total EHEC concentration for the pen regardless of the DFM regimen (Table 3). These pen-level observations agree with previous data for the transmission and fecal load of O157:H7 among a pen in relation to the presence of super shedding cattle (Omisakin *et al.*, 2003; Matthews *et al.*, 2006; Cobbold *et al.*, 2007; Arthur *et al.*, 2009).

This is the first study to compare the efficacy of a high-dose DFM to reduce the fecal concentration of total EHEC in beef cattle compared to a standard-dose DFM. Previous studies have focused on the ability of DFMs to reduce the fecal prevalence of O157:H7 (Brashears *et al.*, 2003; Peterson *et al.*, 2007) and in a dose-dependent manner (Younts-Dahl *et al.*, 2005), while other reports have shown that treatment of cattle with Bovamine does not significantly reduce the prevalence of O157:H7 (Cull *et al.*, 2012) or the top 6 STEC serotypes (Cernicchiaro *et al.*, 2014). To compensate for these varying results, it has been suggested that different concentrations of DFMs and the probiotic species composition have varying effects on foodborne pathogens and are application dependent (Callaway *et al.*, 2013). In addition, measuring the reduction in EHEC prevalence may be misleading in terms of value to food safety (Callaway *et al.*, 2013) as the fecal concentration has a greater impact on carcass contamination (Arthur *et al.*, 2009).

Conclusion

The results of this study indicate that the use of the DFM dosed at 10^9 *Lactobacillus acidophilus* CFU/head/day does not provide a significant reduction in the fecal concentration of total EHEC over the use of the DFM dosed at 10^6 *Lactobacillus acidophilus* CFU/head/day. Also, this study highlights the utility of the described assay, which is capable of enumerating and detecting nontop 7 EHEC in addition to the top 7, thus providing a more complete picture of the total EHEC concentration in cattle feces. Use of this assay could facilitate the evaluation of other preharvest interventions and provide information relevant to food safety.

Acknowledgments

The authors thank the cooperating feedlots and processing plant for access to sample collection. Additional recognition goes to Frank Reno, Lawnie Luedtke, and Greg Smith for technical support and Jody Gallagher for secretarial assistance.

Disclosure Statement

No competing financial interests exist.

References

- Arthur TM, Brichta-Harhay DM, Bosilevac JM, Kalchayanand N, Shackelford SD, Wheeler TL, Koohmaraie M. Super shedding of *Escherichia coli* O157:H7 by cattle and the impact on beef carcass contamination. *Meat Sci* 2010;86:32–37.
- Arthur TM, Keen JE, Bosilevac JM, Brichta-Harhay DM, Kalchayanand N, Shackelford SD, Wheeler TL, Nou X, Koohmaraie M. Longitudinal study of *Escherichia coli* O157:H7 in a beef cattle feedlot and role of high-level shedders in hide contamination. *Appl Environ Microbiol* 2009;75:6515–6523.
- Bibbal D, Loukiadis E, Kerouredan M, Peytavin De Garam C, Ferre F, Cartier P, Gay E, Oswald E, Auvray F, Brugere H. Intimin gene (*eae*) subtype-based real-time PCR strategy for specific detection of Shiga toxin-producing *Escherichia coli* serotypes O157:H7, O26:H11, O103:H2, O111:H8, and O145:H28 in cattle feces. *Appl Environ Microbiol* 2014;80:1177–1184.
- Bielaszewska M, Prager R, Kock R, Mellmann A, Zhang W, Tschape H, Tarr PI, Karch H. Shiga toxin gene loss and transfer in vitro and in vivo during enterohemorrhagic *Escherichia coli* O26 infection in humans. *Appl Environ Microbiol* 2007;73:3144–3150.
- Boerlin P, Chen S, Colbourne JK, Johnson R, De Grandis S, Gyles C. Evolution of enterohemorrhagic *Escherichia coli* hemolysin plasmids and the locus for enterocyte effacement in shiga toxin-producing *E. coli*. *Infect Immun* 1998;66:2553–2561.
- Bosilevac JM, Koohmaraie M. Prevalence and characterization of non-O157 shiga toxin-producing *Escherichia coli* isolates from commercial ground beef in the United States. *Appl Environ Microbiol* 2011;77:2103–2112.
- Brashears MM, Galyean ML, Loneragan GH, Mann JE, Killinger-Mann K. Prevalence of *Escherichia coli* O157:H7 and performance by beef feedlot cattle given *Lactobacillus* direct-fed microbials. *J Food Prot* 2003;66:748–754.
- Brooks JT, Sowers EG, Wells JG, Greene KD, Griffin PM, Hoekstra RM, Strockbine NA. Non-O157 Shiga toxin-producing *Escherichia coli* infections in the United States, 1983–2002. *J Infect Dis* 2005;192:1422–1429.
- Callaway TR, Edrington TS, Loneragan GH, Carr MA, Nisbet DJ. Current and near-market intervention strategies for reducing Shiga toxin producing *Escherichia coli* (STEC) shedding in cattle. *Agric Food Anal Bacteriol* 2013;3:103–120.
- Cernicchiaro N, Pearl DL, McEwen SA, LeJeune JT. Assessment of diagnostic tools for identifying cattle shedding and super-shedding *Escherichia coli* O157:H7 in a longitudinal study of naturally infected feedlot steers in Ohio. *Foodborne Pathog Dis* 2011;8:239–248.
- Cernicchiaro N, Pearl DL, McEwen SA, Zerby HN, Fluharty FL, Loerch SC, Kauffman MD, Bard JL, LeJeune JT. A randomized controlled trial to assess the impact of dietary energy sources, feed supplements, and the presence of super-shedders on the detection of *Escherichia coli* O157:H7 in feedlot cattle using different diagnostic procedures. *Foodborne Pathog Dis* 2010;7:1071–1081.
- Cernicchiaro N, Renter DG, Cull CA, Paddock ZD, Shi X, Nagaraja TG. Fecal shedding of non-O157 serogroups of Shiga toxin-producing *Escherichia coli* in feedlot cattle vaccinated with an *Escherichia coli* O157:H7 SRP vaccine or fed a *Lactobacillus*-based direct-fed microbial. *J Food Prot* 2014;77:732–737.
- Cobbold RN, Hancock DD, Rice DH, Berg J, Stilborn R, Hovde CJ, Besser TE. Rectoanal junction colonization of feedlot cattle by *Escherichia coli* O157:H7 and its association with supershedders and excretion dynamics. *Appl Environ Microbiol* 2007;73:1563–1568.
- Cull CA, Paddock ZD, Nagaraja TG, Bello NM, Babcock AH, Renter DG. Efficacy of a vaccine and a direct-fed microbial against fecal shedding of *Escherichia coli* O157:H7 in a randomized pen-level field trial of commercial feedlot cattle. *Vaccine* 2012;30:6210–6215.
- Delannoy S, Beutin L, Fach P. Discrimination of enterohemorrhagic *Escherichia coli* (EHEC) from non-EHEC

- strains based on detection of various combinations of type III effector genes. *J Clin Microbiol* 2013;51:3257–3262.
- Feng P. Shiga toxin-producing *Escherichia coli* (STEC) in fresh produce—a food safety dilemma. *Microbiol Spectr* 2014;2:EHEC-0010-2013.
- Greenquist MA, Drouillard JS, Sargeant JM, Depenbusch BE, Shi X, Lechtenberg KF, Nagaraja TG. Comparison of rectoanal mucosal swab cultures and fecal cultures for determining prevalence of *Escherichia coli* O157:H7 in feedlot cattle. *Appl Environ Microbiol* 2005;71:6431–6433.
- Jacob ME, Shi X, An B, Nagaraja TG, Bai J. Evaluation of a multiplex real-time polymerase chain reaction for the quantification of *Escherichia coli* O157 in cattle feces. *Foodborne Pathog Dis* 2012;9:79–85.
- Lejeune JT, Wetzel AN. Preharvest control of *Escherichia coli* O157 in cattle. *J Anim Sci* 2007;85:E73–E80.
- Livezey KW, Groschel B, Becker MM. Use of the *ecfI* gene to detect Shiga toxin-producing *Escherichia coli* in beef samples. *J Food Prot* 2015;78:675–684.
- Luedtke BE, Bono JL, Bosilevac JM. Evaluation of real time PCR assays for the detection and enumeration of enterohemorrhagic *Escherichia coli* directly from cattle feces. *J Microbiol Methods* 2014;105C:72–79.
- Matthews L, Low JC, Gally DL, Pearce MC, Mellor DJ, Heesterbeek JA, Chase-Topping M, Naylor SW, Shaw DJ, Reid SW, Gunn GJ, Woolhouse ME. Heterogeneous shedding of *Escherichia coli* O157 in cattle and its implications for control. *Proc Natl Acad Sci U S A* 2006;103:547–552.
- McAllister TA, Beauchemin KA, Alazzeh AY, Baah J, Teather RM, Stanford K. Review: The use of direct fed microbials to mitigate pathogens and enhance production in cattle. *Can J Anim Sci* 2011;91:193–211.
- Menrath A, Wieler LH, Heidemanns K, Semmler T, Fruth A, Kemper N. Shiga toxin producing *Escherichia coli*: Identification of non-O157:H7-super-shedding cows and related risk factors. *Gut Pathog* 2010;2:7.
- Niu YD, Xu Y, McAllister TA, Rozema EA, Stephens TP, Bach SJ, Johnson RP, Stanford K. Comparison of fecal versus rectoanal mucosal swab sampling for detecting *Escherichia coli* O157:H7 in experimentally inoculated cattle used in assessing bacteriophage as a mitigation strategy. *J Food Prot* 2008;71:691–698.
- Omisakin F, Macrae M, Ogden ID, Strachan NJ. Concentration and prevalence of *Escherichia coli* O157 in cattle feces at slaughter. *Appl Environ Microbiol* 2003;69:2444–2447.
- Perelle S, Dilasser F, Grout J, Fach P. Screening food raw materials for the presence of the world's most frequent clinical cases of Shiga toxin-encoding *Escherichia coli* O26, O103, O111, O145 and O157. *Int J Food Microbiol* 2007;113:284–288.
- Peterson RE, Klopfenstein TJ, Erickson GE, Folmer J, Hinkley S, Moxley RA, Smith DR. Effect of *Lactobacillus acidophilus* strain NP51 on *Escherichia coli* O157:H7 fecal shedding and finishing performance in beef feedlot cattle. *J Food Prot* 2007;70:287–291.
- Pihkala N, Bauer N, Eblen D, Evans P, Johnson R, Webb J, Williams C. Risk Profile for Pathogenic Non-O157 Shiga Toxin-Producing *Escherichia coli* (non-O157 STEC). Washington, DC: Office of Public Health Science, Office of Policy and Program Development, Food Safety and Inspection Service & United States Department of Agriculture, 2012.
- Rice DH, Sheng HQ, Wynia SA, Hovde CJ. Rectoanal mucosal swab culture is more sensitive than fecal culture and distinguishes *Escherichia coli* O157:H7-colonized cattle and those transiently shedding the same organism. *J Clin Microbiol* 2003;41:4924–4929.
- Sivaganesan M, Haugland RA, Chern EC, Shanks OC. Improved strategies and optimization of calibration models for real-time PCR absolute quantification. *Water Res* 2010;44:4726–4735.
- Trabulsi LR, Keller R, Tardelli Gomes TA. Typical and atypical enteropathogenic *Escherichia coli*. *Emerg Infect Dis* 2002;8:508–513.
- Verstraete K, Van Coillie E, Werbrouck H, Van Weyenberg S, Herman L, Del-Favero J, De Rijk P, De Zutter L, Joris MA, Heyndrickx M, De Reu K. A qPCR assay to detect and quantify Shiga toxin-producing *E. coli* (STEC) in cattle and on farms: A potential predictive tool for STEC culture-positive farms. *Toxins (Basel)* 2014;6:1201–1221.
- Wheeler TL, Kalchayanand N, Bosilevac JM. Pre- and post-harvest interventions to reduce pathogen contamination in the U.S. beef industry. *Meat Sci* 2014;98:372–382.
- Younts-Dahl SM, Osborn GD, Galyean ML, Rivera JD, Lonergan GH, Brashears MM. Reduction of *Escherichia coli* O157 in finishing beef cattle by various doses of *Lactobacillus acidophilus* in direct-fed microbials. *J Food Prot* 2005;68:6–10.

Address correspondence to:

Terrance M. Arthur, PhD

U.S. Department of Agriculture

Agricultural Research Service

Roman L. Hruska U.S. Meat Animal Research Center

State Spur 18D

Clay Center, NE 68933-0166

E-mail: terrance.arthur@ars.usda.gov