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Design, Biological Evaluation, and Computer-Aided Analysis of Dihydrothiazepines as Selective Antichlamydial Agents

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Author Contributions

M.C.-S. and S.P.O. conceived the idea. L.J.d.C. synthesized the compounds, performed antichlamydial assays, antibacterial assays, cytotoxicity, stability, and *in silico* studies. M.A.S. synthesized the compounds. J.F. performed antichlamydial assays. K.M.P.d.O. and J.V.d.A.d.S. performed antifungal assays and Ames test. J.B.C., S.H., and L.J.d.C. performed anaerobic experiments. D.J.F. and S.K. performed ClpX anti-ATPase assays. L.J.d.C. wrote the manuscript, all authors reviewed it and approved its content.

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ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jmedchem.2c01894>.

Detailing of the antichlamydial potency in different unit measures; detailing of the prediction accuracy of KPLS-radial 2D QSAR Model (radial 13); experimental *versus* KPLS-radial 2D QSAR model (radial 13) predicted values of antichlamydial activity (pIC₅₀); color maps for the 2D fingerprint (radial 13) QSAR model for all of the molecules used to generate the QSAR model; dose–response curves for the effects of **5b** and **5c** reported as percentage of recovered IFU; viability analysis for molecules **11d** and **11e** against the HEP-2 cell line; and spectral information of the studied molecules (PDF) Calculated *in silico* permeability parameters (CSV) Molecular formula strings and biological data (CSV)

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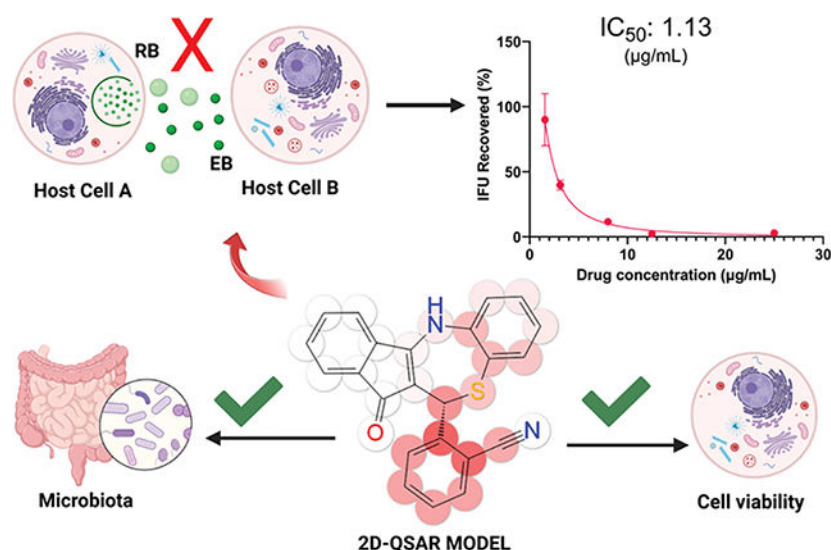
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Abstract

Chlamydia trachomatis (CT) causes the most prevalent sexually transmitted bacterial disease in the United States. The lack of drug selectivity is one of the main challenges of the current antichlamydial pharmacotherapy. The metabolic needs of CT are controlled, among others, by cylindrical proteases and their chaperones (*e.g.*, ClpX). It has been shown that dihydrothiazepines can disrupt CT-ClpXP. Based on this precedent, we synthesized a dihydrothiazepine library and characterized its antichlamydial activity using a modified semi-high-throughput screening assay. Then, we demonstrated their ability to inhibit ClpX ATPase activity *in vitro*, supporting ClpX as a target. Further, our lead compound displayed a promising selectivity profile against CT, acceptable cytotoxicity, no mutagenic potential, and good *in vitro* stability. A two-dimensional quantitative structure–activity relationship (2D QSAR) model was generated as a support tool in the identification of more potent antichlamydial molecules. This study suggests dihydrothiazepines are a promising starting point for the development of new and selective antichlamydial drugs.

Graphical Abstract



INTRODUCTION

Chlamydia trachomatis (CT) is an obligate intracellular Gram-negative bacterium¹ and the cause of the most prevalent sexually transmitted bacterial disease in the United States.² More than 1.8 million cases were reported in 2019, representing a 19% increase since 2015.³ The medical cost of CT infection treatments in the U.S. is estimated to be ~\$700 million.⁴ CT transmission occurs almost exclusively through sexual contact, although there are reported cases of infections during labor, which can lead to newborn conjunctivitis.⁵ CT infection is especially problematic in reproductive-aged women because it can result in pelvic inflammatory disease (PID), tubal factor infertility, ectopic pregnancy, and chronic pelvic pain.⁶ CT can result in trachoma⁷ (the leading cause of infectious blindness worldwide),⁸ facilitate human immunodeficiency virus (HIV) infection,⁹ and is associated with preterm labor, low birth weight, and perinatal mortality.¹⁰ Further, the reported infection rate for chlamydia of African Americans/Blacks is six times that of Whites.³

CT displays a biphasic developmental cycle.¹¹ The cycle begins when the elementary body (EB), the infectious but nondividing form of the bacterium, interacts with membranes of host cells, leading to its uptake.¹² After entering the host cell, the EB remains within a host-derived vesicle, known as an inclusion, in which the EB differentiates into the reticulate body (RB) form.¹¹ RBs are the replicative but noninfectious form of CT that, after repeated cycles of bacterial cell division, undergo secondary differentiation to EBs. Later, EBs are released by lysis or inclusion extrusion and a new infective cycle ensues.¹³

Broad-spectrum antibiotics such as azithromycin (AZM) and doxycycline (Doxy) are the drugs of choice to eradicate this infection.¹⁴ However, accumulating evidence suggests that AZM presents failure rates; *i.e.*, 16% for the treatment of urethritis¹⁵ and 14% for female urogenital chlamydial infections.¹⁶ These levels are higher than the desired World Health Organization's target chlamydia treatment failure rate of <5%.¹⁷ Further, broad-spectrum drugs affect commensal flora,¹⁸ altering their normal function¹⁹ and both AZM and Doxy may compromise the microbiome.^{20,21} Of note, studies have shown that gut

microbial dysbiosis can contribute to the development of diabetes,²² obesity,²³ asthma,²⁴ and other conditions.²⁵ Further, these drugs can foster the development of general antibiotic resistance.^{26,27} For example, treatment of CT with broad-spectrum antibiotics, including AZM, can lead to nasopharyngeal carriage of macrolide-resistant *Staphylococcus aureus*,²⁸ macrolide-resistant gonorrhea,²⁹ and *Mycoplasma genitalium*.³⁰ Additionally, treatment of trachoma with AZM has led to induction of resistance in *Streptococcus pneumoniae*.^{28,31} Thus, narrow-spectrum/selective drugs can prevent damage or alterations in the patient's microbiome³² and limit the development of general resistance. Unfortunately, there are few reports of molecules that are specific for *Chlamydia*, and none of those molecules has received U.S. Food and Drug Administration (FDA) approval.^{26,33–36} In this context, novel scaffolds or targets are needed to advance antibacterial research.

Proteolytic enzymes can play an important role as new drug targets.³⁷ Affecting the degradation machinery is an attractive, yet barely explored, strategy to control pathogens.³⁸ The ClpXP protease system is well conserved in bacteria and relevant for protein turnover.³⁹ This degradation complex is composed of a serine protease subunit, known as ClpP, and an unfoldase chaperone, *e.g.*, ClpX (which has ATPase activity). The chaperone component recognizes, unfolds, and transfers target proteins to ClpP for degradation.⁴⁰ The degraded proteins include damaged or nonfunctional proteins as well as transcriptional regulators and other specific targets.⁴¹ The ClpP protease component of the ClpXP complex has been more extensively studied as a drug target than the unfoldases.⁴² Some of the structures that target the ClpP are (i) acyldepsipeptide (ADEP) derivatives;⁴³ (ii) activators of self-compartmentalizing proteases (ACP);⁴⁴ (iii) sclerotiamide;⁴⁵ (iv) β -lactones;⁴⁶ (v) phenyl esters;⁴⁷ or (vi) bortezomib⁴⁸ (Figure 1). However, only a few reports describe small molecules targeting the ClpX ATPase.⁴⁹ The dihydrothiazepine scaffold reported by Fetzer and co-workers⁵⁰ is the first reversible inhibitor of ClpX. Their most potent dihydrothiazepine derivative (**334**—Figure 1) disrupts the hexameric state of Methicillin-resistant *S. aureus* (MRSA) ClpX and blocks its ATPase activity with an IC₅₀ value of 0.8 μ M. Additionally, according to the authors, the whole ClpXP proteolytic complex collapses in response to dihydrothiazepine derivative action. Recently, it has been demonstrated that the ClpXP proteolytic system is critical to CT development^{51–53} and that **334** (Figure 1) can disrupt chlamydial ClpX function affecting *Chlamydia* growth in cell culture.³⁹ Thus, targeting the ClpX component of the CT proteolytic system with dihydrothiazepine derivatives could produce anti-CT agents.

In the present work, we report the synthesis and biological evaluation of dihydrothiazepines with antichlamydial activity. Some of the synthesized derivatives showed substantial reduction in chlamydial inclusion number and size in infected HEp-2 cells. To investigate the selectivity of our most promising structure, we tested its antimicrobial activity against clinically relevant Gram-positive/Gram-negative bacteria, yeast, and bacteria from the human gastrointestinal (GI) and genital tract microbiota. Inhibition of ClpX ATPase activity was assessed to acquire initial insights on the mechanism of action responsible for chlamydial eradication. Further, we tested the toxicity of our most promising analogues toward human cells, their stability in serum and simulated gastric and intestinal fluids. Moreover, the potential mutagenicity was investigated using an Ames test. To understand

the activity and develop more potent structures, we generated a two-dimensional (2D) quantitative structure–activity relationship (QSAR) model. We used this model to guide the design of compounds with superior potency than the parent molecules. Our results indicate that our best dihydrothiazepine derivative is selective for CT and represents a starting point for the development of new antichlamydial drugs.

RESULTS AND DISCUSSION

Chemistry.

First, we explored modifications on the ortho, meta, and para positions of ring A. The synthesis consisted of Knoevenagel condensation of indan-1,3-dione (**1**) with the corresponding benzaldehyde (**2**) following the method of Fetzer et al.⁵⁵ Then, dihydrothiazepines (**5a–y**) were obtained by the cyclization of 2-benzylideneindan-1,3-diones (**3a–y**) with 2-aminothiophenol (**4**) (Scheme 1).

To place an amine on ring A, we performed catalytic hydrogenation of the nitro derivative **5n** (Scheme 2). To investigate the importance of the thioether on the seven-member ring heterocycle, we oxidized this group to a sulfone with *meta*-chloroperoxybenzoic acid (*m*CPBA).

To determine the importance of ring B on the antichlamydial activity, we removed it by reacting selected 2-benzylideneindan-1,3-dione intermediates (**3**) with 2-amino-ethanethiol (**8**) using the same reaction conditions (Scheme 3).

Since ring B proved to be key for activity, we explored how modifications on this ring affect the activity. First, we placed a trifluoromethyl group on the meta position. Further, we replaced the thioether by a secondary amine on the seven-member ring heterocycle (diazepine derivatives). The synthetic route for these modifications is presented in Scheme 4.

Biological Evaluation.

Determination of Antichlamydial Activity.—We tested our compounds against *C. trachomatis* serovar L2. As mentioned, CT has a biphasic developmental cycle.⁵⁶ Once the EB is internalized into the cell, it resides in it as an inclusion where it differentiates into the replicative RB form.⁵⁷ Since the inclusions are linked to the growth and replication of the pathogen,⁵⁸ we quantitatively determined the antichlamydial activity by analyzing the number of inclusions after treatment and comparing them to untreated (UTD) samples and drug controls. To do so, we adapted an immunofluorescence assay (IFA) described previously^{36,52,59} to develop a semi-high-throughput screening platform (see full description in the Experimental Procedures section). This allowed us to determine the IC₅₀ (the concentration to induce 50% of inclusion inhibition) of all analogues using a screening approach that is faster than other methods described in the literature.^{52,60–62}

We observed that substitutions at ortho and meta positions on ring A had a positive effect on the antichlamydial activity (Table 1). These positions allow substituents with diverse electronic characteristic (both electron-withdrawing and electron-donating groups maintained the activity). In accordance, our top five most active compounds (**5a–e**)

presented substitutions at one or both of these positions in ring A. On the contrary, groups in the para position caused a reduction in the IC₅₀ (Table 1). In particular, bulky groups at the para position had the most significant drop in potency (a benzyloxy group (**5s**) led to a 14-fold decrease in the IC₅₀ and a phenyl (**5v**) group completely abolished the activity). Of note, derivatives displaying a thiophene instead of a benzene in ring A (**11a** and **5y**, Table 4) retained activity.

Further, we observed that acidic groups abolished the antichlamydial activity. This can be seen with derivatives **5t** and **5u**, which present carboxylic acid in ring A at ortho and para positions, respectively. This observation can be partially explained by the ionization of these two molecules at the pH used in the assays (~7.4) (calculated pK_a: 4.10 and 3.48 for **5t** and **5u**, respectively), which can diminish their permeability into the host cell preventing them from displaying antichlamydial activity. This theory is supported by the fact that the esterified version of **5t**, compound **5j**, displays antichlamydial activity (Table 1), supporting the notion of permeability and activity. However, the polar derivatives **5f** and **5n**, with a nitro group at meta and para positions, respectively, possess antichlamydial activity (IC₅₀ of 6.35 and 9.01 µg/mL).

To analyze these results, we also calculated *in silico* permeability parameters (values for all of the compounds can be found in File S1; Table 2 shows selected compounds). Among the evaluated parameters, two descriptors indicated the derivatives having a nitro group should have better permeability than the carboxylic acids: (i) the predicted apparent Caco-2 cell permeability (QPPCaco) and (ii) the predicted apparent Madin–Darby Canine Kidney (MDCK) cell permeability (QPPMDC). First, QPPCaco values for the acid derivatives are lower than the ones predicted for the nitro analogues (101.98 and 345.75 nm/s for **5t** and **5u** against 429.39 and 605.69 nm/s for **5n** and **5f**, respectively). The same pattern is seen on the second permeability prediction: 82.60 and 298.59 for **5t** and **5u** against 307.34 and 444.67 for **5n** and **5f**, respectively. Thus, the predicted permeability for the nitro derivatives is considerably better than that for the carboxylic acid analogues (the ester analogue, **5j**, also has higher predicted permeability than the acid—Table 2). Meanwhile, the predicted octanol/water partition coefficient (QPlogPo/w (pH 7.4)) and solvent accessible surface area (SASA) were similar between the analogues. Of course, electronic effects might also contribute to the distinct antichlamydial activities. Resonance forms, position of the substituents, and electronegativity cannot be discarded.⁶³

Further, the removal of ring B led to a significant drop in the IC₅₀ values (Table 3) when compared to the molecules displaying the same substitution pattern but presenting such a ring (for derivative **9d**, due to no inhibition seen at the concentration range tested, we could not calculate the IC₅₀). Structural studies need to be done to assess if there is a π -stacking interaction that is critical for activity or if this is related to cell permeability/lipophilicity.

Interestingly, the introduction of a trifluoromethyl at the meta position on ring B (Table 4) led to a slight increase in the potency of all of the derivatives (when compared to analogues possessing the same substitution on the rest of the structure—Table 1). For instance, the IC₅₀ of compound **5q** improved from 14.69 to 12.30 µg/mL (**11c**), compound **5y** from 7.36 to 6.53 µg/mL (**11a**), and compound **5l** from 7.50 to 6.93 µg/mL (**11b**).

A potential explanation for the slight improvement in the potency of **11a–c** may be a result of increased permeability into host cells. The *in silico* permeability parameters (Table 2) analyzed for these molecules support this hypothesis. Molecules lacking the trifluoromethyl (**5y**, **5l**, **5q**) group present lower values for the predicted QPPCaco and QPPMDCK cell permeabilities as well as lower predicted octanol/water partition coefficient values (QPlogPo/w—pH 7.4) when compared to analogues that display this functional group (see File S1 and Table 2).

Electronic effects might also contribute to the beneficial activity of the trifluoromethyl group at the meta position on ring B. Figure 2a displays the calculated quantum mechanics molecular electrostatic potential (MEP) for various analogues. The MEP surface suggests an electron density deficiency on ring B for the CF₃ analogues when compared to their unsubstituted counterparts. A potential explanation (assuming aromatic amino acids are nearby and keeping in mind the binding pocket of the ClpX is unknown at this moment) might be improved π -stacking with Tyr, Phe, or Trp in the binding pocket. In fact, the MEP for these amino acids shows the electron-rich nature (dark red color) of the aromatic ring (especially Trp and Try) side chains (Figure 2b). Thus, electronic effects complementary to the CF₃ substituted B-ring (Figure 2a) would be favored. We understand that the existence of such interactions depends also on other factors like the orientation of the interacting groups and distance. However, due to the absence of a crystal structure and the binding pocket identification, our current investigation tools are limited. Another beneficial interaction mediated by the CF₃ might be the establishment of hydrogen bonds between the fluorine atoms and residues in the binding pocket. Altogether, the data suggest that the B-ring is crucial for the antichlamydial activity and that both rings A and B are permissive to modifications while maintaining activity against CT.

Next, we assessed the SAR for the dihydrothiazepine seven-member ring. We noticed the exchange of sulfur for a nitrogen increased the IC₅₀ values, but it did not abolish activity (Table 4, compounds **13a–c**). Further, oxidation of the thioether to a sulfone also reduced the activity but still the derivative containing this modification (**7a**) showed intermediate activity (10 μ g/mL).

We found the permeability parameters could be reasonably correlated to better antichlamydial activity for specific comparison cases, for instance, carboxylic acids *versus* nitro structures, or molecules with CF₃ groups (described above), or compound **13a–c** analogues (S replaced by NH). Due to the intracellular nature of CT, it is expected that permeability will affect the activity (improved membrane permeability leads to higher accumulation of drug inside the cell⁶⁵). In fact, the most potent molecules (**5a–d**) present high-permeability-related parameters (Table 2) and not active compounds like **9d** display very low predicted permeability. However, not all highly lipophilic molecules presented good activity. For instance, low active molecule **5s** (22.76 μ g/mL) presents high predicted permeability (QPPCaco: 4104.20; QPPMDCK: 3532.70 nm/s). Further, the most active compound (**5a**) displays lower values for the predicted permeability (QPPCaco: 1794.20 and QPPMDCK: 1465.80) than the second or third most active structures (**5b**: 3707.80, 3212.50 and **5c**: 3639.20, 3188.80 for QPPCaco and QPPMDCK, respectively). Further, the acetyl ester (para position, **5j**) of the phenol (**5p**) is more active than the methyl

ether (**5q**) of the same molecule, although its permeability is lower. This may suggest electronic effects are as important as the permeability parameter (although the occurrence of intracellular ester hydrolysis may not be discarded since the similar acid derivative—**5u**, presenting a carboxylic acid at the meta position—displays ClpX anti-ATPase activity but no antichlamydial activity in the cellular assay). These illustrate the multifactorial nature of drug activity.⁶⁶ In accordance, small structural changes will affect the biological activity⁶⁷ (especially when comparing a cellular assay directly to an enzymatic experiment⁶⁸).

Fingerprint-Based Two-Dimensional (2D) QSAR Studies.—Although it is common to infer a SAR through the qualitative evaluation of a library of molecules, some intricacies of the relationships are better elucidated using a visual projection of the quantitative contributions of different chemical groups.⁶⁹ Accordingly, to analyze the key structural features of the dihydrothiazepine library, a 2D quantitative structure–activity relationship model (QSAR) was generated. This ligand-based tool is necessary for this study because there is no published chlamydial ClpX crystal structure, and the binding pocket has not been identified. Further, the use of a three-dimensional (3D) QSAR model (based on 3D descriptors) will be inadequate since the antichlamydial activity reported is based on racemic mixtures. Additionally, the biologically active conformation is unknown at the moment. Although the active conformation of the ligand is not absolutely indispensable for all 3D QSAR methods,⁷⁰ it increases the reliability of the models and is a requirement for most of the 3D QSAR approaches.⁷¹

To develop our model, we used 2D fingerprint descriptors and AutoQSAR, a machine-learning software that builds and applies QSAR models through automation.⁷² The program computes fingerprints and uses machine-learning statistical methods to create a predictive QSAR model that can describe the studied biological activity. The kernel-based partial least-square (KPLS) regression method using 2D fingerprint radial descriptors produced a statistically relevant QSAR model (Figures S1, 3, and 4). The best KPLS model generated using radial descriptors (model KPLS—radial 13) possesses good predictive accuracy in the training as well as in the test set (Table S3). The obtained model displays satisfactory statistical results as evaluated by the correlation coefficient (R^2 —a measure of the data explained by the model⁷³); the coefficient of determination between the observed and predicted biological activities for the test set (Q^2 —a measure of predictability⁷⁴); low standard deviation (SD) of the regression calculated based on the training set; and low root mean square error (RMSE) in the test set predictions (RMSE—expresses the variability not explained by the model⁷⁵). Further details regarding the predictive accuracy in the test and training set and statistical parameters/reference values for the QSAR model validation are presented in Tables S2 and S3 and Figure S1.

The used fingerprints descriptors allow the KPLS model to be decomposed into individually atomic effects, allowing a convenient way of visualizing favorable and unfavorable aspects of the chemical structures. The evaluation of the individual atomic contribution to the model is illustrated in Figure 3a using active and Figure 3b using molecules presenting low antichlamydial activity (color maps for the entire dataset can be found in Figure S2). Groups that contribute positively to activity are highlighted in red and the detrimental ones in blue. Color intensity reflects the strength of the effect. The 2D KPLS-QSAR model provides an

efficient approach to visualize molecules (Figures 3 and 4). The compounds in panel “a” are more active than the ones presented in panel “b”.

Other relevant features are highlighted by this QSAR model. For instance, for molecule **5s**, the darkest blue color is centered at the methylene group at the para position (ring A), indicating that this substitution pattern is highly unsuited for modifications (Figure 3b). The same effect is seen in **5v** (phenyl group at the para position—ring A), which does not present antichlamydial activity at the tested concentration. These observations could relate to the size and shape of the binding pocket, which probably does not accommodate bulky groups at the para position. Further, molecules **6a**, **9c**, and **13a** presenting smaller electron-donating groups at the para position of ring A (amine and methyl groups, respectively) are also detrimental for the activity.

The QSAR model indicates the favorable effect of lipophilic groups regardless of the position within the dihydrothiazepine core (Figure 4a). As mentioned above, substituents at the para position have a detrimental effect on the biological activity (ring A is highlighted in blue when such positions are substituted). It is possible that the presence of lipophilic and electronegative substituents (*e.g.*, trifluoromethyl or chloro at ring A) may counterbalance this unfavorable effect (note the red sphere around these groups in Figure 4a, molecules **5h** and **5k**). This could also be associated to the relationship between increasing lipophilicity and a superior ability to cross the membrane of the host cell and of the inclusions to access the target.

Another important feature is the unfavorable effect of hydrogen bond (H bond) donor groups at the A ring as shown in Figure 4b as demonstrated by the more intense spherical blue color at those groups (Figure 4b, molecules **5p**, **6a**, and **334**). Their effects are more evident when the group is placed at the para position *versus* the meta position (for **334**, the A ring displays more faded blue color than **5p** and **6a**). Interestingly, even though **334** possessed an inhibitory effect against ClpX of *S. aureus*,⁵⁵ it only showed moderate activity (IC₅₀ ~ 16 µg/mL) against *C. trachomatis*. This might be a result of differences in the composition of the binding pocket between ClpX orthologues in these different microorganisms (52.09% of protein similarity according to Uniprot databank alignment tool⁷⁶). Further, the differences in membrane constitution may influence permeability, and consequently, the biological activity of **334** across bacterial species. To be able to inhibit ClpX (or other targets) in CT, **334** needs to cross the host cell membrane, the inclusion membrane, and the double membrane of CT (a Gram-negative bacteria). On the opposite side, for *S. aureus*, a single membrane needs to be crossed by the drug. The impact of such differences might be clarified once the binding pocket of dihydrothiazepines are identified.

EB Progeny Assay.—To determine the ability of the compounds to inhibit infectious (EB) progeny production and to validate the IC₅₀ obtained by our semi-high-throughput IFA screening, we performed an inclusion forming units (IFU) reinfection assay⁷⁷ using the three most potent structures. Here, we quantified the number of *C. trachomatis* infectious forms (EB) from previously treated cultures at five concentrations (as well as untreated and AZM controls) following infection of a new cell monolayer. The chlamydial infectivity, reported as IFUs, was determined by counting the number of green-fluorescent inclusions

after immunofluorescent staining at 24 h post-secondary infection (Figure 5). In addition to the number of inclusions, we also evaluated the size and appearance of the chlamydial inclusions after treatment with the studied drugs. The most promising compound (**5a**) displayed inhibitory activity against CT with an IC_{50} of 1.13 $\mu\text{g/mL}$ (similar to the IC_{50} value determined in the semi-high-throughput screening—1.61 $\mu\text{g/mL}$, Figure 5a). The dose–response curve displays a concentration-dependent profile for the reduction in the chlamydial progeny, as seen by the reduction in the number of chlamydial inclusions (Figure 5a). Importantly, we noticed a remarkable reduction in the inclusion size compared to the control (UTD) even in lower doses, for instance, 3.12 and 1.56 $\mu\text{g/mL}$ (Figure 5c). Moreover, even at 3.12 $\mu\text{g/mL}$, compound **5a** achieved an inclusion recovery ~60% lower than the untreated samples (Figure 5c). When compared against the drug control AZM (Figure 5b), our tested structure displayed lower potency. However, the superior antichlamydial activity of AZM is also followed by a remarkable lower selectivity when tested against different bacteria, including different species from the human microbiota (Table 5). Moreover, AZM is a commercialized drug that had its potency optimized throughout its preclinical research phase, while the dihydrothiazepine scaffold has not been fully investigated. The obtained IC_{50} values for **5b** and **5c** in the progeny assay also display the same pattern as for the semi-high-throughput screening. The first molecule has superior activity (6.41 $\mu\text{g/mL}$) compared to the latter (7.13 $\mu\text{g/mL}$). The dose–response curves and immunofluorescence images for compounds **5b** and **5c** can be found in Figure S3.

Cytotoxicity.—Following the determination of the antibacterial activity of our library, we evaluated the *in vitro* cytotoxicity of the three most active molecules (**5a–c**) against epithelial type 2 (HEp-2, host cell in the antichlamydial assays), epithelial endometrium adenocarcinoma (HEC-1-A), and immortalized human embryonic kidney (HEK-293) cells. HEC-1-A and HEK-293 cells showed a very good viability profile, over 75% survival at the highest concentration for all tested compounds: 256 $\mu\text{g/mL}$ (Figure 6b,c). HEp-2 cells proved to be more sensitive to the tested molecules. At the highest tested concentration, around 50% cell viability was seen (Figure 6a). Importantly, in the highest concentration used in the antichlamydial assays (25 $\mu\text{g/mL}$), molecules **5a** and **5c** displayed high cell viability values (84 and 78%, respectively). Compound **5b** shows a slightly increased cytotoxicity against HEp-2 cells at this concentration, displaying a viability value of 73%. Compound **5a** was selected for further investigations since it presents higher potency (IC_{50} for the infectious progeny inhibition: 1.13 $\mu\text{g/mL}$ versus 6.41 and 7.13 for **5b** and **5c**, respectively—Figure S3) and a similar cytotoxicity profile when compared to the other derivatives analyzed.

Selectivity.—One major challenge of current antichlamydial treatments is their lack of selectivity. To determine the spectrum of antibacterial effect of **5a**, we tested it against a broad panel of microorganisms spanning from pathogenic bacteria and fungi to bacteria from the human microbiota (Table 5). Molecule **5a** proved to be inactive against Methicillin-resistant *S. aureus* USA 300 JE2, *P. aeruginosa* PA01, *A. baumannii* 2208, and *E. coli* K12 up to the highest concentration analyzed.

Additionally, **5a** also showed no antibacterial activity against *L. rhamnosus* (CGMCC 1.3724) and *L. plantarum* (DSM 20174). This result is encouraging since *L. rhamnosus* is found in the human gut and vaginal microbiota.⁷⁸ Further, both species are widely used as probiotic strains.^{78,79} To further determine the antibacterial activity against bacteria that comprise the human gut microbiota, we tested our compound against *B. producta*,⁸⁰ *C. ramosum*,⁸¹ *B. thetaiotaomicon*,⁸² *A. caccae*,⁸³ and *E. coli* MG1655. Compound **5a** was completely inactive against all species tested up to the highest concentration evaluated. On the other hand, AZM showed very potent antibacterial activity against most tested bacteria. In particular, for the gut microbiota tested strains, AZM displays MIC values as low as <0.06 µg/mL for *B. producta* DSM 2950 and *C. ramosum* DSM 1402, 0.5 µg/mL for *B. thetaiotaomicon* DSM 2079, and 2 µg/mL for *E. coli* MG1655. Regarding the lactobacillus strains, AZM also presented low MIC values that ranged around 2 to <1 µg/mL. The only gut microbiota bacteria that showed no susceptibility to AZM in the concentration range tested was *A. caccae* DSM 14662. The fact that compound **5a** did not affect the tested gut microbiota strains is very interesting, since, as pointed out, gut microbiota symbiosis is linked to the development of numerous diseases²⁵ and to increased rates of antibiotic resistance to other pathogens.

Molecule **5a** also proved to be inactive (MIC > 1000 µg/mL) against *C. albicans* (90028), *C. glabrata* (2001), *C. krusei* (6558), and *C. tropicalis* (750). The lack of activity against *Candida* is expected given that it is a eukaryotic organism. These species are commonly found in the gastrointestinal (GI) and genital tract microbiota of healthy humans. However, during host immunosuppression or alterations in the bacterial microbiota, these microorganisms can disseminate and cause life-threatening illness.^{84–87}

Overall, these results suggest that normal microbiota in humans might be unaffected by this class of compounds, which in turn suggests that the development of widespread resistance or alterations in the bacterial composition of the microbiota is unlikely. We hypothesize that the lack of activity against other classes of microorganisms might be a result of the unique cell wall structure of CT⁸⁸ when compared to other bacteria and/or the unique developmental cycle of Chlamydia.¹¹ In fact, the ClpXP complex⁵¹ is essential for CT development,^{39,51,53} whereas it is dispensable for other types of bacteria like *E. coli*.⁸⁹ These factors might play a role in the selectivity of this compound.

ClpX Anti-ATPase Activity.—To investigate the mechanism of action, we studied the inhibition of CT ClpX ATPase activity *in vitro*. We assessed the ability of **5a**, **5c**, **5u**, and **9d** to block ATP depletion over time using recombinant CT ClpX and a Kinase-Glo endpoint assay (Figure 7). All tested derivatives at 25 µg/mL significantly reduced Ct ClpX ATP consumption compared to the dimethyl sulfoxide (DMSO) solvent control by ~20%. While *in vitro* recombinant ClpX inhibition was expected for compounds **5a** and **5c** based on the antichlamydial assays, the *in vitro* activity of **5u** and **9d** contrasted with those experiments where no inhibition was observed. It is conceivable that membrane permeability negatively impacted *in vivo* activity for **5u** and **9d**. Compound **5u**, as discussed above, is ionized in the pH of the cell culture assay. This fact reinforces the highly influential effect of permeability in the activity of this scaffold and explains the difference in the cell culture assay *versus* the *in vitro* ClpX inhibition. Furthermore, the same rationale can

be applied to explain the *in vitro* ClpX inhibition of **9d**. This structure presents very low predicted permeability compared to other highly active molecules of our library according to Caco-2 cell permeability and MDCK cell permeability parameters (File S1). For instance, **5a** displays high potency in the cell culture assay and high-predicted-permeability Caco-2 cell permeability of 1794.16 nm/s and MDCK cell permeability of 1465.85 nm/s. On the contrary, **9d** displays 149.74 and 112.61 nm/s for the same parameters, respectively (File S1).

The activities of **5a** and **5c** against ClpX *in vitro* support that CT inhibition in the cell culture assays by these compounds is due, at least in part, to reduced ClpX activity. However, the results from the Sieber group, using an *S. aureus* ClpX knockout strain, indicate that the dihydrothiazepine scaffold could also have other targets within the bacterium.⁵⁵ Thus, a second target cannot be discarded.

Stability Studies.—To analyze the drug-like properties of **5a** to a greater extent, we evaluated its stability in human serum (Figure 8a), simulated gastric fluid (SGF, pH 1.40, Figure 8b), and simulated intestinal fluid (SIF, pH 7.60, Figure 8c). As shown in Figure 8, all of the tested compounds exhibited a satisfactory stability profile for the serum, SGF, and SIF assays during the study. For the stability in serum, at 180 min, the percentage of drug remaining in the solution was around 100%, and at 24 h showed a decrease to 80%. In SGF, the percentage of drug remaining at 180 min was around 90% with a decrease to 70% after 24 h of exposure. Furthermore, for SIF, the same stable pattern was visualized with the percentage of drug remaining around 85% at 24 h.

Ames Test.—Analysis of the mutagenic potential in drug discovery intends to rapidly recognize unpromising candidates, removing them from development.⁹⁰ Particularly, for antibacterial drugs, the evaluation of the potential mutagenic effect is highly significant since certain antibiotics induce elevated rates of mutation in bacteria.⁹¹ The elevated rate of mutagenesis expands the genetic diversity of bacterial populations, which might increase the rate at which bacteria develop antibiotic resistance.^{91,92} To further characterize the potential of **5a**, we evaluated its mutagenic activity using a bacterial reverse mutation assay (Ames test). The basis of this assay is the detection of substances, which by induction of mutations, can revert previous mutations present in the tester strains.⁹³

Assays were performed with *Salmonella typhimurium* strains TA98 and TA100. Since molecules can be converted to mutagenic compounds after metabolism, we performed the assay in the presence and absence of metabolic activation from the microsomal fraction (S9). The results are displayed in Table 6. The mutagenicity index (MI) was calculated according to the formula: MI = number of induced revertants/number of spontaneous revertants. The sample is considered to have mutagenic potential when the mutagenicity index was ≥ 2 in at least one of the concentrations tested and when there was a dose–response relationship between the concentrations tested and the number of induced revertants.⁹⁴

Analysis of the data presented in Table 6 suggests the studied molecule does not display mutagenic activity against *S. typhimurium* TA98 or TA100 strains. The calculated MI is less than 2 at all tested concentrations with or without metabolic activation. Further, a positive

dose–response relationship between concentrations and number of revertant colonies was not observed for the tested samples. Some concentrations showed a significant increase in the number of revertant colonies per plate ($*P < 0.05$, 1500, and 5000 $\mu\text{g/mL}$ for S9- and TA100 strain) when compared against the negative control, but overall, based on these data, the molecule can be considered nonmutagenic. Lastly, strain TA98 is used to detect frameshift mutations and strain TA100 detects substitution of DNA bases.⁹³ Compound **5a** was considered nonmutagenic to both TA98 and TA100 strains, suggesting that this dihydrothiazepine does not have the potential to cause gene mutations by frameshift or substitution of DNA bases. Of note, caveats remain for this assay because the compound may not penetrate into *S. typhimurium*. However, if this is the case, it reinforces the potential selectivity of this compound for *Chlamydia*.

Experimental Validation of the QSAR Model (KPLS—Radial 13).—The reliability of a QSAR model is conditioned to its ability to result in confident predictions, which can aid in the design of new molecules not used in the generation of the model.⁷⁵ Therefore, we designed three new antichlamydial dihydrothiazepines (**11d**, **11e**, **5x**, Figure 9a) to experimentally validate our QSAR model. In our new molecular design, we focused on modifications of the most active compounds from the initial dataset (**5a** and **5b**). To these initial structures, a trifluoromethyl group was inserted at ring B (structures **11d** and **11e**). Furthermore, since the presence of a sp^3 carbon from the methoxy in **5b** is detrimental for activity (marked in blue in the QSAR model representation in Figure 3a), we exchanged this group for a methyl substituent to get derivative **5x**. From this, we used our QSAR model to predict the antichlamydial activity of the proposed new molecules. Next, we synthesized and determined the antichlamydial activity of the designed new molecules using our semi-high-throughput screening assay. Details of the new compounds and predicted *versus* experimental activity comparisons can be found in Figure 9a,b. These simple modifications, as predicted by the QSAR model, resulted in more potent structures. Compound **11d** presented an IC_{50} value of 0.70 $\mu\text{g/mL}$ against 1.61 $\mu\text{g/mL}$ for the parent molecule **5a**. The same pattern is observed for structures **11e** (IC_{50} 1.08 $\mu\text{g/mL}$) and **5x** (IC_{50} 2.78 $\mu\text{g/mL}$) *versus* 3.32 $\mu\text{g/mL}$ for **5b** (parent molecule). The viability profile of the new most potent compounds (**11d** and **11e**) against HEp-2 cells indicated a very promising cytotoxic activity for these two structures (viability at 256 $\mu\text{g/mL}$: ~100%, Figure S4).

As Figure 9 shows, our KPLS-QSAR model predicted the antichlamydial activity for the new derivatives with a low prediction error (Figure 9b). Noteworthy, the model can effectively predict the activity trend of the molecules in a satisfactory manner. In addition, predictions of biological activities of new molecules using a QSAR model are conditioned to such molecules falling within the applicability domain (AD) of the model.⁹⁵ The AD corresponds to the chemical space covered by the nature of compounds in the training set.⁹⁶ We tested our new molecules **11d**, **11e**, and **5x** using the AutoQsar generated domain label alert,⁹⁷ which suggested that these compounds are within the applicability domain of the KPLS model. These results indicate that the presented QSAR model can be a support tool in the design and guided synthesis of new dihydrothiazepines. Therefore, the model's prediction power grants us the ability to rationalize our process of finding new highly potent molecules by focusing our synthesis on the molecules with superior predicted activity.

Further, this model can have its prediction power improved even more as we expand our library to cover a larger chemical space.

We understand our model (due to the nature of the intracellular assays discussed here) is not able to distinguish between variations in the IC₅₀ caused by differences in the permeability of the compounds or in their potency. There is a possibility that other targets in addition to the ClpX are affected, which will result in the antichlamydial activity being a result of multiple factors. However, we theorize the use of a QSAR model based on the overall antichlamydial activity is a better representation of potency than using different QSAR models (each of them based on one specific biological measurement). Experiments that seek to increase the permeability of dihydrothiazepines are currently underway.

CONCLUSIONS

The lack of drug selectivity is one of the main challenges of current therapies against CT infection owing to disruption of the microbiome and risk for the development of antibiotic resistance in other (non-CT) bacteria. In this study, we synthesized a new library of dihydrothiazepine derivatives and characterized their antichlamydial activity using a newly developed semi-high-throughput screening assay. Some of the synthesized derivatives presented substantial reduction in chlamydial inclusion number and size in infected HEp-2 cells. Further, our generated 2D fingerprint (radial 13) QSAR model was used to better elucidate the SAR using a visual projection of the quantitative contributions of different groups onto the chemical structures. Interestingly, our lead compound **5a** displayed a very promising selectivity profile in our initial evaluation when tested against a panel of both pathogenic bacterial species and microorganisms from the human microbiota. Additionally, this compound showed a good cytotoxicity profile against the tested human cell lines and no mutagenic potential was seen in the Ames test. Furthermore, **5a** proved to be stable in human serum and simulated gastric and intestinal fluids. A set of our dihydrothiazepines inhibited the ClpX ATPase activity *in vitro*, supporting ClpX as a target of this class of compounds. Our results indicate that our lead derivative **5a** is selective for CT and represents a starting point for the development of new selective antichlamydial drugs. In addition, we were able to use a validated QSAR model to guide the design of compounds with superior potency when compared to the parent molecules. This tool provides us the ability to acquire more potent molecules in the future in a quicker and rationalized manner.

EXPERIMENTAL PROCEDURES

Chemistry.

The solvents and reagents were from Fisher Scientific and used with no further purification. All chemical reactions were performed using oven-dried glassware. Qualitative analysis of reactions was performed by thin-layer chromatography (TLC) with silica gel as the adsorbent (Merck silica gel IB2-F plates 0.25 mm thickness). The visualization techniques employed to analyze TLC results were either ultraviolet light (254 nm) or iodine on silica. Chromatographic separations were performed on silica gel columns (normal phase silica columns, 20–40 μ m, RediSep Rf Gold high-performance columns) by a flash chromatography method on a RF 200i Flash Chromatography System from Teledyne

ISCO. The solvent system used was hexanes/ethyl acetate (Hex/EtOAc) with an increasing strength of the polar component from 0 to 100% during 25 min (unless otherwise stated). To perform structural characterization of the resulting molecules we used proton nuclear magnetic resonance (^1H NMR, Bruker instrument, 500 or 600 MHz) with deuterated dimethyl sulfoxide ($\text{DMSO}-d_6$), methanol ($\text{CD}_3\text{OD}-d_4$), or chloroform (CDCl_3) as solvents. ^1H NMR chemical shifts are reported in parts per million (δ ppm). Chemical purities of the final compounds were determined by high-performance liquid chromatography (HPLC)-UV, which confirmed >95% purity for all biologically tested compounds. The purity determination assay was performed using the following protocol: Agilent Technologies 1220 infinity LC instrument coupled with a multiple wavelength UV detector (acquisition 254 nm), Kinetex column from Phenomenex: $5\ \mu\text{m}$ (C18 reversed phase), $250 \times 4.6\ \text{mm}^2$ with a mobile phase of methanol/water, 1 mL/min flow, gradient from 0 to 100% methanol from 0 to 23 min, and maintained it until 25 min. All reported yields refer to final tested molecules. The compounds were named using the naming algorithm employed by ChemDraw Professional 15.0 (<https://perkinelmerinformatics.com/products/research/chemdraw/>).

General Procedure for the Knoevenagel Condensation Reaction (A).—

Knoevenagel condensation was performed to obtain 2-benzylidene-1*H*-indene-1,3(2*H*)-dione intermediates (**3a–y**, Scheme 1) by adapting the procedure reported by Fetzer and co-workers.⁵⁵ A suspension of ~ 1 mmol of indan-1,3-dione (**1**) in ethanol or methanol (see individual descriptions below, 10 mL) was added to the appropriate benzaldehyde (**2**) (1.10 equiv with respect to indan-1,3-dione) along with L-proline (0.30 equiv) and molecular sieves 4 Å. Specifically for derivative **3r**, 1 mmol of starting material **1** was reacted with 3-hydroxybenzaldehyde in 10 mL of water (no catalyst).⁹⁸ The reaction mixture was stirred at room temperature (RT) for 16–24 h (depending on the intermediate to be synthesized). Next, the reaction content was filtered using vacuum and washed with methanol (3×10 mL) to obtain a solid mass. These intermediates were used in the following reaction without further purification.

General Procedure for Cyclization in Acidic Media (B).—To obtain final compounds **5a–v**, 2-benzylidene-1*H*-indene-1,3(2*H*)-dione intermediates (**3**) were cyclized by adapting the method of Fetzer and co-workers (Scheme 1).⁵⁵ To a solution of a given benzylidene-1*H*-indene-1,3(2*H*)-dione (**3**) intermediate (amount indicated below according to different compounds) in isopropanol (*i*PrOH, 6 mL) were added acetic acid (AcOH, 4 mL), 2-aminothiophenol (**4**) (1.5 equiv with respect to benzylidene intermediate), and molecular sieves 4 Å. For some final compounds, this reaction was performed using *p*-toluenesulfonic acid (0.5 equiv amount indicated below according to the specific final compound), instead of acetic acid. The reaction mixture was stirred at RT or using higher temperature for 18–24 h depending on the intermediate to be synthesized (see individual descriptions below). The solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–100% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained products are listed below.

General Procedure for Nitro Reduction (C).—To a mixture of methanol (MeOH, 10 mL) and tetrahydrofuran (THF, 10 mL) was added 11-(4-nitrophenyl)-5,11-dihydro-12*H*-benzo[*b*]indeno[1,2-*e*]-[1,4]thiazepin-12-one (**5n**—0.362 mmol, Scheme 2). To this solution was added 20 mg of palladium on carbon (Pd/C, 10%). This mixture was stirred under a hydrogen gas atmosphere (with a balloon apparatus) for 24 h at room temperature. Next, the solution was filtered, the solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–100% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained products are listed below.

General Procedure for Thioester Oxidation (D).—To a solution of **5w** (68 mg, 0.18 mmol) in dry dichloromethane (DCM, 8 mL) was added *m*CPBA (107 mg, 0.620 mmol, Scheme 2), and the mixture was stirred for 5 h at room temperature. The mixture was quenched with water and extracted with ethyl acetate. The organic layer was dried over sodium sulfate and filtered. The solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–100% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained product are listed below.

General Procedure for Cyclization of Aminoethanethiol Derivatives (E).—Procedure to obtain final derivatives without ring B (**9a–d**, Scheme 3): To a solution of benzylidene-1*H*-indene-1,3(2*H*)-dione (**3**) intermediate (amount indicated below according to different compounds) in isopropanol (*i*PrOH, 6 mL) were added acetic acid (AcOH, 4 mL), 2-aminoethanethiol (**8**) (1.5 equiv with respect to benzylidene intermediate), and molecular sieves 4 Å. For some final compounds, this reaction was performed using *p*-toluenesulfonic acid (0.5 equiv amount indicated below according to the specific final compound), instead of acetic acid. The reaction mixture was stirred at RT or using higher temperature for 18–24 h depending on the intermediate to be synthesized. The solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–70% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained products are listed below.

General Procedure for Cyclization of Ring B Substituted Derivatives (F).—Procedure to obtain final derivatives containing a trifluoromethyl group on ring B (**11a–e**, Scheme 4): To a solution of benzylidene-1*H*-indene-1,3(2*H*)-dione intermediate (**3**) (amount indicated below according to different compounds) in isopropanol (*i*PrOH, 15 mL) were added 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (**10**) (1.5 equiv with respect to benzylidene intermediate) and molecular sieves 4 Å. The reaction mixture was stirred at room temperature for 18–24 h depending on the intermediate to be synthesized (see individual descriptions below). The solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–70% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained products are listed below.

General Procedure for Cyclization of Diazepine Derivatives (G).—Procedure to obtain final derivatives containing the diazepine group (**13a–c**, Scheme 4): In a three-neck round-bottom flask, to a solution of benzylidene-1*H*-indene-1,3(2*H*)-dione intermediate (**3**) (amount indicated below according to different compounds) in isopropanol (*i*PrOH, 6 mL) were added acetic acid (AcOH, 4 mL), *ortho*-phenylenediamine (**12**) (1.5 equiv with respect to benzylidene intermediate), and molecular sieves 4 Å. For some final compounds, this reaction was performed using *p*-toluenesulfonic acid (0.5 equiv amount indicated below according to the specific final compound) instead of acetic acid. The reaction mixture was stirred at room temperature for 18–24 h under a nitrogen gas atmosphere. The solvent was removed under vacuum, and the crude product was absorbed onto silica gel followed by purification using flash column chromatography eluting with a 0–70% gradient of EtOAc in hexanes. The physical characters and the spectral data of the obtained products are listed below.

Characterization of the Compounds.

2-((1,3-Dioxo-1,3-dihydro-2*H*-inden-2-ylidene)methyl)benzonitrile (**3a**).—

According to procedure A, 150 mg of indan-1,3-dione (**1**, 1.03 mmol) was added in 10 mL of MeOH. To this solution was added 148 mg of 2-cyanobenzaldehyde (1.13 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Orange solid, 258.00 mg (96.7%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 8.69 (s, 1H), 8.01 (br s, 6H), 7.89–7.76 (m, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆): 188.52, 187.62, 142.14, 139.88, 138.21, 136.50, 136.36, 134.57, 133.47, 132.98, 132.90, 132.25, 132.17, 23.46, 117.25, 113.85.

2-(2-Methoxybenzylidene)-1*H*-indene-1,3(2*H*)-dione (**3b**).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 115 mg of 2-methoxy-benzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 18 h at RT. Yellow solid, 131.60 mg (64.7%). ¹H NMR (500 MHz, CDCl₃): δ 8.90 (dd, *J* = 7.90, 1.65 Hz, 1H), 8.51 (s, 1H), 8.02–7.97 (m, 2H), 7.81–7.78 (m, 2H), 7.52 (td, *J* = 7.50, 1.70 Hz, 1H), 7.08 (t, *J* = 7.58 Hz, 1H), 6.95 (d, *J* = 8.29 Hz, 1H), 3.94 (s, 3H). ¹³C NMR (125 MHz, CDCl₃): 190.75, 189.41, 160.67, 142.53, 141.53, 140.23, 135.50, 135.29, 135.13, 134.10, 128.42, 123.32, 123.31, 122.18, 120.53, 110.80, 55.90.

2-(2-Fluoro-3-methylbenzylidene)-1*H*-indene-1,3(2*H*)-dione (**3c**).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of EtOH. To this solution was added 103 μL of 2-fluoro-3-methylbenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 118.50 mg (58.7%). ¹H NMR (500 MHz, CDCl₃): δ 8.26 (t, *J* = 6.95 Hz, 1H), 8.25 (s, 1H), 8.04–7.99 (m, 2H), 7.84–7.80 (m, 2H), 7.38 (t, *J* = 7.18 Hz, 1H), 7.18 (t, *J* = 7.73 Hz, 1H), 2.33 (s, 3H). ¹³C NMR (125 MHz, CDCl₃): 190.12, 189.14, 161.82 (d, *J*_{CF} = 256.01 Hz), 142.84, 140.51, 138.37 (d, *J*_{CF} = 8.03 Hz), 137.15 (d, *J*_{CF} = 6.40 Hz), 135.73 (d, *J*_{CF} = 15.20 Hz), 131.54, 130.40, 125.45 (d, *J*_{CF} = 17.59 Hz), 124.03 (d, *J*_{CF} = 4.51 Hz), 123.74 (d, *J*_{CF} = 2.82 Hz), 121.27 (d, *J*_{CF} = 2.82 Hz), 14.86 (d, *J*_{CF} = 4.64 Hz).

2-(3-Chlorobenzylidene)-1H-indene-1,3(2H)-dione (3d).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 113 μL of 3-chlorobenzaldehyde (0.814 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 18 h at RT. Yellow solid, 154.30 mg (74.8%). ^1H NMR (500 MHz, CDCl_3): δ 8.56 (t, J = 1.85 Hz, 1H), 8.25 (d, J = 7.75 Hz, 1H), 8.05–8.01 (m, 2H), 7.86–7.82 (m, 2H), 7.81 (s, 1H), 7.53–7.51 (m, 1H), 7.45 (t, J = 7.87 Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3): 189.90, 188.91, 145.04, 142.71, 140.27, 135.81, 135.63, 134.89, 134.72, 133.42, 132.96, 132.24, 130.42, 130.10, 123.69, 123.64.

2-(3-(Trifluoromethyl)benzylidene)-1H-indene-1,3(2H)-dione (3e).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 147 mg of 3-(trifluoromethyl)benzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. White solid, 146.5 mg (62.9%). ^1H NMR (500 MHz, CDCl_3): δ 8.80 (s, 1H), 8.57 (d, J = 7.84 Hz, 1H), 8.07–8.02 (m, 2H), 7.90 (s, 1H), 7.87–7.84 (m, 2H), 7.80 (d, J = 7.85 Hz, 1H), 7.65 (d, J = 7.85 Hz, 1H). ^{19}F NMR δ –62.86 (s). ^{13}C NMR (125 MHz, CDCl_3): 189.75, 188.89, 144.62, 142.71, 140.33, 136.88, 135.91, 135.75, 133.66, 131.62, 131.36, 130.85, 130.47 (q, J_{CF} = 3.81 Hz), 129.46, 129.26 (q, J_{CF} = 3.53 Hz), 123.78, 123.71.

2-(3-Nitrobenzylidene)-1H-indene-1,3(2H)-dione (3f).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 123 mg of 3-nitrobenzaldehyde (0.814 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. White solid, 146.0 mg (67.9%). ^1H NMR (600 MHz, CDCl_3): δ 9.57 (s, 1H), 8.68 (d, J = 7.68 Hz, 1H), 8.43 (d, J = 8.11 Hz, 1H), 8.07–8.04 (m, 2H), 8.00–7.99 (m, 3H), 7.85 (d, J = 7.97 Hz, 1H). ^{13}C NMR (150 MHz, CDCl_3): 188.91, 188.56, 147.85, 142.26, 142.10, 139.79, 139.77, 136.34, 136.25, 134.05, 131.62, 130.31, 127.05, 126.74, 123.48, 123.32.

2-(2,4-Dichlorobenzylidene)-1H-indene-1,3(2H)-dione (3g).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 148 mg of 2,4-dichlorobenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 18 h at RT. Yellow solid, 208.80 mg (89.5%). Spectral data are identical to those published in the literature.⁵⁵

2-(4-Chlorobenzylidene)-1H-indene-1,3(2H)-dione (3h).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 114 mg of 4-chlorobenzaldehyde (0.814 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 150.0 mg (72.7%). ^1H NMR (600 MHz, CDCl_3): δ 8.42 (d, J = 8.26 Hz, 2H), 8.03–7.99 (m, 2H), 7.84–7.81 (m, 3H), 7.48 (d, J = 8.28 Hz, 2H). ^{13}C NMR (150 MHz, CDCl_3): 190.15, 189.18, 145.34, 142.66, 140.23, 139.67, 135.69, 135.53, 135.51, 131.69, 129.60, 129.30, 123.59, 123.57.

2-(4-Methylbenzylidene)-1H-indene-1,3(2H)-dione (3i).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 100 μL of 4-methylbenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25

mmol). The reaction was stirred for 20 h at RT. Yellow solid, 131.0 mg (68.6%). ^1H NMR (500 MHz, $\text{CD}_3\text{OD}-d_4$): δ 8.25 (d, J = 6.98 Hz, 2H), 7.87 (s, 2H), 7.75–7.69 (m, 3H), 7.20 (d, J = 7.07 Hz, 2H), 2.32 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3 , some drops of $\text{CD}_3\text{OD}-d_4$): 190.94, 189.39, 147.40, 144.87, 139.87, 135.46, 135.21, 134.48, 130.51, 129.61, 128.01, 124.52, 123.22, 21.87.

4-((1,3-Dioxo-1,3-dihydro-2H-inden-2-ylidene)methyl)phenyl Acetate (3j).—

According to procedure A, 146 mg of indan-1,3-dione (**1**, 1.00 mmol) was added in 10 mL of MeOH. To this solution was added 154 μL of 2-acetoxy-benzaldehyde (1.10 mmol) and 32 mg of L-proline (0.28 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 241.7 mg (82.8%). ^1H NMR (500 MHz, CDCl_3): δ 8.55 (d, J = 8.74 Hz, 2H), 8.04–8.00 (m, 2H), 7.88 (s, 1H), 7.85–7.81 (m, 2H), 7.28–7.26 (m, 2H), 2.35 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3): 190.33, 189.22, 168.92, 154.44, 145.79, 142.64, 140.22, 136.01, 135.59, 135.42, 130.93, 129.12, 123.53, 123.50, 122.16, 21.36.

2-(4-(Trifluoromethoxy)benzylidene)-1H-indene-1,3(2H)-dione (3k).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 120 μL of 4-(trifluoromethoxy)benzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 136.5 mg (55.7%). ^1H NMR (500 MHz, CDCl_3): δ 8.52 (s, 2H), 8.04–7.99 (m, 2H), 7.84–7.81 (m, 3H), 7.80 (d, J = 2.04 Hz, 2H). ^{13}C NMR (125 MHz, CDCl_3): 190.24, 189.29, 152.65 (q, J_{CF} = 1.55 Hz), 145.07, 142.83, 140.44, 136.34, 135.92, 135.77, 131.70, 129.94, 123.81, 123.78, 120.77 (q, J_{CF} = 257.84 Hz), 120.71.

2-(4-Bromobenzylidene)-1H-indene-1,3(2H)-dione (3l).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 156 mg of 4-bromobenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 17 h at RT. Yellow solid, 205.3 mg (85.2%). Spectral data identical to the literature.⁹⁹

3-((1,3-Dioxo-1,3-dihydro-2H-inden-2-ylidene)methyl)benzonitrile (3m).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 111 mg of 3-cyanobenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 22 h at RT. White solid, 179.7 mg (90.3%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 8.94 (s, 1H), 8.68 (d, J = 9.48 Hz, 1H), 8.07–7.98 (m, 5H), 7.90 (s, 1H), 7.78 (t, J = 9.42 Hz, 1H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 188.95, 188.49, 142.32, 142.07, 139.72, 137.64, 136.34, 136.22, 135.47, 133.64, 131.37, 129.94, 123.42, 123.30, 118.29, 111.80.

2-(4-Nitrobenzylidene)-1H-indene-1,3(2H)-dione (3n).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 128 mg of 4-nitrobenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 24 h at RT. Yellow solid, 174.90 mg (81.4%). ^1H NMR (500 MHz, CDCl_3): δ 8.55 (d, J = 8.80 Hz, 2H), 8.33 (d, J = 8.86 Hz, 2H), 8.08–8.04 (m, 2H), 7.90–7.87 (m, 3H). ^{13}C NMR (125 MHz, CDCl_3): 189.26, 188.54, 149.60, 142.80, 142.77, 140.42, 138.57, 136.16, 136.06, 134.40, 132.39, 123.93, 123.88, 123.83.

4-((1,3-Dioxo-1,3-dihydro-2H-inden-2-ylidene)methyl)benzonitrile (3o).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 128 mg of 4-cyanobenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 162.6 mg (81.7%). ¹H NMR (500 MHz, CDCl₃, with some drops of CD₃OD-*d*₄): δ 8.40 (d, *J* = 7.60 Hz, 2H), 7.80 (s, 2H), 7.80–7.77 (m, 3H), 7.71 (d, *J* = 7.60 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃, with some drops of CD₃OD-*d*₄): 189.54, 188.60, 143.47, 142.53, 140.15, 136.76, 136.11, 135.97, 133.72, 132.26, 131.81, 123.71, 123.66, 118.19, 115.15.

2-(4-Hydroxybenzylidene)-1H-indene-1,3(2H)-dione (3p).—

According to procedure A, 146 mg of indan-1,3-dione (**1**, 1 mmol) was added in 10 mL of MeOH. To this solution was added 134 mg of 4-hydroxybenzaldehyde (1.100 mmol) and 56 mg of L-proline (0.50 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 97.4 mg (39.0%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.88 (s br, 1H), 8.54 (d, *J* = 8.79 Hz, 2H), 7.96–7.90 (m, 4H), 7.76 (s, 1H), 6.95 (d, *J* = 8.82 Hz, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆): 190.00, 189.04, 163.30, 146.29, 141.65, 139.22, 137.61, 135.58, 135.41, 125.19, 124.60, 122.76, 122.71, 116.00.

2-(4-Methoxybenzylidene)-1H-indene-1,3(2H)-dione (3q).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 102.5 μL of 4-methoxybenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 20 h at RT. Yellow solid, 152.3 mg (75.0%). Spectral data identical to the literature.⁹⁹

2-(3-Hydroxybenzylidene)-1H-indene-1,3(2H)-dione (3r).—

According to procedure A, 146 mg of indan-1,3-dione (**1**, 1.0 mmol) was added in 10 mL of water. To this solution was added 134 mg of 3-hydroxybenzaldehyde (1.1 mmol). The reaction was stirred for 19 h at RT. Yellow solid, 199.2 mg (79.7%). Spectral data identical to the literature.¹⁰⁰

2-(4-(Benzyloxy)benzylidene)-1H-indene-1,3(2H)-dione (3s).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of EtOH. To this solution was added 179 mg of 4-(benzyloxy)benzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 19 h at RT. Yellow solid, 160.8 mg (61.1%). ¹H NMR (500 MHz, CDCl₃): δ 8.57 (d, *J* = 8.86 Hz, 2H), 8.00–7.97 (m, 2H), 7.84 (s, 1H), 7.81–7.77 (m, 2H), 7.45–7.40 (m, 4H), 7.36 (t, *J* = 7.25 Hz, 1H), 7.09 (d, *J* = 8.95 Hz, 2H), 5.18 (s, 2H). ¹³C NMR (125 MHz, CDCl₃): 190.95, 189.67, 163.33, 146.91, 142.53, 140.12, 137.33, 136.12, 135.23, 135.02, 128.88, 128.47, 127.67, 127.62, 126.82, 126.80, 123.23, 115.33, 70.41.

4-((1,3-Dioxo-1,3-dihydro-2H-inden-2-ylidene)methyl)benzoic Acid (3t).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 127 mg of 4-carboxaldehyde (1.1 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 18 h at RT. Yellow solid, 181.5 mg (84.7%). Spectral data identical to the literature.⁵⁵

2-((1,3-Dioxo-1,3-dihydro-2H-inden-2-ylidene)methyl)benzoic Acid (3u).—

According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 127 mg of 2-carboxaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 23 h at RT. White solid, 155.6 mg (72.7%). Spectral data are identical to those published in the literature.⁹⁹

2-([1,1'-Biphenyl]-4-ylmethylene)-1H-indene-1,3(2H)-dione (3v).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 150 mg of [1,1'-biphenyl]-4-carbaldehyde (0.824 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 206.0 mg (86.2%). Spectral data are identical to those published in the literature.¹⁰¹

2-(3-Methoxybenzylidene)-1H-indene-1,3(2H)-dione (3w).—According to procedure A, 146 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 118 mg of 3-methoxybenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 16 h at RT. Yellow solid, 190.0 mg (84.9%). ¹H NMR (600 MHz, CDCl₃): δ 8.46 (s, 1H), 8.04–8.01 (m, 2H), 7.89 (s, 1H), 7.84–7.81 (m, 2H), 7.79 (t, *J* = 7.62 Hz, 1H), 7.41 (t, *J* = 7.92 Hz, 1H), (dd, *J* = 8.28, 2.46 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃): 190.44, 189.32, 159.87, 147.34, 142.73, 140.16, 135.59, 135.40, 134.51, 129.79, 129.30, 127.89, 123.52, 123.51, 120.97, 117.02, 55.68.

2-(2-Methylbenzylidene)-1H-indene-1,3(2H)-dione (3x).—According to procedure A, 112 mg of indan-1,3-dione (**1**, 0.770 mmol) was added in 10 mL of MeOH. To this solution was added 98.2 μL of 2-methylbenzaldehyde (0.847 mmol) and 28 mg of L-proline (0.25 mmol). The reaction was stirred for 18 h at RT. Yellow solid, 132.6 mg (64.7%). ¹H NMR (600 MHz, CDCl₃): δ 8.54 (d, *J* = 7.76 Hz, 1H), 8.24 (s, 1H), 8.04–7.99 (m, 2H), 7.84–7.81 (m, 2H), 7.42 (td, *J* = 7.44, 1.02 Hz, 1H), 7.33 (t, *J* = 7.57 Hz, 1H), 7.28 (d, *J* = 7.56 Hz, 1H), 2.55 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): 190.41, 188.94, 144.49, 142.65, 140.98, 140.20, 135.57, 135.38, 132.91, 132.63, 131.60, 130.70, 129.33, 126.05, 123.50, 123.47, 20.50.

2-(Thiophen-2-ylmethylene)-1H-indene-1,3(2H)-dione (3y).—According to procedure A, 300 mg of indan-1,3-dione (**1**, 2.05 mmol) was added in 20 mL of MeOH. To this solution was added 210 μL of 2-thiophenecarboxaldehyde (2.26 mmol) and 56 mg of L-proline (0.50 mmol). The reaction was stirred for 16 h at RT. Orange solid, 464.4 mg (94.2%). Spectral data are identical to those published in the literature.⁹⁹

2-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzonitrile (5a).—According to general procedure B, 100 mg of **3a** (0.386 mmol), 61.9 μL of 2-aminothiophenol (0.579 mmol), 4 mL of acetic acid, 6 mL of *i*PrOH, and molecular sieves 4 Å were reacted for 20 h at RT. Orange solid, 79.73 mg (56.4%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.26 (s, 1H), 8.15 (d, *J* = 7.37 Hz, 1H), 7.80 (dd, *J* = 7.56, 1.14 Hz, 1H), 7.63 (d, *J* = 8.22 Hz, 1H), 7.58 (td, *J* = 7.56, 1.02 Hz, 1H), 7.44 (t, *J* = 7.35 Hz, 1H), 7.41–7.38 (m, 2H), 7.25 (td, *J* = 7.56, 0.94 Hz, 1H), 7.18 (td, *J* = 7.74, 1.26 Hz, 1H), 6.94–6.91 (m, 2H), 6.62 (d, *J* = 7.84 Hz, 1H), 5.63 (s, 1H). ¹³C NMR (150 MHz, DMSO-

d_6): 189.17, 157.25, 145.17, 143.50, 139.19, 136.05, 133.43, 132.54, 132.26, 131.90, 130.34, 129.71, 127.62, 126.78, 125.14, 123.81, 122.93, 120.69, 119.54, 117.46, 110.69, 106.89, 44.17. High-resolution mass spectrometry-electrospray ionization (HRMS-ESI) (m/z) calcd for $C_{23}H_{14}N_2OS$ $[M + H]^+$ 367.0900, found 367.0914. HPLC purity: 95.4%.

11-(2-Methoxyphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5b).—According to general

procedure B, 50 mg of **3b** (0.203 mmol), 32.6 μ L of 2-aminothiophenol (0.305 mmol), 4 mL of acetic acid, 6 mL of *i*PrOH, and molecular sieves 4 Å were reacted for 19 h at 70 °C. Red solid, 15.05 mg (19.8%). 1H NMR (600 MHz, DMSO- d_6): δ 10.04 (s, 1H), 8.11 (d, J = 7.44 Hz, 1H), 7.60 (d, J = 7.97 Hz, 1H), 7.55 (td, J = 7.56, 0.96 Hz, 1H), 7.40 (d, J = 7.33 Hz, 1H), 7.35–7.32 (m, 2H), 7.03 (td, J = 8.28, 1.62 Hz, 1H), 6.94 (d, J = 8.10 Hz, 1H), 6.90–6.87 (m, 2H), 6.40 (t, J = 7.47 Hz, 1H), 6.28 (t, J = 7.59, 1.47 Hz, 1H), 5.62 (s, 1H), 3.91 (s, 3H). ^{13}C NMR (150 MHz, DMSO- d_6): 189.16, 156.87, 155.75, 143.21, 139.36, 136.11, 132.65, 131.61, 130.09, 129.95, 128.91, 127.95, 126.36, 124.55, 124.33, 123.20, 120.37, 119.05, 119.03, 110.75, 109.23, 55.68, 40.03. HRMS-ESI (m/z) calcd for $C_{23}H_{17}NO_2S$ $[M + Na]^+$ 394.0878, found 394.2260. HPLC purity: 98.3%.

11-(2-Fluoro-3-methylphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5c).—According to procedure B, 50 mg of **3c**

(0.188 mmol), 30.20 μ L of 2-aminothiophenol (0.282 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 48.80 mg (69.1%). 1H NMR (500 MHz, DMSO- d_6): δ 10.12 (s, 1H), 8.13 (d, J = 7.38 Hz, 1H), 7.61 (d, J = 8.06 Hz, 1H), 7.56 (t, J = 7.44 Hz, 1H), 7.42 (t, J = 7.45 Hz, 1H), 7.39–7.36 (m, 2H), 6.99–6.91 (m, 3H), 6.56 (t, J = 7.62 Hz, 1H), 6.24 (t, J = 7.25 Hz, 1H), 5.56 (s, 1H), 2.23 (s, 3H). ^{19}F NMR δ –121.87 (s). ^{13}C NMR (150 MHz, DMSO- d_6): 189.12, 157.69 (d, J_{CF} = 244.66 Hz), 157.01, 143.33, 139.25, 136.05, 132.56, 131.73, 130.16 (d, J_{CF} = 7.86 Hz), 129.29, 129.07 (d, J_{CF} = 13.71 Hz), 125.16 (d, J_{CF} = 2.15 Hz), 124.86, 123.87 (d, J_{CF} = 17.15 Hz), 123.73, 123.47, 122.65 (d, J_{CF} = 3.86 Hz), 120.52, 119.27, 108.07, 40.05, 14.14 (d, J_{CF} = 3.92 Hz). HRMS-ESI (m/z) calcd for $C_{23}H_{16}FNOS$ $[M + H]^+$ 374.1009, found 374.1008. HPLC purity: 95.8%.

11-(3-Chlorophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5d).—According to procedure B, 50 mg of **3d** (0.186 mmol), 20.55 μ L of 2-

aminothiophenol (0.219 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 43.00 mg (61.8%). 1H NMR (500 MHz, DMSO- d_6): δ 10.11 (s, 1H), 8.11 (d, J = 7.39 Hz, 1H), 7.60–7.55 (m, 2H), 7.43 (t, J = 7.39 Hz, 1H), 7.39–7.34 (m, 2H), 7.09–7.06 (m, 3H), 7.00–6.94 (m, 3H), 5.55 (s, 1H). ^{13}C NMR (125 MHz, DMSO- d_6): 189.25, 156.68, 144.81, 143.35, 139.18, 136.03, 132.60, 132.46, 131.70, 130.13, 129.51, 129.24, 127.04, 126.36, 125.74, 124.88, 123.86, 123.50, 120.50, 119.28, 108.20, 45.16. HRMS-ESI (m/z) calcd for $C_{22}H_{14}ClNOS$ $[M + H]^+$ 376.0557, found 376.0559. HPLC purity: 97.5%.

11-(3-(Trifluoromethyl)phenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5e).—According to procedure B, 50

mg of **3e** (0.166 mmol), 18.7 μ L of 2-aminothiophenol (0.199 mmol), 4 mL of acetic

acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 18.26 mg (26.9%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.15 (s, 1H), 8.12 (d, *J* = 7.40 Hz, 1H), 7.59–7.55 (m, 2H), 7.43 (d, *J* = 7.55 Hz, 1H), 7.39–7.34 (m, 3H), 7.32–7.28 (m, 3H), 6.92–6.91 (m, 2H), 5.68 (s, 1H). ¹⁹F NMR δ –61.18 (s). ¹³C NMR (150 MHz, DMSO-*d*₆): 189.27, 156.83, 143.84, 143.38, 139.15, 136.04, 132.60, 131.76, 131.11, 130.20, 129.29, 128.84, 128.40 (q, *J*_{CF} = 31.51 Hz), 124.95, 123.80, 123.57, 123.40 (q, *J*_{CF} = 4.01 Hz), 123.11 (q, *J*_{CF} = 3.89 Hz), 120.56, 119.34, 107.97, 45.29. HRMS-matrix-assisted laser desorption ionization (MALDI) (*m/z*) calcd for C₂₃H₁₄F₃NOS [M + H]⁺ 410.0820, found 410.0726. HPLC purity: 96.7%.

11-(3-Nitrophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5f).—According to procedure B, 50 mg of **3f** (0.179 mmol), 20.2 μL of 2-aminothiophenol (0.214 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 18.3 mg (26.9%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.29 (s, 1H), 8.14 (d, *J* = 7.38 Hz, 1H), 7.90 (dd, *J* = 8.16, 1.59 Hz, 1H), 7.83 (t, *J* = 1.73 Hz, 1H), 7.61–7.56 (m, 2H), 7.51–7.49 (m, 1H), 7.45–7.43 (m, 1H), 7.40–7.34 (m, 3H), 6.95–6.90 (m, 2H), 5.76 (s, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆): 189.26, 156.96, 147.20, 144.78, 143.45, 139.11, 136.08, 133.68, 132.58, 131.81, 130.27, 129.45, 129.35, 125.06, 123.68, 123.59, 121.44, 121.42, 120.61, 119.42, 107.64, 45.06. HRMS-MALDI (*m/z*) calcd for C₂₂H₁₄N₂O₃S [M + Na]⁺ 409.0623, found 409.0959. HPLC purity: 98.13%.

11-(2,4-Dichlorophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5g).—According to procedure B, 50 mg of **3g** (0.16 mmol), 26.5 μL of 2-aminothiophenol (0.247 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Orange solid, 27.2 mg (40.0%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 10.22 (s, 1H), 8.14 (d, *J* = 7.38 Hz, 1H), 7.63 (d, *J* = 8.14 Hz, 1H), 7.59–7.56 (m, 2H), 7.45–7.37 (m, 3H), 6.97–6.89 (m, 2H), 6.90 (dd, *J* = 8.45 Hz, 1H), 6.48 (d, *J* = 8.43 Hz, 1H), 5.58 (s, 1H). ¹³C NMR (125 MHz, DMSO-*d*₆): 189.06, 157.22, 143.38, 139.18, 138.18, 136.15, 133.21, 132.48, 132.00, 131.80, 130.24, 129.58, 128.93, 128.75, 126.17, 125.10, 123.61, 123.10, 120.60, 119.41, 107.53, 42.86. HRMS-MALDI (*m/z*) calcd for C₂₂H₁₃Cl₂NOS [M + H]⁺ 410.0167, found 410.0040. HPLC purity: 97.0%.

11-(4-Chlorophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5h).—According to procedure B, 50 mg of **3h** (0.19 mmol), 23.9 μL of 2-aminothiophenol (0.22 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 25 mg (35%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.07 (s, 1H), 8.11 (d, *J* = 7.38 Hz, 1H), 7.57–7.54 (m, 2H), 7.42 (t, *J* = 7.31 Hz, 1H), 7.38–7.33 (m, 2H), 7.12 (d, *J* = 8.53 Hz, 2H), 7.03 (d, *J* = 8.51 Hz, 2H), 6.98 (dd, *J* = 7.68, 1.44 Hz, 1H), 6.94 (td, *J* = 7.74, 1.01 Hz, 1H), 5.53 (s, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆): 189.25, 156.64, 143.38, 141.34, 139.24, 136.07, 132.64, 131.70, 130.87, 130.11, 129.21, 128.99, 127.69, 124.83, 123.94, 123.51, 120.49, 119.22, 108.55, 45.05. HRMS-MALDI (*m/z*) calcd for C₂₂H₁₄ClNOS [M + H]⁺ 376.0557, found 376.0560. HPLC purity: 99.4%.

11-(*p*-Tolyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5i).—According to procedure B, 50 mg of **3i** (0.20 mmol), 25.9 μL of 2-aminothiophenol (0.242 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 24 h at RT. Orange

solid, 42.83 mg (59.4%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 9.99 (s, 1H), 8.10 (d, $J = 7.38$ Hz, 1H), 7.57–7.54 (m, 2H), 7.42 (t, $J = 7.29$ Hz, 1H), 7.38 (d, $J = 6.65$ Hz, 1H), 7.35–7.32 (m, 1H), 6.98 (dd, $J = 7.70$, 1.45 Hz, 1H), 6.94–6.86 (m, 5H), 5.45 (s, 1H), 2.13 (s, 3H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ 189.23, 156.34, 143.37, 139.32, 139.26, 136.05, 135.33, 132.69, 131.61, 129.97, 128.95, 128.32, 127.09, 124.60, 124.27, 123.30, 120.39, 119.08, 109.39, 45.46, 20.48. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{17}\text{NOS}$ [$\text{M} + \text{H}$] $^+$ 356.1104, found 356.1102. HPLC purity: 98.9%.

4-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)phenyl Acetate (5j).—According to procedure B, 150 mg of **3j** (0.51 mmol), 2-aminothiophenol (0.77 mmol), 8 mL of acetic acid, and 12 mL of *i*PrOH were reacted for 24 h at RT. Orange solid, 102 mg (50.0%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 10.04 (s, 1H), 8.11 (d, $J = 7.39$ Hz, 1H), 7.57–7.54 (m, 2H), 7.42 (t, $J = 7.30$ Hz, 1H), 7.37 (d, $J = 6.55$ Hz, 1H), 7.36–7.32 (m, 1H), 7.04 (d, $J = 8.45$ Hz, 2H), 6.99 (dd, $J = 7.70$ Hz, 1.45 Hz, 1H), 6.92 (td, $J = 7.65$, 1.13, 1.45 Hz, 1H), 6.82 (d, $J = 8.60$ Hz, 2H), 5.52 (s, 1H), 2.17 (s, 3H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 189.26, 168.95, 156.48, 148.73, 143.37, 139.70, 139.28, 136.05, 132.66, 131.65, 130.04, 129.08, 128.41, 124.70, 124.06, 123.43, 121.00, 120.44, 119.16, 109.00, 45.16, 20.77. HRMS-MALDI (m/z) calcd for $\text{C}_{24}\text{H}_{17}\text{NO}_3\text{S}$ [$\text{M} + \text{Na}$] $^+$ 422.0827, found 422.2483. HPLC purity: 98.2%.

11-(4-(Trifluoromethoxy)phenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5k).—According to procedure B, 50 mg of **3k** (0.16 mmol), 17.7 μL of 2-aminothiophenol (0.199 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 37.6 mg (56.4%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 10.08 (s, 1H), 8.11 (d, $J = 7.39$ Hz, 1H), 7.58–7.54 (m, 2H), 7.43 (t, $J = 7.29$ Hz, 1H), 7.38–7.33 (m, 2H), 7.14 (d, $J = 8.64$ Hz, 2H), 7.05 (d, $J = 8.34$ Hz, 2H), 6.97 (dd, $J = 7.75$, 1.5 Hz, 1H), 6.94–6.91 (m, 1H), 5.58 (s, 1H). ^{19}F NMR δ –56.89 (s). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 189.26, 156.65, 146.10, 143.38, 141.70, 139.23, 136.02, 132.63, 131.70, 130.10, 129.21, 128.96, 124.73, 123.89, 123.53, 120.48, 120.23, 119.23, 108.51, 44.96. HRMS-MALDI (m/z) calcd for $\text{C}_{23}\text{H}_{14}\text{F}_3\text{NO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$ 426.0769, found 426.0709. HPLC purity: 96.7%.

11-(4-Bromophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5l).—According to procedure B, 50 mg of **3l** (0.16 mmol), 18.1 μL of 2-aminothiophenol (0.168 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Orange solid, 33.7 mg (50.2%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 10.06 (s, 1H), 8.11 (d, $J = 7.35$ Hz, 1H), 7.57–7.54 (m, 2H), 7.42 (t, $J = 7.25$ Hz, 1H), 7.38–7.34 (m, 2H), 7.26 (d, $J = 8.50$ Hz, 2H), 7.00–6.93 (m, 4H), 5.51 (s, 1H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): 189.24, 156.62, 143.37, 141.76, 139.23, 136.06, 132.62, 131.69, 130.60, 130.09, 129.35, 129.21, 124.82, 123.91, 123.50, 120.48, 119.43, 119.21, 108.48, 45.10. HRMS-MALDI (m/z) calcd for $\text{C}_{22}\text{H}_{14}\text{BrNOS}$ [$\text{M} + \text{H}$] $^+$ 420.0051, found 420.0066. HPLC purity: 97.2%.

3-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzonitrile (5m).—According to procedure B, 50

mg of **3m** (0.19 mmol), 31 μL of 2-aminothiophenol (0.29 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at 75 °C. Orange solid, 17.58 mg (22.2%). ^1H NMR (500 MHz, DMSO- d_6): δ 10.13 (s, 1H), 8.12 (d, J = 7.35 Hz, 1H), 7.60–7.55 (m, 2H), 7.50 (d, J = 7.62 Hz, 1H), 7.45–7.42 (m, 2H), 7.39–7.35 (m, 3H), 7.29 (t, J = 7.77 Hz, 1H), 6.96–6.91 (m, 2H), 5.64 (s, 1H). ^{13}C NMR (125 MHz, DMSO- d_6): 189.73, 157.47, 144.52, 143.91, 139.70, 136.52, 133.12, 132.42, 132.18, 130.93, 130.71, 130.62, 129.84, 129.50, 125.43, 124.12, 124.11, 120.99, 119.82, 119.04, 111.11, 108.06, 45.52. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{14}\text{N}_2\text{OS}$ [$\text{M} + \text{H}$] $^+$ 367.0900, found 367.0899. HPLC purity: 96.3%.

11-(4-Nitrophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5n).—According to procedure B, 50 mg of **3n** (0.58 mmol), 92.3 μL of 2-aminothiophenol (0.868 mmol), 50 mg of *p*-toluenesulfonic acid monohydrate (0.260 mmol), and 20 mL of *i*PrOH were reacted for 24 h at RT. Orange solid, 213.25 mg (95.3%). ^1H NMR (500 MHz, DMSO- d_6): δ 10.16 (s, 1H), 8.14 (d, J = 7.39 Hz, 1H), 7.94 (d, J = 8.77 Hz, 2H), 7.60–7.56 (m, 2H), 7.43 (t, J = 7.31 Hz, 1H), 7.40–7.35 (m, 2H), 7.30 (d, J = 8.72 Hz, 2H), 6.97 (dd, J = 7.70, 1.45 Hz, 1H), 6.92 (t, J = 7.41 Hz, 1H), 5.71 (s, 1H). ^{13}C NMR (125 MHz, DMSO- d_6): 189.29, 156.84, 150.07, 145.78, 143.35, 139.15, 135.98, 132.58, 131.78, 130.21, 129.44, 128.39, 124.95, 123.66, 123.46, 122.99, 120.57, 119.36, 107.58, 45.19. HRMS-ESI (m/z) calcd for $\text{C}_{22}\text{H}_{14}\text{N}_2\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$ 387.0798, found 387.0796. HPLC purity: 97.1%.

4-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzonitrile (5o).—According to procedure B, 50 mg of **3o** (0.19 mmol), 31 μL of 2-aminothiophenol (0.29 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 24 h at RT. Orange solid, 67.6 mg (95.3%). ^1H NMR (500 MHz, $\text{CD}_3\text{OD}-d_4$): δ 7.92 (d, J = 7.35 Hz, 1H), 7.55–7.51 (m, 1H), 7.49 (dd, J = 8.10, 0.93 Hz, 1H), 7.45–7.42 (m, 4H), 7.33 (dd, J = 8.35, 1.47 Hz, 1H), 7.24 (d, J = 8.25 Hz, 2H), 7.01 (dd, J = 7.75, 1.45 Hz, 1H), 6.94 (td, J = 7.50, 1.10 Hz, 1H), 5.57 (s, 1H). ^{13}C NMR (125 MHz, DMSO- d_6): δ 189.28, 156.83, 147.95, 143.35, 139.19, 135.97, 132.59, 131.75, 130.16, 129.37, 128.11, 124.89, 123.62, 123.54, 120.53, 119.31, 118.65, 109.00, 107.61, 45.40. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{14}\text{N}_2\text{OS}$ [$\text{M} + \text{H}$] $^+$ 367.0900, found 367.0898. HPLC purity: 99.2%.

11-(4-Hydroxyphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5p).—According to procedure B, 97.4 mg of **3p** (0.39 mmol), 2-aminothiophenol (0.58 mmol), 8 mL of acetic acid, 12 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Red solid, 13.00 mg (9.3%). Spectral data are identical to those published in the literature.⁵⁵ HPLC purity: 97.4%.

11-(4-Methoxyphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5q).—According to procedure B, 50 mg of **3q** (0.22 mmol), 25.3 μL of 2-aminothiophenol (0.265 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 14.3 mg (17.4%). ^1H NMR (500 MHz, DMSO- d_6): δ 9.98 (s, 1H), 8.06 (d, J = 7.38 Hz, 1H), 7.56–7.53 (m, 2H), 7.41 (t, J = 7.28 Hz, 1H), 7.37–7.31 (m, 2H), 6.99 (dd, J = 7.70, 1.45 Hz, 1H), 6.94–6.91 (m, 3H), 6.62 (d, J = 8.75 Hz, 2H), 5.44 (s, 1H), 3.60 (s, 3H). ^{13}C NMR (125 MHz, DMSO- d_6):

189.20, 157.59, 156.31, 143.38, 139.32, 136.09, 134.25, 132.69, 131.61, 129.97, 128.94, 128.30, 124.61, 124.37, 123.32, 120.38, 119.07, 113.06, 109.61, 54.88, 45.27. HRMS-ESI (m/z) calcd for $C_{23}H_{17}NO_2S$ $[M + H]^+$ 372.1053, found 372.1051. HPLC purity: 97.8%.

11-(3-Hydroxyphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (334).—According to procedure

B, 60 mg of **3r** (0.17 mmol), 2-aminoethanethiol (19.4 mg, 0.252 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 19 h at RT. Red solid, 44.4 mg (52.0%). Spectral data are identical to those published in the literature.⁵⁵ HPLC purity: 95.9%.

11-(4-Aminophenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (6a).—According to procedure C, 140 mg of **5n** (0.362 mmol), 10 mL of dried

MeOH, and 10 mL of dried THF were added in a three-neck round-bottom flask under a nitrogen atmosphere. To this solution was added 20 mg of palladium on carbon (Pd/C, 10%) and the solution was stirred for 24 h at RT. Red solid, 65.7 mg (52.3%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 9.92 (s, 1H), 8.07 (d, J = 7.36 Hz, 1H), 7.53 (td, J = 7.38, 1.02 Hz, 2H), 7.40 (t, J = 7.31 Hz, 1H), 7.35 (d, J = 6.93 Hz, 1H), 7.31 (td, J = 7.38, 1.46 Hz, 1H), 6.99 (dd, J = 7.74, 1.50 Hz, 1H), 6.92 (td, J = 7.50, 1.20 Hz, 1H), 6.65 (d, J = 8.40 Hz, 2H), 6.22 (d, J = 4.25 Hz, 2H), 5.32 (s, 1H), 4.84 (s, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆): 189.15, 156.17, 146.90, 143.39, 139.40, 136.21, 132.80, 131.54, 129.88, 129.14, 128.71, 127.83, 124.84, 124.52, 123.18, 120.32, 118.94, 113.17, 110.32, 45.83. HRMS-ESI (m/z) calcd for $C_{22}H_{16}N_2OS$ $[M + H]^+$ 357.1056, found 357.1059. HPLC purity: 95.9%.

11-(4-(Benzyloxy)phenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e]

[1,4]thiazepin-12-one (5s).—According to procedure B, 50 mg of **3s** (0.15 mmol), 23.5 μ L of 2-aminothiophenol (0.220 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at 80 °C. Orange solid, 13.7 mg (20.0%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.99 (s, 1H), 8.10 (d, J = 7.35 Hz, 1H), 7.56–7.53 (m, 2H), 7.41 (t, J = 7.25 Hz, 1H), 7.37–7.29 (m, 7H), 7.00 (d, J = 7.62 Hz, 1H), 6.94–6.91 (m, 3H), 6.70 (d, J = 8.65 Hz, 2H), 5.45 (s, 1H), 4.94 (s, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆): 189.20, 156.71, 156.31, 143.38, 139.32, 136.95, 136.08, 134.51, 132.68, 131.60, 129.96, 128.94, 128.33, 127.74, 127.65, 124.62, 124.35, 123.34, 120.38, 119.07, 113.94, 109.57, 69.04, 45.26. HRMS-MALDI (m/z) calcd for $C_{29}H_{21}NO_2S$ $[M + H]^+$ 448.1365, found 448.0505. HPLC purity: 96.4%.

4-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzoic

Acid (5t).—According to procedure B, 50 mg of **3t** (0.18 mmol), 28.8 μ L of 2-aminothiophenol (0.269 mmol), 25 mg of *p*-toluenesulfonic acid monohydrate (0.130 mmol), and 10 mL of *i*PrOH were reacted for 20 h at RT. Orange solid, 30.8 mg (44.7%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 12.80 (s, 1H), 10.10 (s, 1H), 8.12 (d, J = 7.40 Hz, 1H), 7.64 (d, J = 8.35 Hz, 2H), 7.58–7.54 (m, 2H), 7.42 (t, J = 7.30 Hz, 1H), 7.38 (d, J = 7.55 Hz, 1H), 7.36–7.32 (m, 1H), 7.13 (d, J = 8.25 Hz, 2H), 6.96 (dd, J = 7.65 Hz, 1H), 6.92–6.89 (m, 1H), 5.59 (s, 1H). ¹³C NMR (125 MHz, DMSO-*d*₆): 189.29, 166.97, 156.69, 147.20, 143.37, 139.25, 136.01, 132.67, 131.70, 130.10, 129.19, 128.86, 128.69, 127.33, 124.77, 123.86, 123.51, 120.50, 119.25, 108.32, 45.49. HRMS-MALDI (m/z) calcd for $C_{23}H_{15}NO_3S$ $[M + Na]^+$ 408.0671, found 408.0961. HPLC purity: 96.2%.

2-(12-Oxo-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzoic Acid (5u).—According to procedure B, 50 mg of **3u** (0.179 mmol), 28.8 μ L of 2-aminothiophenol (0.269 mmol), 25 mg of *p*-toluenesulfonic acid monohydrate (0.13 mmol), and 10 mL of *i*PrOH were reacted for 19 h at RT. Orange solid, 56.7 mg (82.1%). ^1H NMR (600 MHz, DMSO- d_6): δ 13.11 (s, 1H), 10.09 (s, 1H), 8.12 (d, J = 7.41 Hz, 1H), 7.75 (dd, J = 7.68, 1.32 Hz, 1H), 7.57–7.54 (m, 2H), 7.42 (t, J = 7.33 Hz, 1H), 7.37 (d, J = 6.84 Hz, 1H), 7.30 (td, J = 7.14, 1.58 Hz, 1H), 7.10 (td, J = 7.50, 0.92 Hz, 1H), 6.98 (td, J = 7.68, 1.32 Hz, 1H), 6.88–6.82 (m, 2H), 6.60 (s, 1H), 6.53 (d, J = 7.81 Hz, 1H). ^{13}C NMR (125 MHz, DMSO- d_6): 189.34, 168.90, 157.15, 143.21, 142.69, 139.34, 135.98, 132.71, 131.66, 130.58, 130.38, 130.05, 129.45, 129.06, 126.72, 126.52, 124.64, 124.02, 123.19, 120.45, 119.11, 108.67, 42.85. HRMS-MALDI (m/z) calcd for $\text{C}_{23}\text{H}_{15}\text{NO}_3\text{S}$ [$\text{M} + \text{Na}$] $^+$ 408.0671, found 408.0302. HPLC purity: 97.9%.

11-([1,1'-Biphenyl]-4-yl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5v).—According to procedure B, 50 mg of **3v** (0.16 mmol), 21 μ L of 2-aminothiophenol (0.19 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 43.3 mg (64.4%). ^1H NMR (600 MHz, DMSO- d_6): δ 10.05 (s, 1H), 8.13 (d, J = 7.41 Hz, 1H), 7.59 (dd, J = 8.22, 1.02 Hz, 1H), 7.56 (td, J = 7.56, 1.14 Hz, 1H), 7.54–7.52 (m, 2H), 7.43 (t, J = 7.32 Hz, 1H), 7.39–7.37 (m, 5H), 7.34 (td, J = 7.44, 1.38 Hz, 1H), 7.30 (t, J = 7.36 Hz, 1H), 7.11 (d, J = 8.26 Hz, 2H), 7.03 (dd, J = 7.72, 1.42 Hz, 1H), 6.92 (dd, J = 7.50, 1.12 Hz, 1H), 5.54 (s, 1H). ^{13}C NMR (150 MHz, DMSO- d_6): 189.30, 156.42, 143.39, 141.53, 139.40, 139.30, 137.92, 136.06, 132.66, 131.66, 130.03, 129.06, 128.83, 127.80, 127.30, 126.35, 125.93, 124.66, 124.13, 123.41, 120.43, 119.15, 109.05, 45.43. HRMS-MALDI (m/z) calcd for $\text{C}_{28}\text{H}_{19}\text{NOS}$ [$\text{M} + \text{H}$] $^+$ 418.1259, found 418.1215. HPLC purity: 97.8%.

11-(3-Methoxyphenyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5w).—According to procedure B, 100 mg of **3w** (0.381 mmol), 49 μ L of 2-aminothiophenol (0.53 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Orange solid, 68.00 mg (48.03%). Spectral data are identical to those published in the literature.⁵⁵

5-(4-Methoxyphenyl)-1,5-dihydro-6H-indeno[1,2-e][1,4]thiazepin-6-one (9a).—According to procedure E, 50 mg of **3q** (0.22 mmol), 2-aminoethanethiol (0.332 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Orange solid, 18.1 mg (30.0%). ^1H NMR (500 MHz, DMSO- d_6): δ 8.36 (s, 1H), 7.65 (d, J = 7.30 Hz, 1H), 7.44 (td, J = 7.50, 0.80 Hz, 1H), 7.35 (t, J = 7.25 Hz, 1H), 7.25 (d, J = 6.95 Hz, 1H), 7.20 (d, J = 8.55 Hz, 2H), 6.84 (d, J = 8.72 Hz, 2H), 5.00 (s, 1H), 3.94 (m, 1H), 3.70 (s, 3H), 3.51 (m, 1H), 2.87 (m, 1H), 2.65 (m, 1H). ^{13}C NMR (150 MHz, DMSO- d_6): 189.97, 161.65, 157.70, 139.18, 134.87, 133.72, 131.03, 129.82, 128.64, 119.42, 118.17, 113.49, 106.88, 55.01, 48.31, 40.46, 27.55. HRMS-ESI (m/z) calcd for $\text{C}_{19}\text{H}_{17}\text{NO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$ 324.1053, found 324.1051. HPLC purity: 95.4%.

5-(4-Bromophenyl)-1,5-dihydro-6H-indeno[1,2-e][1,4]thiazepin-6-one (9b).—According to procedure E, 100 mg of **3l** (0.27 mmol), 2-aminoethanethiol (0.48 mmol),

4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 24 h at RT. Orange solid, 61.8 mg (52.0%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.43 (s, 1H), 7.66 (d, *J* = 7.30 Hz, 1H), 7.48–7.44 (m, 3H), 7.36 (t, *J* = 7.35 Hz, 1H), 7.25 (t, *J* = 8.30 Hz, 3H), 5.04 (s, 1H), 3.95–3.90 (m, 1H), 3.57–3.52 (m, 1H), 2.91–2.87 (m, 1H), 2.65–2.59 (m, 1H). ¹³C NMR (125 MHz, DMSO-*d*₆): 190.01, 161.65, 142.77, 139.14, 133.63, 131.12, 131.01, 129.94, 129.80, 119.52, 119.32, 118.33, 105.83, 48.56, 40.64, 27.55. HRMS-ESI (*m/z*) calcd for C₁₈H₁₄BrNOS [M + H]⁺ 372.0051, found 372.0050. HPLC purity: 95.5%.

5-(*p*-Tolyl)-1,5-dihydro-6H-indeno[1,2-*e*][1,4]thiazepin-6-one (9c).—According to procedure E, 68 mg of **3i** (0.27 mmol), 2-aminoethanethiol (0.41 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 24 h at RT. Orange solid, 24.5 mg (29.0%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.36 (s, 1H), 7.65 (d, *J* = 7.28 Hz, 1H), 7.44 (t, *J* = 7.42 Hz, 1H), 7.35 (t, *J* = 7.34 Hz, 1H), 7.25 (d, *J* = 6.94 Hz, 1H), 7.17 (d, *J* = 8.00 Hz, 2H), 7.09 (d, *J* = 7.99 Hz, 2H), 5.00 (s, 1H), 3.95–3.91 (m, 1H), 3.54–3.49 (m, 1H), 2.88–2.84 (m, 1H), 2.68–2.63 (m, 1H), 2.25 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆): 190.00, 161.73, 140.11, 139.19, 135.35, 133.75, 131.04, 129.84, 128.71, 127.49, 119.44, 118.19, 106.69, 48.30, 40.75, 27.63, 20.55. HRMS-MALDI (*m/z*) calcd for C₁₉H₁₇NOS [M + H]⁺ 308.1103, found 308.1102. HPLC purity: 98.5%

4-(6-Oxo-2,3,5,6-tetrahydroindeno[1,2-*e*][1,4]thiazepin-5-yl)benzonitrile (9d).—According to general procedure E, 100 mg of **3o** (0.386 mmol), 44.6 mg of 2-aminoethanethiol (0.579 mmol), 10 mL of *i*PrOH, 4 mL of acetic acid, and molecular sieves were reacted for 24 h at RT. Orange solid, 53.18 mg (43.3%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.48 (br s, 1H), 7.75 (d, *J* = 8.30 Hz, 2H), 7.69 (d, *J* = 7.30 Hz, 1H), 7.46 (t, *J* = 7.76 Hz, 3H), 7.37 (t, *J* = 7.35 Hz, 1H), 7.27 (d, *J* = 6.97 Hz, 1H), 5.16 (s, 1H), 3.96–3.91 (m, 1H), 3.59–3.24 (m, 1H), 2.91–2.87 (m, 1H), 2.60–2.56 (m, 1H). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 190.08, 161.63, 149.21, 139.15, 133.54, 132.20, 131.18, 130.00, 128.52, 119.58, 118.87, 118.43, 109.03, 105.03, 48.87, 41.19, 27.50. HRMS-ESI (*m/z*) calcd for C₁₉H₁₄N₂OS [M + H]⁺ 319.0899, found 319.0900. HPLC purity: 96.6%

11-(Thiophen-2-yl)-7-(trifluoromethyl)-5,11-dihydro-12H-benzo[*b*]indeno[1,2-*e*][1,4]thiazepin-12-one (11a).—According to general procedure F, 150 mg of **3y** (0.624 mmol), 224 mg of 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (0.749 mmol), 15 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Orange solid, 139 mg (53.8%). ¹H NMR (500 MHz, CD₃OD-*d*₄): δ 7.87 (d, *J* = 7.34 Hz, 1H), 7.79 (s, 1H), 7.51 (td, *J* = 7.35, 1.35 Hz, 1H), 7.45–7.40 (m, 2H), 7.37 (d, *J* = 8.08 Hz, 1H), 7.22 (dd, *J* = 8.35, 1.40 Hz, 1H), 7.07 (d, *J* = 5.00 Hz, 1H), 6.65–6.63 (m, 1H), 6.57 (d, *J* = 3.51 Hz, 1H), 5.72 (s, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆): δ 189.16, 155.61, 146.17, 143.79, 138.95, 137.40, 132.14, 132.01, 130.32, 129.29 (q, *J*_{CF} = 32.14 Hz), 126.39, 125.69, 125.60, 123.74 (q, *J*_{CF} = 270.90 Hz), 120.86, 120.54 (q, *J*_{CF} = 3.47 Hz), 119.81 (q, *J*_{CF} = 3.83 Hz), 119.45, 110.69, 41.51. ¹⁹F NMR (CD₃OD-*d*₄) δ –64.37(s). HRMS-MALDI (*m/z*) calcd for C₂₁H₁₂F₃NOS₂ [M + H]⁺ 416.0384, found 415.9460. HPLC purity: 97.6%

11-(4-Bromophenyl)-7-(trifluoromethyl)-5,11-dihydro-12H-benzo[*b*]indeno[1,2-*e*][1,4]thiazepin-12-one (11b).—According to general procedure F, 100 mg of **3l**

(0.319 mmol), 144 mg of 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (0.479 mmol), 10 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Orange solid, 62.1 mg (39.8%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 10.18 (br s, 1H), 8.08 (d, *J* = 7.38 Hz, 1H), 7.98 (s, 1H), 7.59 (td, *J* = 7.40, 1.20 Hz, 1H), 7.45–7.40 (m, 2H), 7.30–7.26 (m, 3H), 7.21 (d, *J* = 8.06 Hz 1H), 7.02 (d, *J* = 8.45 Hz 2H), 5.62 (s, 1H). ¹⁹F NMR δ –63.37 (s). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 189.46, 156.22, 144.14, 141.52, 139.14, 137.07, 132.19, 131.99, 130.82, 130.22, 129.32 (q, *J*_{CF} = 32.13 Hz), 129.29, 128.78, 123.72 (q, *J*_{CF} = 271.25 Hz), 120.79, 120.56 (q, *J*_{CF} = 3.77 Hz), 119.85 (q, *J*_{CF} = 4.33 Hz), 119.75, 119.31, 109.12, 45.21. HRMS-MALDI (*m/z*) calcd for C₂₃H₁₃BrF₃NOS [M + H]⁺ 487.9925, found 487.9705. HPLC purity: 98.5%

11-(Thiophen-2-yl)-5,11-dihydro-12H-benzo[*b*]indeno[1,2-*e*][1,4]thiazepin-12-one (5y).—According to procedure B, 50 mg of **3y** (0.21 mmol), 26.5 μL of 2-aminothiophenol (0.248 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at RT. Red solid, 39.6 mg (55.0%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.05 (s, 1H), 8.10 (d, *J* = 7.38 Hz, 1H), 7.56–7.54 (m, 2H), 7.43 (t, *J* = 7.28 Hz, 1H), 7.39 (d, *J* = 6.50 Hz, 1H), 7.35 (td, *J* = 7.26, 1.56 Hz, 1H), 7.18 (td, *J* = 8.04, 1.44 Hz, 2H), 7.00 (td, *J* = 7.41, 1.17 Hz, 1H), 6.67–7.66 (m, 1H), 6.54–7.53 (m, 1H), 5.67 (s, 1H). ¹³C NMR (150 MHz, DMSO-*d*₆): 188.94, 155.94, 146.53, 143.01, 139.03, 136.36, 132.54, 131.69, 130.15, 129.11, 126.23, 125.35, 125.24, 124.75, 124.24, 123.44, 120.53, 119.32, 110.05, 41.32. HRMS-ESI (*m/z*) calcd for C₂₀H₁₃NOS₂ [M + Na]⁺ 370.0337, found 370.1523. HPLC purity: 98.3%.

11-(3-Methoxyphenyl)-5,11-dihydro-12H-benzo[*b*]indeno[1,2-*e*][1,4]thiazepin-12-one 10,10-Dioxide (7a).—According to general procedure D, to a solution of **5w** (68 mg, 0.18 mmol) in dry dichloromethane (DCM, 8 mL) was added *m*CPBA (107 mg, 0.620 mmol), and the mixture was stirred for 5 h at room temperature. Yellow solid, 45.3 mg (62.2%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.05 (s, 1H), 8.18 (d, *J* = 7.38 Hz, 1H), 7.85 (d, *J* = 8.15 Hz, 1H), 7.73 (td, *J* = 7.38, 1.44 Hz, 1H), 7.64 (td, *J* = 7.26, 1.62 Hz, 1H), 7.50–7.46 (m, 3H), 7.20 (d, *J* = 7.60 Hz, 1H), 7.04 (d, *J* = 7.92 Hz, 1H), 6.74 (dd, *J* = 8.04, 2.04 Hz, 1H), 6.65–6.63 (m, 2H), 5.68 (s, 1H), 3.55 (s, 3H). ¹³C NMR (150 MHz, DMSO-*d*₆): 190.64, 158.79, 155.50, 138.96, 136.41, 135.54, 134.82, 132.36, 132.13, 130.56, 129.15, 129.08, 127.17, 124.28, 123.32, 121.46, 121.03, 120.11, 114.85, 114.10, 100.15, 66.36, 54.96. HRMS-MALDI (*m/z*) calcd for C₂₃H₁₇NO₄S [M + Na]⁺ 426.0776, found 426.0566. HPLC purity: 96.8%.

11-(4-Methoxyphenyl)-7-(trifluoromethyl)-5,11-dihydro-12H-benzo[*b*]indeno[1,2-*e*][1,4]thiazepin-12-one (11c).—According to general procedure F, 150 mg of **3q** (0.378 mmol), 195 mg of 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (0.568 mmol), 15 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Orange solid, 83 mg (33%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.12 (s, 1H), 8.06 (d, *J* = 7.40 Hz, 1H), 7.97 (s, 1H), 7.58 (td, *J* = 7.50, 1.20 Hz, 1H), 7.44–7.39 (m, 2H), 7.25 (dd, *J* = 8.16, 1.53 Hz, 1H), 7.20 (d, *J* = 8.07 Hz, 1H), 6.97 (d, *J* = 8.72 Hz, 2H), 6.65 (d, *J* = 8.80 Hz, 2H), 5.55 (s, 1H), 3.60 (s, 3H). ¹⁹F NMR δ –61.32 (s). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 189.43, 157.78, 155.95, 144.14, 139.22, 137.08, 133.97, 132.26, 131.91, 130.10, 129.31,

129.10 (q, $J_{\text{CF}} = 32.17$ Hz), 128.27, 123.74 (q, $J_{\text{CF}} = 271.13$ Hz), 120.70, 120.36 (q, $J_{\text{CF}} = 3.50$ Hz), 119.66 (q, $J_{\text{CF}} = 3.98$ Hz), 119.18, 113.24, 110.23, 54.91, 45.45. HRMS-MALDI (m/z) calcd for $\text{C}_{24}\text{H}_{16}\text{F}_3\text{NO}_2\text{S}$ $[\text{M} + \text{H}]^+$ 440.0926, found 440.0735. HPLC purity: 98.4%.

11-(p-Tolyl)-10,11-dihydrobenzo[b]indeno[1,2-e][1,4]diazepin-12(5H)-one (13a).

—According to general procedure G, 150 mg of **3i** (0.605 mmol), 15 mL of isopropanol, 98.1 mg of *ortho*-phenylenediamine (0.646 mmol), 3 mL of acetic acid, and molecular sieves were added in a three-neck round-bottom flask and reacted for 24 h at RT under nitrogen gas. Red solid, 54.2 mg (26.4%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 9.83 (s, 1H), 7.98 (d, $J = 7.29$ Hz, 1H), 7.48 (t, $J = 7.45$ Hz, 1H), 7.36–7.34 (m, 2H), 7.31 (d, $J = 6.84$ Hz, 1H), 7.02 (d, $J = 8.09$ Hz, 2H), 6.95 (d, $J = 7.98$ Hz, 2H), 6.98–6.76 (m, 2H), 6.67–6.65 (m, 1H), 6.22 (d, $J = 4.69$ Hz, 1H), 5.28 (d, $J = 4.69$ Hz, 1H), 2.15 (s, 3H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ 188.76, 156.01, 141.38, 138.71, 138.37, 135.29, 134.47, 130.92, 130.22, 128.57, 126.91, 124.37, 122.60, 121.51, 120.26, 119.83, 118.78, 107.78, 107.75, 55.94, 20.51. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{18}\text{N}_2\text{O}$ $[\text{M} + \text{H}]^+$ 339.1492, found 339.1497. HPLC purity: 95.1%.

11-(p-Tolyl)-10,11-dihydrobenzo[b]indeno[1,2-e][1,4]diazepin-12(5H)-one (13b).

—According to general procedure G, 150 mg of **3o** (0.579 mmol), 93.1 mg of *ortho*-phenylenediamine (0.868 mmol), 3 mL of acetic acid, 15 mL of *i*PrOH, and molecular sieves were added in a three-neck round-bottom flask and reacted for 24 h at RT. Red solid, 62.16 mg (37.7%). ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δ 9.99 (br s, 1H), 8.01 (d, $J = 7.32$ Hz, 1H), 7.65 (d, $J = 8.30$ Hz, 2H), 7.50 (t, $J = 7.47$ Hz, 1H), 7.40–7.37 (m, 2H), 7.33 (d, $J = 8.30$ Hz, 3H), 6.84–6.81 (m, 2H), 6.69–6.67 (m, 1H), 6.40 (d, $J = 4.69$ Hz, 1H), 5.38 (d, $J = 4.66$ Hz, 1H). ^{13}C NMR (125 MHz, $\text{DMSO}-d_6$): δ 188.79, 156.32, 149.84, 138.20, 138.06, 134.28, 132.11, 131.10, 130.19, 129.88, 127.95, 124.71, 122.55, 121.81, 120.77, 120.01, 119.09, 118.78, 109.21, 106.20, 56.13. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{15}\text{N}_3\text{O}$ $[\text{M} + \text{H}]^+$ 350.1288, found 350.1291. HPLC purity: 96.5%.

11-(4-Methoxyphenyl)-10,11-dihydrobenzo[b]indeno[1,2-e][1,4]diazepin-12(5H)-one (13c).

—According to general procedure G, 73 mg of **3q** (0.323 mmol), *ortho*-phenylenediamine (69.8 mg, 0.646 mmol), 4 mL of acetic acid, 10 mL of *i*PrOH, and molecular sieves were added in a three-neck round-bottom flask and reacted for 24 h at RT. Red solid, 40.25 mg (41.0%). ^1H NMR (600 MHz, $\text{DMSO}-d_6$): δ 9.83 (s, 1H), 7.97 (d, $J = 7.32$ Hz, 1H), 7.48 (td, $J = 7.56$, 1.02 Hz, 1H), 7.38–7.33 (m, 2H), 7.30 (d, $J = 6.94$ Hz, 1H), 7.05 (d, $J = 8.72$ Hz, 2H), 6.80–6.76 (m, 2H), 6.72–6.69 (m, 2H), 6.68–6.66 (m, 1H), 6.19 (d, $J = 4.28$ Hz, 1H), 5.26 (d, $J = 4.14$ Hz, 1H), 3.63 (s, 3H). ^{13}C NMR (150 MHz, $\text{DMSO}-d_6$): 188.72, 157.65, 155.97, 138.76, 138.34, 136.42, 134.44, 130.91, 130.25, 129.64, 128.05, 124.36, 122.59, 121.50, 120.25, 119.81, 118.75, 113.29, 107.95, 55.60, 54.84. HRMS-ESI (m/z) calcd for $\text{C}_{23}\text{H}_{18}\text{N}_2\text{O}_2$ $[\text{M} + \text{Na}]^+$ 377.1266, found 377.2529. HPLC purity: 95.1%.

11-(o-Tolyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (5x).

—According to procedure B, 50 mg of **3x** (0.201 mmol), 32.2 μL of 2-aminothiophenol (0.302 mmol), 4 mL of acetic acid, and 6 mL of *i*PrOH were reacted for 18 h at 80 °C.

Orange solid, 5.53 mg (7.4%). ¹H NMR (500 MHz, DMSO-*d*₆): δ 10.08 (s, 1H), 8.12 (d, *J* = 7.38 Hz, 1H), 7.60 (d, *J* = 8.08 Hz, 1H), 7.55 (t, *J* = 7.51 Hz, 1H), 7.41 (t, *J* = 7.31 Hz, 1H), 7.36–7.32 (m, 2H), 7.11 (d, *J* = 7.39 Hz, 1H), 6.92 (t, *J* = 7.38 Hz, 1H), 6.87 (d, *J* = 4.24 Hz, 2H), 6.64 (t, *J* = 7.46 Hz, 1H), 6.31 (d, *J* = 7.64 Hz, 1H), 5.47 (s, 1H), 2.54 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆): δ 189.14, 156.82, 143.44, 139.72, 139.40, 136.10, 135.09, 132.68, 131.68, 130.25, 130.02, 129.07, 126.64, 125.71, 124.81, 124.65, 123.86, 123.21, 120.43, 119.11, 109.66, 43.16, 19.16. HRMS-ESI (*m/z*) calcd for C₂₃H₁₇NOS [M + Na]⁺ 378.0929, found 378.3829. HPLC purity: 97.7%.

2-(12-Oxo-7-(trifluoromethyl)-11,12-dihydro-5H-benzo[b]indeno[1,2-e][1,4]thiazepin-11-yl)benzonitrile (11d).—According to general procedure F, 100

mg of **3a** (0.386 mmol), 133 mg of 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (0.579 mmol), 10 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Orange solid, 28.3 mg (17.0%). ¹H NMR (500 MHz, CD₃OD): δ 10.37 (s, 1H), 8.12 (d, *J* = 7.34 Hz, 1H), 8.04 (s, 1H), 7.83 (d, *J* = 7.52 Hz, 1H), 7.62 (td, *J* = 7.50, 1.05 Hz, 1H), 7.48–7.42 (m, 2H), 7.30–7.22 (m, 3H), 7.16 (d, *J* = 8.08 Hz, 1H), 6.71 (d, *J* = 7.72 Hz, 1H), 5.70 (s, 1H). ¹⁹F NMR (CD₃OD) δ –61.45 (s). ¹³C NMR (150 MHz, DMSO-*d*₆): δ 189.36, 156.90, 144.76, 144.29, 139.11, 137.15, 133.65, 132.50, 132.18, 132.12, 130.47, 129.76 (q, *J* = 32.84 Hz), 127.96, 127.72, 126.74, 124.15 (q, *J* = 260.63 Hz), 121.00, 120.86 (q, *J* = 3.66 Hz), 120.22 (q, *J* = 3.59 Hz), 119.63, 117.33, 110.73, 107.53, 44.29. HRMS-ESI (*m/z*) calcd for C₂₄H₁₃F₃N₂OS [M + CH₃OH + H]⁺ 467.1035, found 467.1649. HPLC purity: 99.6%.

11-(2-Methoxyphenyl)-7-(trifluoromethyl)-5,11-dihydro-12H-benzo[b]indeno[1,2-e][1,4]thiazepin-12-one (11e).—According

to general procedure F, 100 mg of **3b** (0.378 mmol), 130 mg of 2-amino-4-(trifluoromethyl) benzenethiol hydrochloride (0.568 mmol), 10 mL of *i*PrOH, and molecular sieves were reacted for 24 h at RT. Orange solid, 85.6 mg (51.5%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.18 (s, 1H), 8.08 (d, *J* = 7.40 Hz, 1H), 8.00 (s, 1H), 7.58 (td, *J* = 7.50, 1.08 Hz, 1H), 7.42 (t, *J* = 7.33 Hz, 1H), 7.38 (d, *J* = 6.90 Hz, 1H), 7.20 (dd, *J* = 8.10, 1.50 Hz, 1H), 7.11 (d, *J* = 8.05 Hz, 1H), 7.06 (td, *J* = 7.74, 1.62 Hz, 1H), 6.96 (d, *J* = 8.25 Hz, 1H), 6.45 (td, *J* = 7.50, 0.78 Hz, 1H), 6.35 (dd, *J* = 7.56, 1.44 Hz, 1H), 5.68 (s, 1H), 3.92 (s, 3H). ¹⁹F NMR (DMSO-*d*₆): δ –61.32 (s). ¹³C NMR (150 MHz, DMSO-*d*₆): δ 189.36, 156.53, 155.76, 143.92, 139.24, 137.10, 132.23, 131.91, 130.09, 129.75, 129.20, 129.06 (q, *J* = 32.16 Hz), 128.34, 126.20, 123.77 (q, *J* = 270.99 Hz), 120.69, 120.28 (q, *J* = 3.40 Hz), 119.59 (q, *J* = 3.74 Hz), 119.17, 119.16, 110.93, 109.78, 55.70, 40.17. HRMS-ESI (*m/z*) calcd for C₂₄H₁₆F₃NO₂S [M + Na]⁺ 462.0762, found 462.4431. HPLC purity: 99.5%.

Quantitative Structure–Activity Relationship (QSAR) Model.—Molecular modeling was performed using Maestro interface¹⁰² (Schrödinger package, version 2020–4). Molecules were modeled from a structure retrieved from the ChemSpider database (ID 2117364) followed by standard molecular mechanics geometry optimization using Maestro standard settings. The experimental antichlamydia IC₅₀ values obtained from the semi-high-throughput imaging inclusion forming units (IFU) screen were converted to the corresponding pIC₅₀ using the pIC₅₀ value calculator from the Sanjeev's Lab¹⁰³ (<https://www.sanjeevslab.org/tools.html>). Only molecules with a determined value of antichlamydia

activity were used to generate the QSAR models. Therefore, molecules with no inhibition seen within the concentration range tested (25–0.39 $\mu\text{g/mL}$) do not compose the dataset used in the QSAR study. The dataset was automatically divided into training (21 compounds—77% of the total dataset) and test set (compounds **5c**, **5d**, **5h**, **5n**, **7a**, **13c**—23% of the total dataset) using autoQSAR tool⁷² from the Schrödinger package, version 2020–4. AutoQSAR, is a machine-learning tool to build, validate, and deploy quantitative structure–activity relationship models in an automated manner.⁷² For the descriptor generation, the following set of binary fingerprints were calculated: radial, linear, dendritic, and molprint2D.¹⁰⁴ For feature selection, the maximum allowed correlation between any pair of independent variables was set to 0.80. The maximum number of generated models was set to 50 and all of the AutoQSAR available methods to generate models were employed.⁷² The model selection was based on the predictive power (Q^2 values) and the internal ranking score from the software (based on the performance).¹⁰⁵ Model visualization and preparation of the color maps was performed using Maestro interface and AutoQSAR tools.

In Silico Drug-Likeness Properties.—The entire library of dihydrothiazepines (including the molecules with no defined value for the antichlamydial IC_{50}) were modeled according to the description in the QSAR section above. Such dataset was used to calculate drug-likeness properties using standard settings of Qikprop tool¹⁰⁶ (Schrödinger package, version 2020–4). Predictions are related to drug absorption as oral dosage form.

Quantum Mechanics Molecular Electrostatic Potential Surface.—All calculations were performed using Jaguar software¹⁰⁷ incorporated in the Schrödinger package, version 2020–4 and using the computer resources of the Holland Computing Center from the University of Nebraska—Lincoln.¹⁰⁸ The selected molecules (**5q**, **5y**, **5l**, **11a**, **11b**, **11c**) were modeled according to the description in the QSAR section. Tryptophan, tyrosine, and phenylalanine were modeled using Maestro build tool by direct insertion of these amino acids from the internal software database and optimization using the same protocol as described for dihydrothiazepines. Next, the molecules were optimized using density functional theory (DFT) level using the functional Becke, three-parameter, Lee–Yang–Parr (B3LYP), with the basis 6–31G(d,p). Structures were optimized using water as the solvent (Poisson–Boltzmann continuum solvation as model) and the accuracy of the calculations was set to accurate. All of the other parameters were used as the standard settings of Jaguar. The calculated quantum mechanics molecular electrostatic potential (MEP) surfaces were displayed using Maestro interface¹⁰² (Schrödinger package, version 2020–4).

Biology.

Antichlamydial Semi-High-Throughput Imaging Screen. HEp-2 Cell Culture.—

Human epithelial cell line HEp-2 was routinely cultured in Dulbecco's modified Eagle's medium (DMEM; Gibco ref 11965–092) supplemented with 10% fetal bovine serum (FBS) and 10 $\mu\text{g/mL}$ gentamicin at 37 °C in a humidified environment with 5% CO_2 . The same culture conditions were used for both infected and uninfected cell culture samples.

Semi-High-Throughput Imaging Inclusion Forming Units (IFU) Screen.—

Antichlamydial activity can be analyzed by the effect of the drugs on the number of

inclusions using an immunofluorescence assay (IFA). Therefore, HEP-2 cells were first seeded in a 96-well plate (PerkinElmer Cell carrier plate, black well walls, and optically clear bottom) compatible with the Operetta microscope. After incubation for 24 h at 37 °C in a humidified environment with 5% CO₂, cells were infected with *C. trachomatis* serovar L2 at a multiplicity of infection (MOI) of 0.6, and the test molecules were added at seven concentration points: 25, 12.5, 6.25, 3.12, 1.56, 0.78, and 0.39 µg/mL. The drug solutions were prepared by serial dilution in a separated 96-well plate and carefully transferred to the cell plate after 8 h post-infection (hpi). After 16 h (total incubation of the cells for 24 h), the cells were washed with 100 µL of phosphate-buffered saline/azide (PBS-azide—0.025%) and fixed with methanol (100 µL) for 10 min. Next, the samples were stained using primary goat anti-MOMP antibody (Meridian Biosciences, ref B65266G) and a secondary donkey anti-goat antibody labeled with Alexa488 (Invitrogen, ref A32814). Finally, the samples were stained with 4',6-diamidino-2-phenylindole (DAPI). All staining solutions were used in 1:1000 concentration diluted in PBS. The green-fluorescent inclusions (number) were automatically identified, counted, and compared against the vehicle DMSO control (untreated) and positive control azithromycin (1 µg/mL) using the microplate imager Operetta High Content Analysis System (PerkinElmer, Inc.). The following parameters were considered by the system to identify the inclusions: green fluorescence intensity (>600), area (>40 µm²), and roundness (>0.85). All of the samples were tested in triplicate. The IC₅₀ was determined using GraphPad Prism software, version 9.3.1 for Windows.¹⁰⁹

EB Progeny Assay.—Aiming to further evaluate the effect of the most active compounds against *C. trachomatis* L2 (Ctr L2), HEP-2 cells were seeded in a 24-well plate and infected with Ctr L2 at a multiplicity of infection of 1. The samples were treated with drugs in triplicate at 8 hpi. Next, after 16 h (total incubation of the cells for 24 h), the cells were lysed to release *C. trachomatis* elementary bodies, which were collected in sucrose storage medium (2SP) and stored at −80 °C. Next, the collected cell lysates were used to infect a fresh HEP-2 cell monolayer in a series of 10-fold dilutions. Samples were incubated for 24 h at 37 °C in a humidified environment with 5% CO₂ in media containing cycloheximide (2 µg/mL). After 24 hpi, the cells were fixed with methanol for 15 min and the recoverable EBs from the initial infection were stained using primary goat anti-MOMP antibody (Meridian Biosciences, ref B65266G) and a secondary donkey anti-goat antibody labeled with Alexa488 (Invitrogen, ref A32814). The fluorescent inclusions were counted from 15 fields of view (FOV) at 20× magnification and images were acquired using an Olympus CKX53 fluorescence microscope equipped with an EP50 camera. Channel merge was processed using ImageJ software. Images were assembled with Adobe Photoshop 26.0.2 (Adobe Systems Inc., San Jose, CA). The experiment was performed in triplicate. The IC₅₀ was determined using GraphPad Prism software, version 9.3.1 for Windows.¹⁰⁹ Infectious progeny was calculated using the same procedure as described before for a volume of 0.25 mL (volume used per well).^{52,110}

Cytotoxicity Assay. Cell Culture.—Epithelial endometrium adenocarcinoma cells (HEC-1-A) were cultured in McCoy's 5A media (Gibco, ref 16600082) containing 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin (Gibco, 15140–122) at 37 °C in a humidified environment with 5% CO₂. Human embryonic kidney 293 cells (HEK-293)

were cultured in DMEM media (Gibco, ref 11965–092) containing 10% fetal bovine serum (FBS), 1% penicillin/streptomycin (Gibco, 15140–122), and 200 mmol L-glutamine at 37 °C in a humidified environment with 5% CO₂. HEp-2 well cultured as described above.

XTT Assay.—The cells were seeded at a density of 5×10^3 cells per well in 96-well plates. After 24 h, the cells were treated within a concentration range from 256 to 2 µg/mL (in triplicate). As control, DMSO was employed at a concentration equivalent to the ones used in drug-treated cells. Next, the plates were incubated for an additional 20 h. After the incubation, the wells were washed (2×) with 100 µL of PBS to remove any residual compound precipitation and 100 µL of new media was added to all samples. Next, 70 µL of the XTT assay reagent (CyQUANT XTT Cell Viability Assay, Invitrogen, ref X12223) was added followed by incubation of the plates for an additional 4 h (total 24 h), accordingly to the manufacturer's instructions. Corrected absorbance readings were measured with a 450 nm filter using a multiscan FC microplate photometer (Thermo Fisher Scientific).

Antibacterial Assay against Clinically Relevant Bacterial Strains. Minimum Inhibitory Concentration (MIC) Determination.—For both tested compounds and controls drugs, the MIC was determined using the broth microdilution method, according to the Clinical and Laboratory Standards Institute (CLSI) reference M07-A9.¹¹¹ The compounds were serially diluted in Mueller–Hinton broth (MHB, Difco, ref 275730) at a concentration range of 125–1.9 µg/mL in 96-well microtiter plates. *S. aureus* USA 300 JE2, *A. baumannii* 2208, *E. coli* K12, and *P. aeruginosa* PA01 were cultivated in trypticase soy agar (TSA—Difco, ref 236940) at 37 °C for 18 h and standardized to 1.5×10^8 colony forming units (CFU)/mL (0.5 McFarland) on saline (0.85%). Afterward, the bacteria solutions were diluted to 1.5×10^6 CFU/mL in MHB and added to the microtiter plates containing 100 µL of drug media (final bacterial concentration: 1.5×10^5 CFU/mL). The plates were incubated for 24 h at 37 °C. A 0.1% triphenyl tetrazolium chloride (TTC) solution was added to all wells before reading the results. The MIC was defined as the lowest drug concentration that showed no visible bacterial growth. The experiment was performed in triplicate in at least two different moments.

Antibacterial Assay against *L. rhamnosus*. Minimum Inhibitory Concentration (MIC) Determination.—*L. rhamnosus* was isolated from the commercially available probiotic product INESSA ADVANCED DAILY BIOTIC (INESSA Wellness, London W1W 7FA, U.K.). For this, the bacterium was aseptically isolated from the respective product on De Man, Rogosa, and Sharpe (MRS) broth (HiMedia, Mumbai, India), incubated at 37 °C for 72 h with 5% CO₂. Next, an isolate typical colony of *L. rhamnosus* was grown in MRS agar (Weber Scientific, NJ). The compounds were serially diluted in MRS broth at a concentration range of 125–1.9 µg/mL in 96-well microtiter plates. *L. rhamnosus* were cultivated in MRS agar at 37 °C with 5% CO₂ for 24 h and standardized to 1.5×10^8 colony forming units (CFU)/mL (0.5 McFarland) on saline (0.85%). Afterward, the bacterial solution was diluted to 1.5×10^6 CFU/mL in MRS broth and added to the microtiter plates containing 100 µL of drug media (final bacterial concentration: 1.5×10^5 CFU/mL). The plates were incubated for 24 h at 37 °C, with 5% CO₂. A 0.1% triphenyl tetrazolium chloride (TTC) solution was added to all wells of the plates to read the results. The MIC

was defined as the lowest drug concentration that showed no visible bacterial growth. The experiment was performed in triplicate in at least two different moments.

Antibacterial Assay against Anaerobic Bacteria. Minimum Inhibitory

Concentration (MIC) Determination.—Drugs were serially diluted in Brucella broth (BRU, Anaerobe systems, ref AS-105) at a concentration range of 125–0.06 $\mu\text{g/mL}$ in 96-well microtiter plates. *B. producta* DSM 2950, *C. ramosum* DSM 1402, *B. thetaiotaomicron* DSM 2079, and *A. caccae* DSM 14662 were cultivated in brain heart infusion (BHI) agar (Anaerobe systems, ref AS-6426) at 37 °C for 24 h followed by another round of growing in Brucella broth (Anaerobe systems, ref AS-105). From the Brucella broth, a sample was centrifuged at 5000 rpm for 5 min. Next, media was replaced by saline (0.85%) and the sample was standardized to 1.5×10^8 CFU/mL (0.5 McFarland). Afterward, bacterial solutions were diluted to 1.5×10^7 CFU/mL in Brucella broth and added to the microtiter plates containing 100 μL of drug media (final bacterial concentration: 1.5×10^6 CFU/mL). The plates were incubated for 48 h at 37 °C under anaerobic conditions using an anaerobic laminar flow workstation. The environment inside the workstation was routinely checked for maintenance of anaerobiosis using anaerobic strip indicators (Thermo Scientific, ref BR0055B). The MIC was defined as the lowest drug concentration that showed no visible bacterial growth. The experiment was performed in triplicate. *L. plantarum* DSM 20174 was cultivated in MRS agar (Difco, ref 288130) at 30 °C for 24 h followed by another round of growing in MRS broth (HiMedia, ref M369) at 30 °C for 24 h. From this point, the same protocol described above was followed, except using MRS broth for bacterial cultivation/compounds dilution and incubation at 30 °C for 24 h.

Antifungal Susceptibility Testing. Minimum Inhibitory Concentration (MIC)

Determination.—For both the tested compound and fluconazole drug control, the MIC was determined using the broth microdilution method, according to the Clinical and Laboratory Standards Institute (CLSI) reference M27-A3.¹¹² Drugs were serially diluted in Roswell Park Memorial Institute 1640 (RPMI 1640—Gibco, ref 11875093) at a concentration range of 1000–1.95 $\mu\text{g/mL}$ in 96-well microtiter plates. *C. albicans* (90028), *C. glabrata* (2001), *C. Krusei* (6558), and *C. tropicalis* (750) (American Type Culture Collection—ATCC) were suspended in sterile saline (0.9%) and adjusted to a concentration of 2.5×10^6 UFC/mL using a spectrophotometer (Quimis, Brazil) with 90% (± 2) transmittance and 530 nm wavelength. Next, the microorganism suspension was diluted 1:20 and 1:50 in RPMI 1640 medium to obtain $(2-3) \times 10^3$ UFC/mL. Drugs were serially diluted in RPMI 1640 medium at a concentration range of 1000–1.9 $\mu\text{g/mL}$ in 96-well microtiter plates. Next, 100 μL of the microorganism suspension was added to the microtiter plates containing 100 μL of drug media. The MIC was defined as the lowest drug concentration that showed no visible microorganism growth. The experiment was performed in triplicate.

Stability Studies. Human Serum Stability.—The studied compound (2 mg/mL) was incubated in human serum AB (50% v/v—Vally Biomedical Inc.) over 24 h in a shaking incubator (37 °C and 75 rpm). Next, 50 μL aliquots were taken at selected time intervals of 0, 30, 60, 90, 120, 150, 180, and 1440 min. To the aliquots was added 50 μL of HPLC-grade

acetonitrile and the solutions were heated to 80 °C for 1 min followed by centrifugation at 12,000 rpm (Eppendorf Centrifuge) for 5 min. Next, a 50 μ L sample of the resulting supernatant was taken from each time point sample and mixed with 50 μ L of 5-methoxy indole (1 mg/mL—standard control) in acetonitrile solution and centrifuged again. An aliquot of the final supernatant (70 μ L) was removed for analysis using HPLC. The stability results were expressed as the percentage of drug remaining *versus* time. The calculations were performed as percentage parent drug remaining at select time points relative to 0 min (100% parent drug).

Simulated Gastric Fluid Stability.—The studied drug (2 mg/mL) was incubated in simulated gastric fluid—pH 1.4 (Ricca Chemical Company, ref 7108–16) containing 3.2 mg/mL of pepsin (MP biomedical LLC, ref 195367) over 24 h in a shaking incubator (37 °C and 75 rpm). At this point, the same experimental procedure described above for serum stability was followed.

Simulated Intestinal Fluid Stability.—The studied drug (2 mg/mL) was incubated in simulated Intestinal fluid—pH 7.40 (Ricca Chemical Company, ref 7109.75–16) containing 10 mg/mL of pancreatin (MP biomedical LLC, ref 102557) over 24 h in a shaking incubator (37 °C and 75 rpm). At this point, the same experimental procedure described above for serum stability was followed.

ClpX ATPase Inhibition Assay.—CT ClpX containing a C-terminal 6x-HIS tag was purified from *E. coli* BL21(DE3) PAX as previously described by Wood et al.³⁹ using cobalt-based immobilized metal affinity chromatography. Protein purity was assessed by running 1 μ g samples on 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gels following by staining with Coomassie Brilliant Blue. CT ClpX sourced from three independent purifications was used for ATPase assays. The assays were performed using Kinase-Glo (ATP) from Promega with 1 μ M ATP and 1.5 μ g of CT ClpX.³⁹ The reactions were incubated at 30 °C for 60 min and terminated *via* addition of the Kinase-Glo reagent. Luminescence (ATP not consumed by ClpX) was measured on a BioTek Synergy HT plate reader. Data are reported as the amount of ATP depleted *versus* an ATP only control. Experiments were run in at least triplicate with replicates used for each individual experiment.

Mutagenic Potential Determination. Ames Test.—The mutagenic potential of the studied molecule was determined by the Ames test using a microsuspension method described by Kado et al.¹¹³ and according to the OCDE guidelines, test no. 471.¹¹⁴ The assays were conducted using *S. typhimurium* strains TA98 and TA100 (Moltox, Molecular Toxicology, Inc.) in the presence and absence of metabolic activation from the microsomal fraction. To prepare the inoculum, a bacterial suspension was incubated in nutrient broth no. 2 at 37 °C for 16 h. Next, this suspension was concentrated by centrifugation (10,000g/10 min, 4 °C) to obtain a bacterial density of $(1-2) \times 10^9$ UFC/mL in PBS 0.2 mM (pH 7.4). In the assay without metabolic activation, 5 μ L of the studied compound diluted in DMSO was mixture with 50 μ L of bacterial inoculum and 50 μ L of PBS 0.2 mM (pH 7.4) to obtain different drug concentrations (50, 150, 500, 1500, and 5000 μ g/mL). For the assay with

metabolic activation, 50 μL of microsomal fraction S9 was added instead of PBS. Next, the samples were preincubated for 90 min at 37 °C. Next, 2 mL of top agar (sodium chloride 0.6% (m/v); agar-agar 0.6% (m/v); 0.5 mM L-histidine, D-biotin 0.5% (w/v)) was added to the prepared samples, homogenized, and poured onto Petri dishes containing minimal glucose agar (MGA): agar-agar 1.5% (m/v); glucose solution 10% (v/v); glucose dextrose (20% (w/v); 0.02% (v/v) of a Vogel–Boner solution 50 \times ; magnesium sulfate heptahydrate 0.02% (v/v); citric acid monohydrate 0.2% (v/v); potassium phosphate dibasic 1% (m/v); and sodium phosphate 0.35% (v/v)). Following 66 h of incubation at 37 °C, the number of revertant colonies per plate (his⁺) was determined. The experiment was performed in triplicate. The positive controls used in the assays without metabolic activation were 4-nitrophenylenediamine (NPD, 10 $\mu\text{g}/\text{plate}$) for TA98 strain and sodium azide (2.5 $\mu\text{g}/\text{plate}$) for TA100 strain. In the assays with metabolic activation, 2-aminoanthracene (2-AA, 0.625 $\mu\text{g}/\text{plate}$) was used for TA98 and TA100 strains. Negative controls were performed with the sample solvent, DMSO (5 $\mu\text{L}/\text{plate}$).

The results were analyzed using the Salanal Statistical Software (U.S. Environmental Protection Agency, Monitoring Systems Laboratory, version 1.0, from the Research Triangle Institute, RTP), adopting the model by Bernstein et al.¹¹⁵ The mutagenicity index (MI) was calculated according to the formula: MI = number of induced revertants/number of spontaneous revertants. The sample was considered to have mutagenic potential when the mutagenicity index was ≥ 2 in at least one of the concentrations tested and when there was a dose–response relationship between the concentrations tested and the number of induced revertants.⁹⁴ Concentrations that had an MI of less than 0.7 were considered cytotoxic.¹¹⁶

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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ABBREVIATIONS USED

ACP	activators of self-compartmentalizing proteases
AD	applicability domain of the model
ADEP	acyldepsipeptide
AZM	azithromycin
CT	Chlamydia trachomatis

DMSO	dimethyl sulfoxide
Doxy	doxycycline
EB	elementary body
HEC-1-A	epithelial endometrium adenocarcinoma cell line
HEK-293	immortalized human embryonic kidney cell line
HEp-2	epithelial type 2 cell line
IFA	immunofluorescence assay
KPLS	kernel-based partial least-square regression method
MRSA	methicillin-resistant <i>Staphylococcus aureus</i>
OECD	organization for economic cooperation and development
PID	pelvic inflammatory disease
Q^2	coefficient of determination between the observed and predicted biological activities for the test set
QPlogPo/w	predicted octanol/water partition coefficient
QPPCaco	predicted apparent Caco-2 cell permeability
QPPMDC	predicted apparent Madin–Darby Canine Kidney (MDCK) cell permeability
R^2	correlation coefficient
RB	reticulate body
RMSE	root mean square error
SAR	structure–activity relationships
SD	standard deviation of the regression

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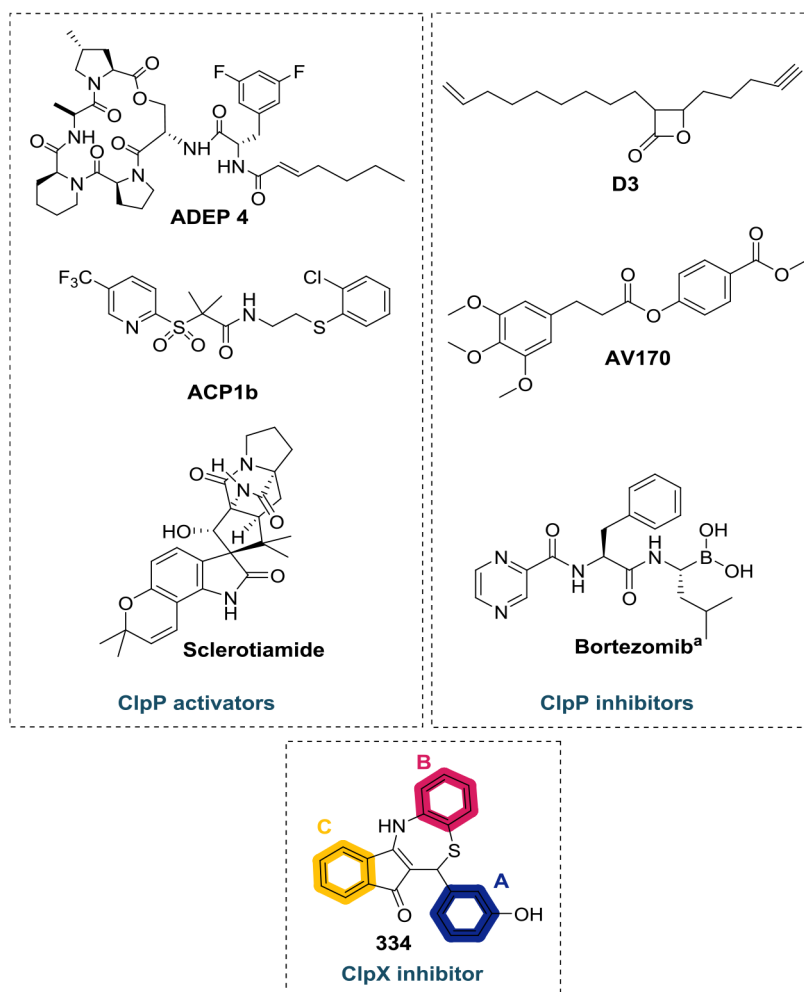


Figure 1. Examples of structures that target the ClpXP protease system. The rings **A**, **B**, and **C** of **334** are color coded as indicated above. (^aBortezomib can exert an activating effect on ClpP depending on its concentration.⁵⁴).

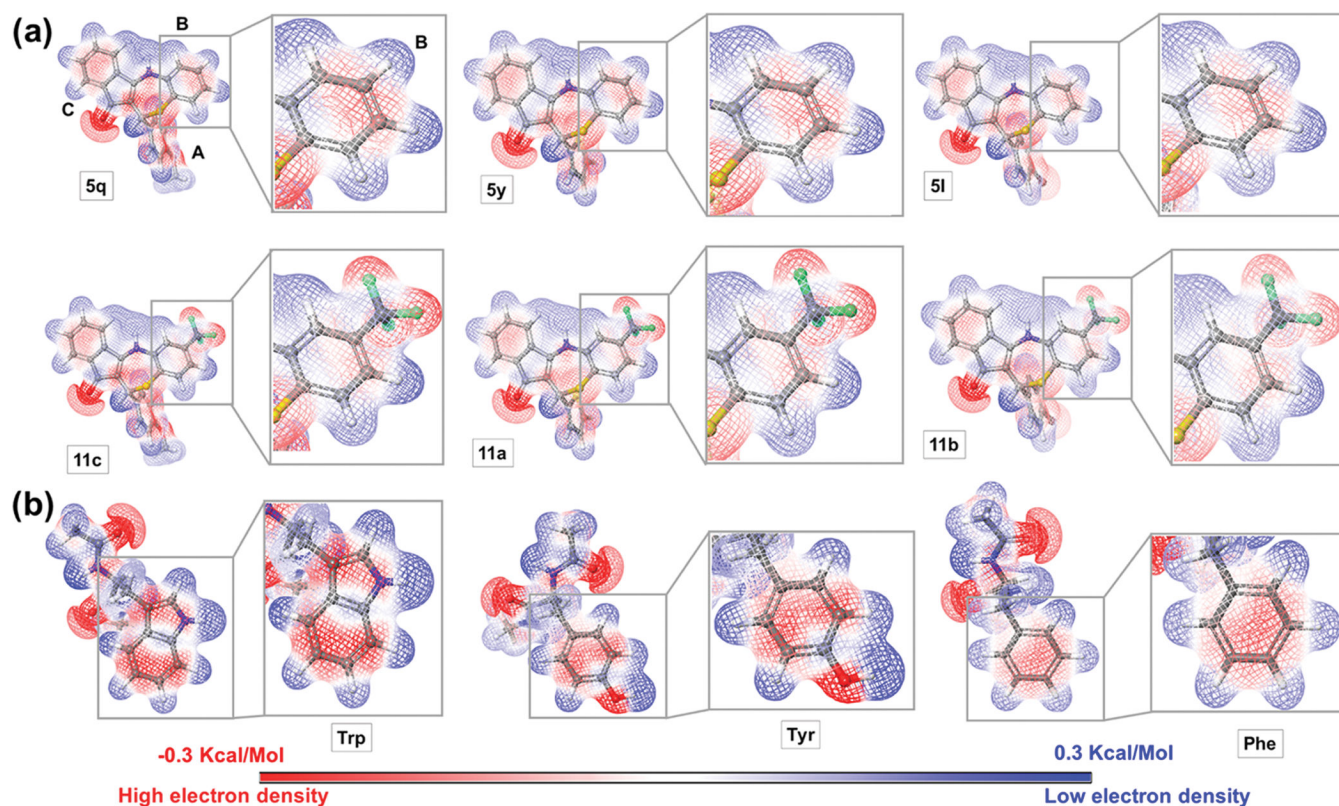
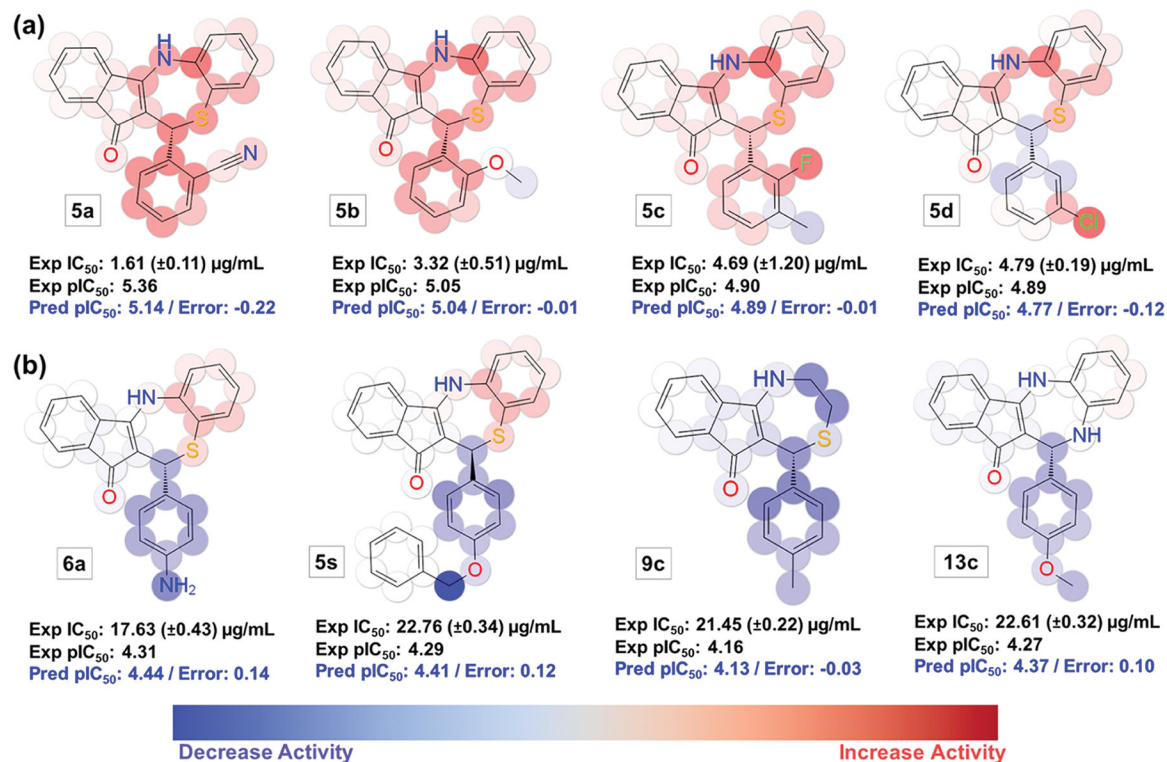
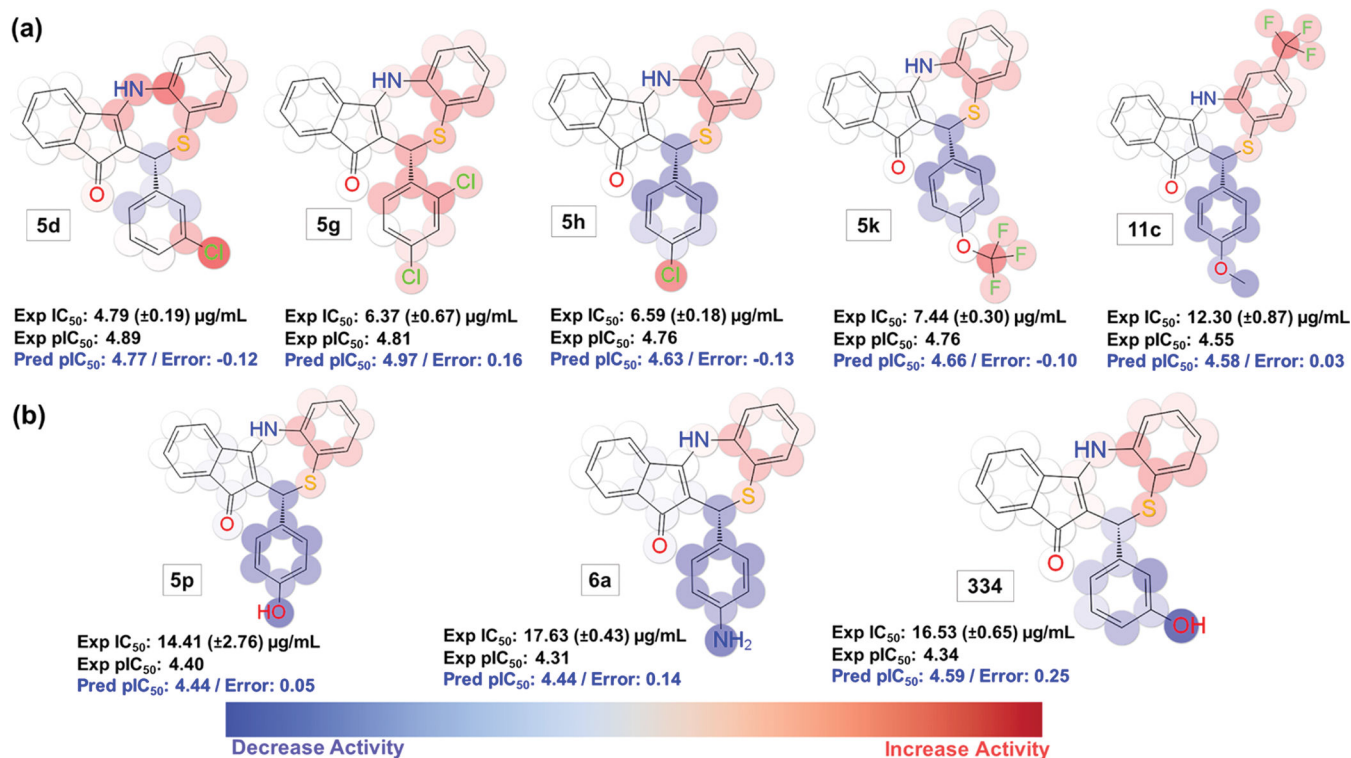


Figure 2.

Molecular electrostatic potential (MEP) surface. (a) Top: compounds with a H atom on ring B. Bottom: compounds containing the trifluoromethyl on ring B. The rest of the molecules possess the same substitution pattern. (b) Tryptophan, tyrosine, and phenylalanine. Detailing of the MEP surface of ring B and the aromatic side chain for Trp, Tyr, and Phe. The most positive potential regions are displayed in deepest blue color and the most negative potential regions in deepest red color.

**Figure 3.**

Color maps for the 2D fingerprint (radial 13) QSAR model. (a) Most active molecules. (b) Molecules with less antichlamydial activity. Atoms displaying a favorable contribution to the antichlamydial activity are colored red and unfavorable are colored blue. Color intensity reflects the strength of the effect. Highly active molecules are shown in predominant red color and less active molecules are presented in predominant blue. (Note: stereochemistry is displayed because the molecules are drawn in the 3D form, which is required for Maestro software to display the QSAR color maps. However, stereochemistry is not considered in the generation of the 2D descriptors used to build the model).

**Figure 4.**

Color maps for the 2D fingerprint (radial 13) QSAR model. (a) Representative molecules of the favorable effect of lipophilic groups on the antichlamydial activity. (b) Representative molecules of the unfavorable effect of H bond donors at ring A on the antichlamydial activity. Atoms displaying a favorable contribution to the antichlamydial activity are colored red and unfavorable are colored blue. Color intensity reflects the strength of the effect. (Note: stereochemistry is displayed because the molecules are drawn in the 3D form, which is required for Maestro software to display the QSAR color maps. However, stereochemistry is not considered in the generation of the 2D descriptors used to build the model).

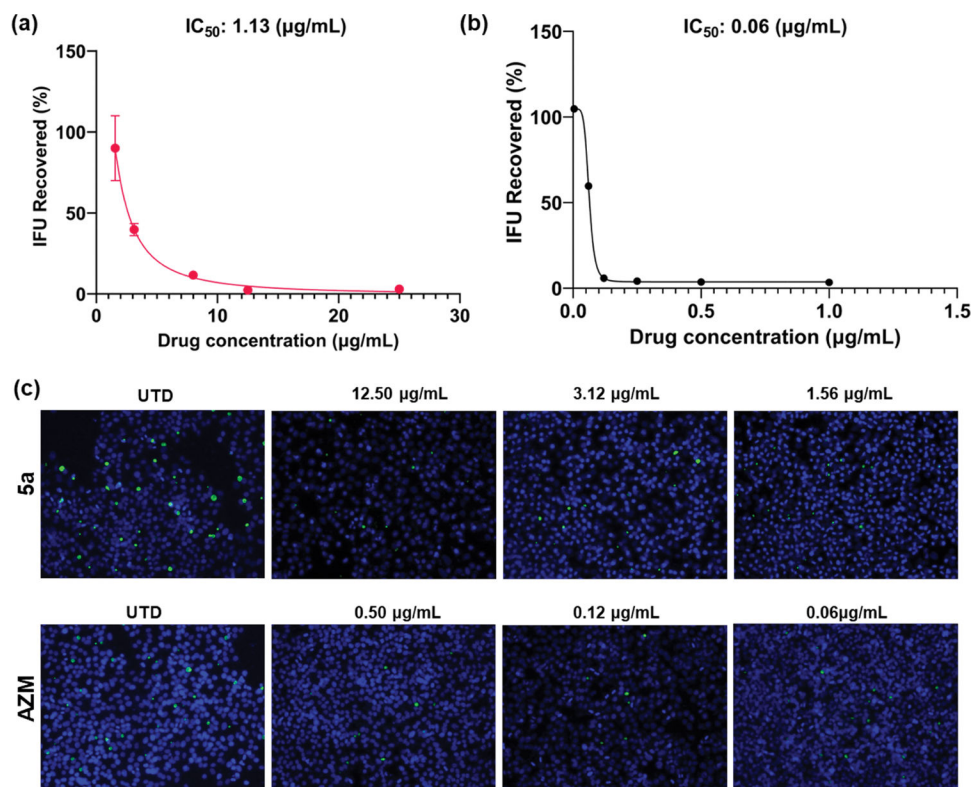


Figure 5. Dose–response curves for the effect of **5a** (a) and AZM (b) reported as the percentage of recovered IFU. Error bars report standard error of the mean (SEM) ($N=3$). (c) Immunofluorescence images for the inhibitory effect of 5a and AZM. Chlamydial inclusions are displayed in green (MOMP - Ctr L2) and HEp-2 cell nuclei in blue (4',6-diamidino-2-phenylindole (DAPI)). Images were acquired on 20× magnification using an Olympus CKX53 fluorescence microscope with EP50 camera. Channel merge was processed using ImageJ software. UTD: untreated; AZM: azithromycin. Omitted error bars indicate a SD value shorter than the size of the symbol presented in the plots.

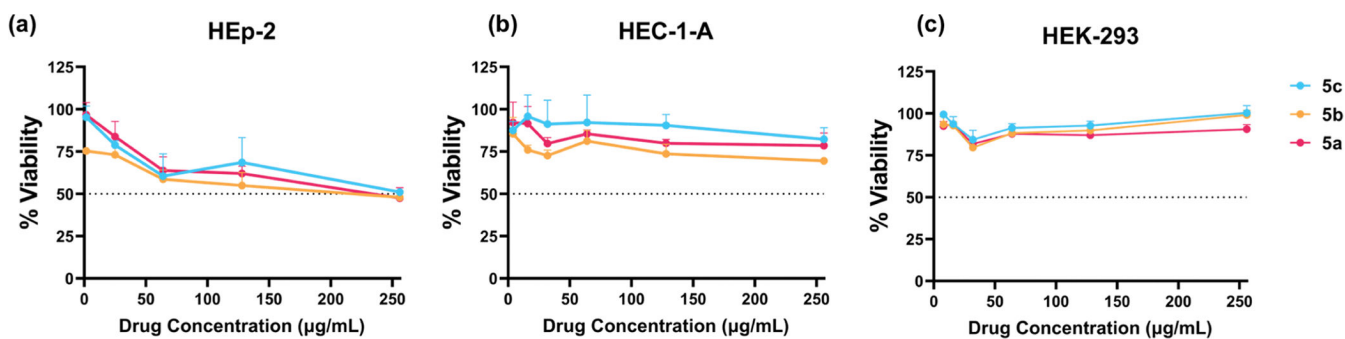


Figure 6.

Viability analysis of the most active compounds against HEP-2 (a), HEC-1-A (b), and HEK-293 (c) cell lines. Error bars report SEM ($N=3$). Each experiment included technical replicates. Omitted error bars indicate an SD value shorter than the size of the symbol presented in the plots.

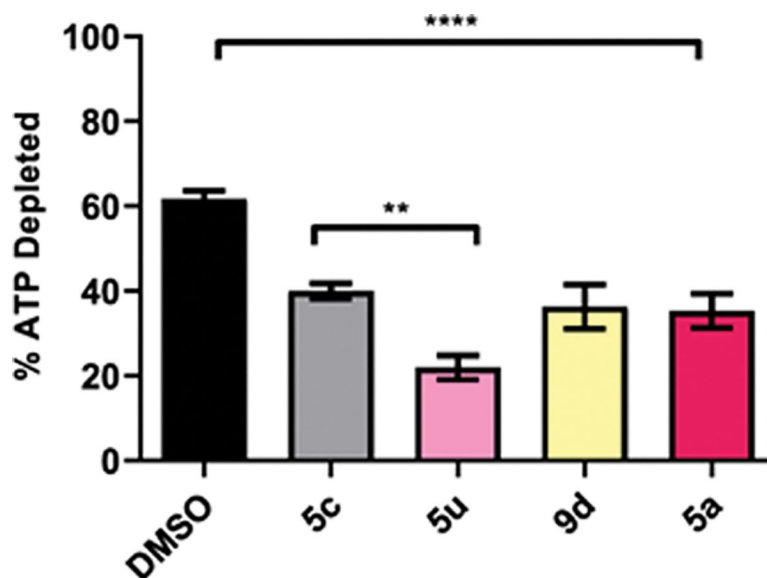


Figure 7.

Inhibition of ClpX ATPase activity. Recombinant ClpX was incubated with either the inhibitor or DMSO as a solvent control. Reactions were halted by the addition of Kinase-Glo at 60 min and the remaining levels of ATP were determined by measuring luminescence as a marker for ClpX ATPase activity. The columns represent the amount of ATP depleted over time *versus* the ATP only control. Error bars report SEM. Each experiment included technical replicates and at least three biological replicates. ** $P < 0.01$; **** $P < 0.0001$; by one-way analysis of variance (ANOVA), Tukey correction.

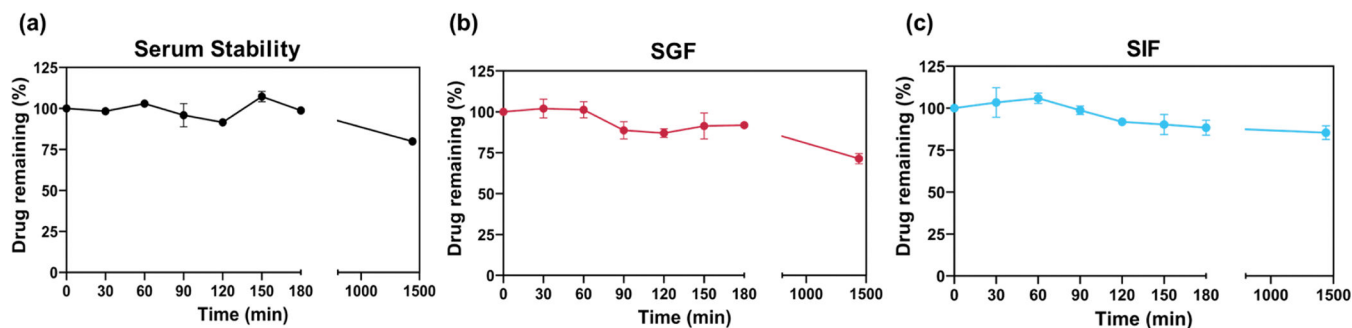
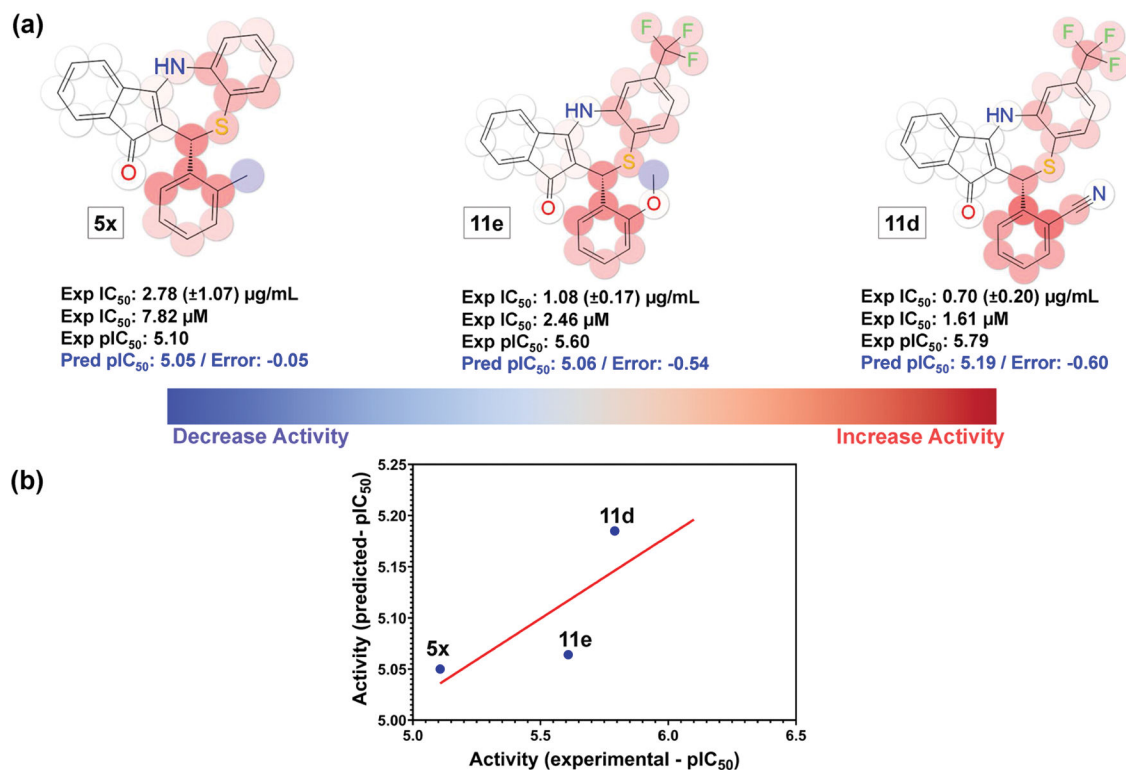
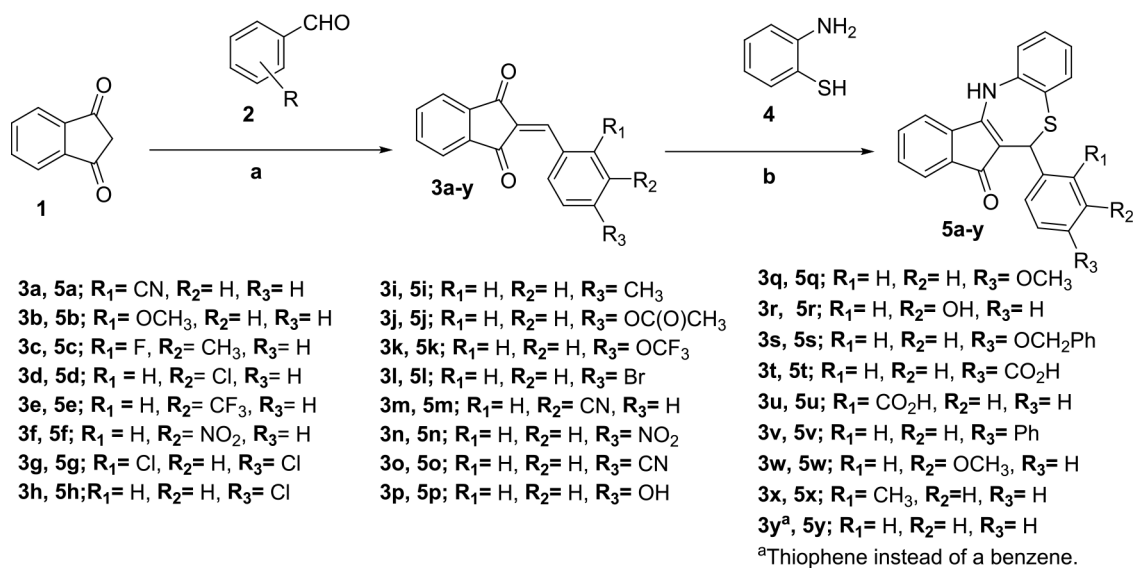


Figure 8.

Stability profile of **5a** in human serum (a), simulated gastric fluid (b), and simulated intestinal fluid (c). The studied compound (2 mg/mL) was incubated in human serum; simulated gastric fluid–pH 1.4 containing 3.2 mg/mL of pepsin; or simulated intestinal fluid–pH 7.40 containing 10 mg/mL of pancreatin over 24 h in a shaking incubator (37 °C and 75 rpm). Relative percentage of drug remaining is presented as a function of incubation time. Error bars represent SD of three independent experiments. Omitted error bars indicate an SD value shorter than the size of the symbol presented in the plots.

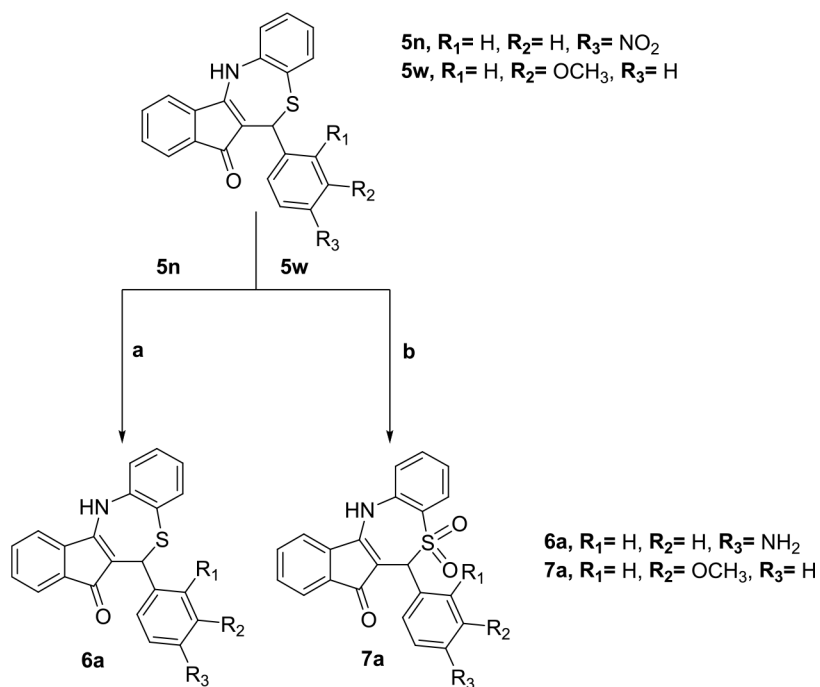
**Figure 9.**

(a) Predictions color maps for the 2D fingerprint (radial 13) QSAR for molecules **11d**, **11e**, and **5x**. (b) Experimental *versus* KPLS-radial 2D QSAR model (radial 13) predicted values of antichlamydial activity (pIC₅₀). Atoms displaying a favorable contribution to the antichlamydial activity are colored red and unfavorable are colored blue. The color intensity reflects the strength of the effect. (Note: stereochemistry is displayed because the molecules are drawn in the 3D form, which is required for Maestro software to display the QSAR color maps. However, stereochemistry is not considered in the generation of the 2D descriptors used to build the model).



Scheme 1. Synthesis of Dihydrothiazepines Containing Modifications on Ring A^a

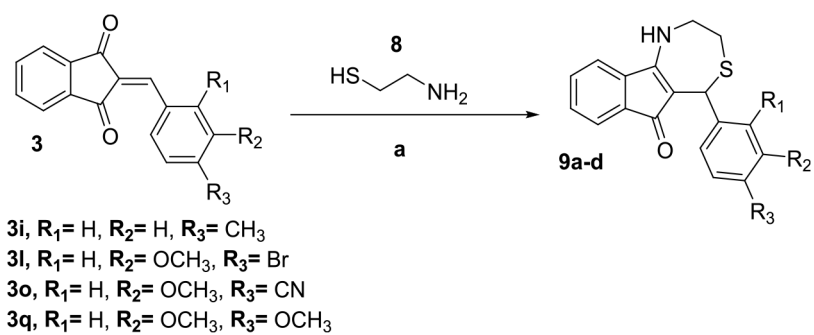
^a(a) L-Proline, MeOH, room temperature (RT), molecular sieve (MS) 4 Å, 16–20 h; (b) isopropanol (*i*PrOH)/HOAc or *p*-toluenesulfonic acid (*p*TsOH)/*i*PrOH, RT, MS 4 Å, 18–24 h.



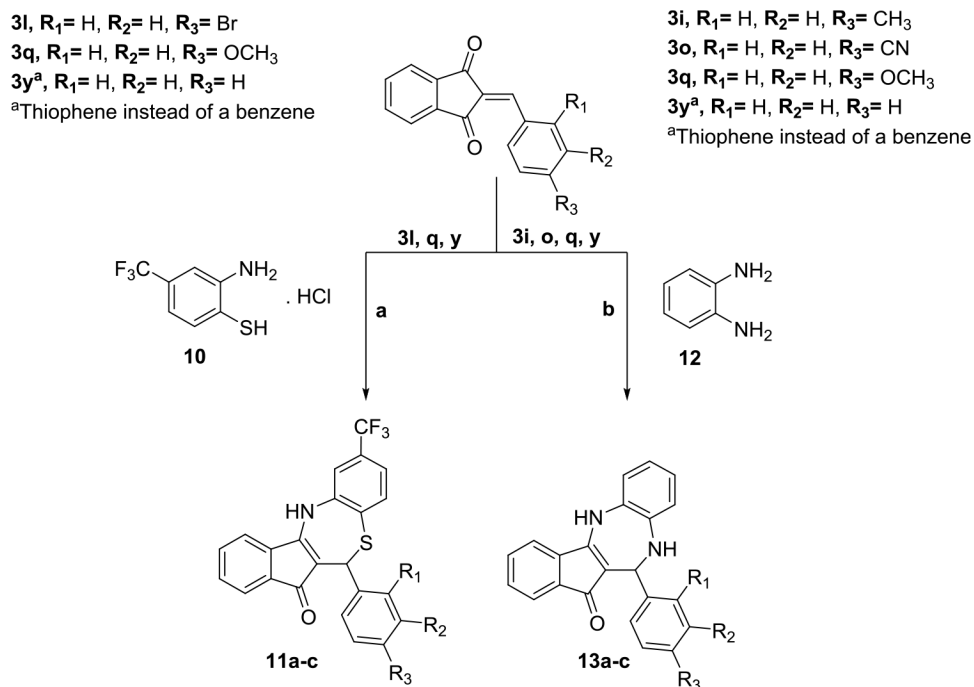
^a(a) Pd/C (10%), H₂(g), tetrahydrofuran (THF)/MeOH, RT, 24 h; (b) *m*CPBA, THF, RT, 5 h.

Scheme 2. Synthesis of Dihydrothiazepines Containing Amine and Sulfone Groups^a

^a (a) Pd/C (10%), H₂(g), tetrahydrofuran (THF)/MeOH, RT, 24 h; (b) *m*CPBA, THF, RT, 5 h.

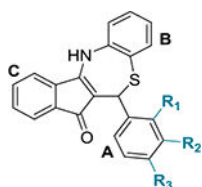
**Scheme 3. Synthesis of Dihydrothiazepines without Ring B^a**

^a (a) *i*PrOH/HOAc or *p*TsOH/*i*PrOH, RT, MS 4 Å, 18–24 h



Scheme 4. Synthesis of Dihydrothiazepines Containing Modification on Ring B and Diazepine Derivatives^a

^a (a) *i*PrOH, RT, MS 4 Å, 18–24 h; (b) *i*PrOH/HOAc or *p*TsOH/*i*PrOH, N₂(g), RT, MS 4 Å, 18–24 h.

Table 1.Chemical Structures of Dihydrothiazepine Derivatives 5a–v and Their IC₅₀ against *C. trachomatis* Serovar L2^a

compound	R ₁	R ₂	R ₃	IC ₅₀ (μg/mL) ^{b,c}
5a	CN	H	H	1.61 ± 0.11
5b	OCH ₃	H	H	3.32 ± 0.51
5c	F	CH ₃	H	4.69 ± 1.20
5d	H	Cl	H	4.79 ± 0.19
5e	H	CF ₃	H	5.18 ± 1.67
5f	H	NO ₂	H	6.35 ± 0.70
5g	Cl	H	Cl	6.37 ± 0.67
5h	H	H	Cl	6.59 ± 0.18
	H	H	CH ₃	6.72 ± 0.58
5j	H	H	OC(O)CH ₃	7.18 ± 0.50
5k	H	H	OCF ₃	7.44 ± 0.30
5l	H	H	Br	7.50 ± 1.01
5m	H	CN	H	9.00 ± 1.15
5n	H	H	NO ₂	9.01 ± 0.25
5o	H	H	CN	10.21 ± 1.47
5p	H	H	OH	14.41 ± 2.76
5q	H	H	OCH ₃	14.69 ± 1.26
334	H	OH	H	16.53 ± 0.65
6a	H	H	NH ₂	17.63 ± 0.43
5s	H	H	OCH ₂ Ph	22.76 ± 0.34
5t	H	H	CO ₂ H	NI
5u	CO ₂ H	H	H	NI
5v	H	H	Ph	NI

^aIC₅₀ indicates 50% inhibition of control Chlamydia inclusions. NI: no inhibition seen at the tested concentration range (25–0.39 μg/mL).^bAll assays were performed in triplicate.^cIC₅₀ values are also presented in μM and pIC₅₀ units in Table S1. The presented data corresponds to mean ± standard deviation (SD).

Table 2. Detailing of *In Silico* Permeability Parameters for Some Selected Compounds and Comparison with IC₅₀ Values

compound	IC ₅₀ (μg/mL)	QlogPo/w ^a	QPPCaco ^b	QPPMDCK ^c	SASA ^d
5a	1.61 ± 0.11	4.08	1794.2	1465.8	563.86
5b	3.32 ± 0.51	4.63	3707.8	3212.5	569.57
5c	4.69 ± 1.20	5.06	3639.2	3188.8	589.42
5d	4.79 ± 0.19	5.21	3600.4	6855.0	582.84
5e	5.18 ± 1.67	5.68	3448.6	10 000.0	605.77
5f	6.35 ± 0.70	4.47	605.7	444.7	624.51
5j	6.72 ± 0.58	4.40	1155.8	900.70	661.59
11a	6.53 ± 0.45	5.75	3352.8	10 000.0	608.64
11b	6.93 ± 0.37	6.45	3384.2	10 000.0	654.63
5y	7.36 ± 0.78	4.80	3269.7	4617.9	563.87
5l	7.50 ± 1.01	5.42	3588.0	8068.2	595.47
5n	9.01 ± 0.25	4.16	429.4	307.3	605.51
11c	12.30 ± 0.87	5.97	3910.5	10 000.0	653.20
5g	14.69 ± 1.26	4.95	3500.7	2967.1	603.41
5s	22.76 ± 0.34	6.82	4104.2	3532.7	734.32
5t	NI	4.14	102.0	82.6	604.58
5u	NI	4.36	345.8	298.6	575.65
9d	NI	2.44	149.7	112.6	561.09

^aQlogPo/w—predicted octanol/water partition coefficient (−2.0 to 6.5).

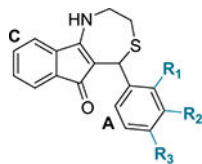
^bQPPCaco—predicted apparent Caco-2 cell permeability in nm/s (<25 poor, >500 great).

^cQPPMDCK—predicted apparent MDCK cell permeability in nm/s (<25 poor, >500 great).

^dTotal solvent accessible surface area (SASA) in square angstroms (300–1000). Range values are related to 95% of known drugs.⁶⁴ Predictions relate to drug absorption as oral dosage form. Values for the entire library of compounds can be found in File S1.

Table 3.

Chemical Structures of Dihydrothiazepine Derivatives 9a–d and Their IC₅₀ against *C. trachomatis* Serovar L2^{a,c}



compound	R ₁	R ₂	R ₃	IC ₅₀ (μg/mL) ^b
9a	H	H	OCH ₃	16.08 ± 0.54
9b	H	H	Br	21.35 ± 2.88
9c	H	H	CH ₃	21.45 ± 0.22
9d	H	H	CN	NI

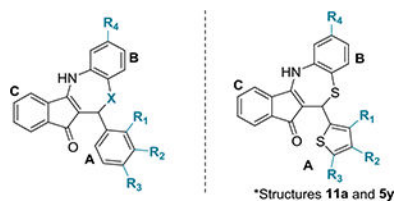
^a IC₅₀ indicates 50% inhibition of control Chlamydia inclusions. NI: no inhibition seen at the tested concentration range (25–0.39 μg/mL).

^b All assays were performed in triplicate.

^c IC₅₀ values are also presented in μM and pIC₅₀ units in Table S1. The presented data corresponds to mean ± SD.

Table 4.

Chemical Structures of Dihydrothiazepine Derivatives 11a–13c and Their IC₅₀ against *C. trachomatis* Serovar L2^{a,c}



compound	R ₁	R ₂	R ₃	R ₄	X	IC ₅₀ (μg/mL) ^b
11a	H	H	H	CF ₃	S	6.53 ± 0.45
11b	H	H	Br	CF ₃	S	6.93 ± 0.37
5y	H	H	H	H	S	7.36 ± 0.78
7a	H	OCH ₃	H	H	SO ₂	10.00 ± 3.69
11c	H	H	OCH ₃	CF ₃	S	12.30 ± 0.87
13a	H	H	CH ₃	H	NH	15.00 ± 3.23
13b	H	H	CN	H	NH	20.88 ± 1.94
13c	H	H	OCH ₃	H	NH	22.61 ± 0.32

^aIC₅₀ indicates 50% inhibition of control Chlamydia inclusions. NA: not applicable. Compounds **11a** and **5y** possess a 2-thiophene ring instead of the benzyl ring A.

^bAll assays were performed in triplicate.

^cIC₅₀ values are also presented in μM and pIC₅₀ units in Table S1.

The presented data corresponds to mean ± SD.

Table 5.

Antimicrobial Activity Panel of Compound 5a

		MIC ^p (μg/mL)														
		Sa ^a	Ab ^b	Ec ^c	Pa ^d	Ec ^e	Lr ^f	Lp ^g	Bp ^h	Cr ⁱ	Bt ^j	Ac ^k	Ca ^l	Cc ^m	Ck ⁿ	Ct ^o
5a		>125	>125	>125	>125	>125	>125	>125	>125	>125	>125	>125	>1000	>1000	>1000	>1000
AZM ^q	2	<1	<1	<1	125	2	1	2	<0.06	<0.06	0.5	>125	ne ^r	ne	ne	ne
FCZ ^s	ne	ne	ne	ne	ne	ne	ne	ne	ne	ne	ne	ne	1	2	16	0.5

^a *S. aureus* USA 300 JE2.

^b *Acinetobacter baumannii* 2208.

^c *Escherichia coli* K12.

^d *Pseudomonas aeruginosa* PA01.

^e *E. coli* nonpathogenic ATCC MG1655.

^f *Lactobacillus rhamnosus* CGMCC 1.3724.

^g *Lactobacillus plantarum* DSM 20174.

^h *Blautia producta* DSM 2950.

ⁱ *Clostridium ramosum* DSM 1402.

^j *Bacteroides thetaotaomicron* DSM 2079.

^k *Anaerostipes cacciae* DSM 14662.

^l *Candida albicans* 90028.

^m *Candida glabrata* 2001.

ⁿ *Candida krusei* 6558.

^o *Candida tropicalis* 750.

^p MIC: minimum inhibitory concentration (μg/mL).

^q AZM: azithromycin.

ne: not evaluated.
FCZ: fluconazole.

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Table 6.

Mutagenic Activity and Mutagenic Index (Displayed in Parenthesis) of 5a against *S. typhimurium* TA98 and TA100 Strains^a

concentration ($\mu\text{g}/\text{plate}$)	number of revertants/plate and (MI)			
	TA98		TA100	
	S9–	S9+	S9–	S9+
0 ^b	25.67 ^c \pm 5.51	24.33 \pm 9.50	206.00 \pm 7.07	201.00 \pm 9.90
50	26.00 \pm 1.41 (1.01)	32.00 \pm 11.31 (1.31)	190.50 \pm 4.95 (0.92)	185.00 \pm 7.07 (0.92)
150	34.50 \pm 0.71 (1.34)	35.00 \pm 1.41 (1.43)	187.50 \pm 3.54 (0.91)	187.00 \pm 5.66 (0.93)
500	38.00 \pm 2.83 (1.48)	43.50 \pm 6.36 (1.78)	233.00 \pm 7.07 (1.13)	191.50 \pm 9.19 (0.95)
1500	39.00 \pm 9.90 (1.51)	30.50 \pm 3.54 (1.25)	241.00 \pm 2.83 [*] (1.16)	198.50 \pm 4.95 (0.98)
5000	37.50 \pm 9.19 (1.46)	43.50 \pm 0.71 (1.78)	241.00 \pm 1.41 [*] (1.16)	227.00 \pm 9.90 (1.12)
control	34.50 \pm 2.12 ^d	137.00 \pm 7.07 ^f	260.70 \pm 15.30 ^e	1256.00 \pm 102.10 ^f

^{*} $P < 0.05$ (ANOVA) concentrations of 5a compared with negative control.

^a The results displayed represent the data for the assays in the absence (S9–) and presence (S9+) of an exogenous metabolism system.

^b Negative control (DMSO).

^c Number of revertant colonies.

^d 4-Nitrophenylenediamine (NPD, 10 $\mu\text{g}/\text{plate}$).

^e Sodium azide (2.5 $\mu\text{g}/\text{plate}$).

^f 2-Aminoanthracene (AA, 2.5 $\mu\text{g}/\text{plate}$).

The results are reported as means \pm SD.