

2015

Fingerprinting soybean germplasm and its utility in genomic research

Qijian Song

Agricultural Research Service, Soybean Genomics and Improvement Lab., Beltsville, Maryland, qijian.song@ars.usda.gov

D. L. Hyten

Soybean Genomics and Improvement Laboratory, Agricultural Research Service, Beltsville, MD, david.hyten@unl.edu

Gaofeng Jia

Agricultural Research Service, Soybean Genomics and Improvement Lab., Beltsville, Maryland

Charles V. Quigley

Agricultural Research Service, Soybean Genomics and Improvement Lab., Beltsville, Maryland

Edward W. Fickus

Agricultural Research Service, Soybean Genomics and Improvement Lab., Beltsville, Maryland

See next page for additional authors

Follow this and additional works at: <https://digitalcommons.unl.edu/agronomyfacpub>

 Part of the [Agricultural Science Commons](#), [Agriculture Commons](#), [Agronomy and Crop Sciences Commons](#), [Botany Commons](#), [Horticulture Commons](#), [Other Plant Sciences Commons](#), and the [Plant Biology Commons](#)

Song, Qijian; Hyten, D. L.; Jia, Gaofeng; Quigley, Charles V.; Fickus, Edward W.; Nelson, Randall L.; and Cregan, P. B., "Fingerprinting soybean germplasm and its utility in genomic research" (2015). *Agronomy & Horticulture -- Faculty Publications*. 818.

<https://digitalcommons.unl.edu/agronomyfacpub/818>

This Article is brought to you for free and open access by the Agronomy and Horticulture Department at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Agronomy & Horticulture -- Faculty Publications by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Authors

Qijian Song, D. L. Hyten, Gaofeng Jia, Charles V. Quigley, Edward W. Fickus, Randall L. Nelson, and P. B. Cregan

Fingerprinting soybean germplasm and its utility in genomic research

Qijian Song^{*}, David L. Hyten^{*1}, Gaofeng Jia^{*}, Charles V. Quigley^{*}, Edward W. Fickus^{*},
Randall L. Nelson[§] and Perry B. Cregan^{*}

^{*} United States Department of Agriculture, Agricultural Research Service, Soybean Genomics
and Improvement Lab., Beltsville, Maryland, USA

[§] United States Department of Agriculture, Agricultural Research Service, Soybean/Maize
Germplasm, Pathology and Genetics Research Unit and Department of Crop Sciences,
University of Illinois, Urbana, Illinois, USA

¹ Pioneer Hi-Bred International Inc, Johnston, Iowa, USA.

URL. The dataset for 19,648 soybean Plant Introductions genotyped with the SoySNP50K
BeadChip is available at SoyBase, the USDA-ARS Soybean Genetics and Genomics Database
(<http://soybase.org/snps/download.php>).

A short running title: Fingerprinting soybean germplasm

Key words: Soybean; germplasm; genotyping; SoySNP50K; genetic diversity; haplotype block map

Correspondence author: Qijian Song, Soybean Genomics and Improvement

Laboratory, Beltsville Agricultural Research Center – West, USDA, ARS, Beltsville, MD

20705-2350. Tel: 301-504-5723, e-mail: Qijian.Song@ars.usda.gov

ABSTRACT

The United States Department of Agriculture, Soybean Germplasm Collection includes 18,480 domesticated soybean and 1,168 wild soybean accessions introduced from 84 countries or developed in the U.S. This collection was genotyped with the SoySNP50K BeadChip containing greater than 50K SNPs. Redundant accessions were identified in the collection and distinct genetic backgrounds of soybean from different geographic origins were observed that could be a unique resource for soybean genetic improvement. We detected a dramatic reduction of genetic diversity based on linkage disequilibrium and haplotype structure analyses of the wild, landrace and North American cultivar populations and identified candidate regions associated with domestication and selection imposed by North American breeding. We constructed the first soybean haplotype block maps in the wild, landrace and North American cultivar populations and observed that most recombination events occurred in the regions between haplotype blocks. These haplotype maps are crucial for association mapping aimed at the identification of genes controlling traits of economic importance. A case-control association test delimited potential genomic regions along seven chromosomes that most likely contain genes controlling seed weight in domesticated soybean. The resulting dataset will facilitate germplasm utilization, identification of genes controlling important traits, and will accelerate the creation of soybean varieties with improved seed yield and quality.

INTRODUCTION

Domesticated in China, the cultivated soybean [*Glycine max* (L). Merr.] was first introduced in North America in 1765 (Hymowitz and Harlan 1983). Extensive soybean collecting started in the 1920's but systematic preservation did occur until the USDA Soybean Collection was established in 1949 (Carter et al. 2004). Few wild soybean [*Glycine soja* (Sieb. and Zucc.)]

accessions were added to the collection until the 1970s. At the time this project was initiated, the USDA Soybean Germplasm Collection contained approximately 19,700 soybean accessions. The collection includes more than 1,100 wild soybeans from China, Korea, Japan and Russia, and more than 18,000 cultivated soybeans from China, Korea, Japan and 84 other countries. Most of the cultivated soybean from China, Korea and Japan are landraces which are not the product of modern plant breeding. The earliest soybean accession in the USDA Soybean Germplasm Collection was collected before 1895. The accessions in the Collection are the sources of genes for soybean improvement not just in the North America (N. Am.), but worldwide.

Although some of the accessions in the USDA Soybean Germplasm Collection have been evaluated for their biotic and abiotic stress resistance, seed constituents and productivity during the past decades, knowledge of molecular variation, diversity, genetic architecture and selection that may underlie the phenotypic variation in the soybean genome based on the entire soybean collection is unknown. Previous studies have documented the existence of linkage disequilibrium (LD) (Hyten et al. 2007; Lam et al. 2010; Li et al. 2008; Zhu et al. 2003), genomic regions of selection (Lam et al. 2010), and the level of genetic diversity (Hyten et al. 2006; Lam et al. 2010; Li et al. 2008; Zhu et al. 2003) among various soybean populations. However, these studies were of limited scale in terms of sample size and/or the number of loci analyzed.

Highly selfing species, like soybean, are in many ways uniquely suitable for haplotype block mapping. Domestication and artificial selection have led to extensive LD and haplotype structure. The availability of inbred accessions makes it possible to observe rather than infer haplotypes. Whole-genome haplotype block mapping has been proposed in humans as a powerful tool to detect genes conditioning complex traits by limiting the number of SNPs to be

typed (Cardon and Abecasis 2003; Chien et al. 2013; Daly et al. 2001; Gabriel et al. 2002; Zhang et al. 2004a).

The N. Am. soybean crop accounts for 29% of world production, however, it may be at a low level of diversity due to several genetic bottlenecks. These include the domestication from wild soybean, the limited set of introduced landraces that form the N. Am. soybean genetic base and intensive selection for enhanced agronomic performance (Hyten et al. 2006). Determination of the genomic regions affected by, and the extent of fixation and linkage disequilibrium caused by the domestication, introduction and selection bottlenecks on a large population scale are fundamental information related to the future genetic improvement of soybean.

Here we report the molecular analysis of the soybean and wild soybean accessions in the USDA Soybean Germplasm Collection with a high-density genotyping array of soybean, the SoySNP50K beadchip (Song et al. 2013), which has been the largest such analysis in a plant species to date. The fingerprinting of the USDA Soybean Germplasm Collection provided a definition of soybean LD and haplotype block structure across the genome, identified genomic regions associated with soybean domestication and selection in N. Am. breeding programs and delimited regions associated with seed weight, an important component of seed yield.

MATERIALS AND METHODS

Soybean Germplasm and DNA analysis

The seeds of 19,648 accessions including 1,168 wild and 18,480 cultivated soybean accessions in the USDA Soybean Germplasm Collection (Urbana, IL) were crushed to a powder in a deep well microtiter plate (Thermo Scientific AB-0932) with a steel ball using a Retsch MM400

Mixer Mill at 30 hz for two minutes and DNA was extracted using the CTAB method (Keim et al. 1988).

SNP Genotyping

A high throughput SNP assay, the SoySNP50K Illumina Infinium II BeadChip containing 52,041 SNPs which were chosen from euchromatic and heterochromatic genome regions was used for genotyping. SNP genotyping was conducted following the procedures described previously (Song et al. 2013). Any polymorphic SNP with a rate of missing and heterozygous allele calls greater than 0.1 among the 19,648 soybean and wild soybean accessions was eliminated. The heterozygous allele calls in the remaining loci were set as missing in the subsequent analysis. The position of SNPs in the soybean genome was based on the Glyma1.01 assembly.

Similarity analysis

Genetic similarity between pairs of genotypes among the 18,480 cultivated and among the 1,168 wild accessions was calculated as the ratio of the number of identical SNP allele calls and the total number of SNPs for which allele calls were made for the pair.

Cluster analysis

Pair-wise distance among the accessions of 806 wild and 5,396 landrace soybeans was obtained based on the allelic dissimilarity of the 42,509 SNPs, the neighbor-joining tree was constructed using the software Mega 5.10 (Tamura et al. 2011).

LD analysis

LD was analyzed within the wild, landrace and N. Am. cultivar populations with 806, 5,396 and 562 accessions, respectively. Only the SNPs with minor allele frequency $\geq 5\%$ were included for LD calculation and construction of haplotype blocks. Calculation of pairwise LD (r^2) among SNPs and identification of haplotype blocks was based upon SNPs within 1Mb windows using the software PLINK (Purcell et al. 2007). Haplotype blocks were identified through estimates of D' for all pairwise combinations of SNPs. Pairs of SNPs were in a haplotype block if the one-tailed upper 95% confidence limit on D' was greater than 0.98 and the lower limit was above 0.7 (Gabriel et al. 2002).

Haplotype block sharing

To compare block boundaries among wild soybeans, landraces, and N. Am. cultivars populations (Dataset S1), the ratio of haplotype sharing across populations was calculated based on the method of Gabriel et al. (2002) with modification. SNPs in the haplotype blocks of the two populations being compared were identified and concordance of all SNP pairs in the same block of one population vs. SNP pairs in the other population was examined. A SNP pair was concordant if the pair was assigned to the same block in both populations, and non-concordant if the assignment was not to the same block. The percentage of haplotype block sharing was calculated as: (the number of concordant SNPs/(number of concordant SNPs + number of non-concordant SNPs)) $\times 100$.

Calculation of fixation index (F_{st})

The F_{st} between the wild and landrace genotypes and the F_{st} between the landrace and N. Am. cultivars were calculated using the software Arlequin v3.1 (Excoffier et al. 2005).

Expected number of genes in and between haplotype blocks

The expected number of genes in and between the haplotype blocks of the euchromatic and heterochromatic regions of each population was calculated based upon an even distribution of genes per physical distance in both the euchromatic and heterochromatic regions. The positions of the 46,430 high-confidence protein coding genes were downloaded at <http://www.phytozome.net> and the observed number of genes in and between the blocks of the euchromatic and heterochromatic regions of each population was counted.

Recombination rate in and between haplotype blocks

The recombination rate in centiMorgans within haplotype blocks was estimated based upon the genetic distance of 21,483 SNPs in the linkage map developed via the analysis of a Williams 82 x PI479752 population with 1,083 recombinant inbred lines which were genotyped with the SoySNP50K chip. Total genetic distance, as well as total length of sequence (bp) between the most distant SNPs in haplotype blocks, was calculated in order to estimate the observed and expected recombination rate within haplotype block regions.

Genome-wide association analysis of seed weight

For the purpose of genome wide association analysis of seed weight (g/100 seeds), the dataset containing seed weight of the *G. max* accessions was downloaded from the Germplasm Resources Information Network (GRIN) <http://www.ars-grin.gov/npgs/searchgrin.html>. Given that the seed weight of the accessions was observed in different years and/or environments, variation of seed weight impacted by different growing environments was expected. In order to eliminate such variation, we used a method to treat seed weight as a dichotomous, i.e. 0, and 1,

rather than a quantitative trait. If the seed weight is ≥ 20 g/100 seeds, the seed weight was set to 1, and if the seed weight is ≤ 10 g/100 seeds, the value was set to 0. Thus, only 3,753 accessions with seed weight ≥ 20 grams or ≤ 10 grams per 100 seeds were included in the association analysis. It is very unlikely that the seed weight classification of the accessions based on this standard would be affected by environmental effects among years and locations. Then a genome-wide association study of the seed size was executed with the PLINK procedure “case-control association test” taking into account population stratification with a defined number of clusters, K. The K estimation of the population was obtained using the software fastStructure (Raj et al. 2014), K=2-15 was chosen. A value of $-\log(p)$, where the p is the genomic control adjusted significance level of each locus was obtained from PLINK. Significant loci were identified when the $-\log(p)$ value was greater than 3.

RESULTS

Redundant and highly similar soybean accessions

Of the 52,041 SNPs in the SoySNP50K bead pool, a total of 42,509 SNPs were polymorphic and had a rate of missing and heterozygous allele calls < 0.1 among the 19,648 soybean and wild soybean accessions characterized. Based on the pair-wise genetic similarity of the accessions calculated from the 42,509 SNPs, 1,682 and 95 accessions were 100% identical to at least one other accession among the 18,480 *G. max* and 1,168 *G. soja* accessions, respectively. Overall 4,303 *G. max* (23%) and 362 *G. soja* (30%) accessions were at least 99.9% identical to another accession in the collection (Table 1).

After eliminating all but one accession within each group of accessions with greater than 99.9% similarity, a total of 806 *G. soja* accessions from China, Korea, Japan and Russia and 14,181 *G.*

max accessions remained. Of the 14,181 *G. max* accessions, a total of 5,396 landraces from China, Korea and Japan, and 562 N. Am. cultivars were used for further analysis (Table S1).

Genetic relationship with geographic origins of soybean accessions

Cluster analysis of the 806 wild soybean accessions from China, Korea, Japan and Russia, and 5,396 landraces from China, Korea and Japan showed that wild soybeans and landraces from different countries were well separated and the clusters of wild soybean and soybean landraces were consistent with their geographic origins (Fig.1). This indicates that landraces from China, Korean and Japan have distinct genetic backgrounds that are unique and that could be useful in genetic soybean improvement.

Extent of LD among wild, landrace and N. Am. cultivar populations

The trend of LD decay among the 806 wild, 5,396 landrace and 562 N. Am. cultivar populations was very similar in the euchromatic and heterochromatic regions. The LD was more extensive in the N. Am. cultivars than in the landraces, and in the landraces than in the wild soybeans. LD was much more extensive in the heterochromatic than in the euchromatic regions in each population (Fig. 2). In the euchromatic regions, the LD declined to half of its maximum value within approximately 20kb, 75kb and 160kb in the wild, landrace and N. Am. cultivar populations, respectively, while in the heterochromatic regions, the LD declined to half of its maximum value within 350kb, 900kb and 970kb in the wild, landrace and N. Am. cultivar populations, respectively. Thus, in both regions, LD was the lowest in the wild soybean and the highest in the N. Am. cultivars.

Haplotype map and haplotype block structure among wild, landrace and N. Am. cultivar populations

The total number of haplotype blocks ranged from 3,000-5,300 among the 806 wild, 5,396 landrace and 562 N. Am. cultivar populations (Table S2, S3 and S4). In each of the populations most of these blocks resided in the euchromatic regions where there were 4,331 blocks in the wild, 4,777 in the landrace and 2,763 in the N. Am. cultivar population. The average block size was 10.7, 39.6 and 79.6kb, respectively, in the three populations (Table 2). Similarly, in the heterochromatic regions, the smallest average block size was detected in the wild and the largest in the N. Am. cultivar population. However, the average haplotype block size was much smaller in euchromatic regions vs. the heterochromatic regions in all three populations. Approximately 10%, 41% and 48% of the euchromatic regions were in haplotype blocks in the wild, landrace and N. Am. cultivars, respectively. The percentage of the heterochromatic regions in haplotype blocks was also the lowest in the wild, followed by the N. Am. cultivar and landrace populations. Most of the haplotype blocks (>83.7%) were shared between any two of the three populations (Table S5).

Analysis of the distribution of haplotype block size in euchromatic regions showed that, although >50% of the haplotype blocks extended <15kb in the landrace and <20kb in the N. Am. cultivar accessions, there were 10% and 24% of the blocks with size >100kb in the landrace and N. Am. cultivar accessions, respectively (Fig. 3a). In the wild soybean, more than 50% of the blocks were less than 5kb and less than 1% of the blocks were greater than 100kb. In the heterochromatic regions, a large proportion of the blocks were >900kb in size which included 42% of the blocks in the N. Am. cultivar accessions followed by 37% in the landraces and 24% in the wild soybean population (Fig. 3b). Clearly, the wild soybean haplotype block structure is

characterized by much smaller blocks. In terms of haplotype diversity, the three populations were quite similar with the mean number of haplotypes/block of 3.4, 3.5 and 3.7 in the wild, N. Am. cultivar and landrace populations, respectively (Table 3). Among these, >66% of the haplotypes in euchromatic and >55% in heterochromatic regions had a frequency >10% among the three populations (Table S6). The relatively low number of haplotypes/block was surprising given the fact that the average number of SNPs per block ranged from 2.9 to 7.8 in the euchromatic regions and from 6.8 to 10.2 in the heterochromatic regions of the wild, landrace and the N. Am. cultivar populations.

Although the number of genes in haplotype blocks was slightly lower than expectation in the heterochromatic regions of the three populations and higher than expectation in the euchromatic regions of the landrace and N. Am. cultivar populations, the bias of gene density within blocks vs. between blocks was not severe (Table S7). However, as expected, the recombination rate within blocks was dramatically reduced and was approximately only 12%-39% and 12-41% of expectation in the blocks of the euchromatic and heterochromatic regions, respectively, in the wild, landrace, and N. Am. cultivar populations (Table S8). The wild population had the most dramatic reduction in recombination frequency and the N. Am. cultivar population had the least.

Genomic regions associated with domestication and selective sweeps associated with soybean breeding

Genome-wide F_{st} was 0.23 between the wild soybean (806 accessions) and landrace (5,396 accessions) populations (Table S9) with a standard deviation of 0.242, and 0.11 between the landrace and N. Am. cultivar (562 accessions) populations, with a standard deviation of 0.147.

Thus, the threshold of F_{st} with a two-tailed significance level of 5% was 0.704 and 0.398 between

the wild and landrace and between the landrace and N. Am. cultivar populations, respectively. Analysis of average F_{st} and the distribution of individual loci above the F_{st} threshold at the 5% significance level among chromosomes suggested that chromosomes underwent different selective pressure during domestication versus that imposed by modern breeding. For example, the average F_{st} between the wild and landrace populations was much greater along Gm12 (0.315) than Gm18 (0.174), and there was a much higher proportion of loci with significant F_{st} (14.05 % vs. 1.59%) on Gm12 versus Gm18. The F_{st} between the landrace and N. Am. cultivar populations ranged from 0.060 on Gm05 to 0.157 on Gm19. The proportion of loci with $F_{st} \geq 0.398$ was 11.86% on Gm19 vs. 2.21% on Gm05 (Table S10).

We examined the potential relationship of the candidate regions with several traits. Determinate versus indeterminate growth habit in the cultivated soybean is thought to be a trait which has undergone strong selection in N. Am. breeding programs. The approximate percentage of accessions with determinate growth habit was 65% in landraces vs. 32% in the N. Am. cultivars based on the data from GRIN (Germplasm Resources Information Network, www.ars-grin.gov/). The gene Glyma19g37890.1 which is located at 44.93Mb~44.97Mb of Gm19 was identified as the gene conditioning determinacy (Tian et al. 2010). In the region, we observed high F_{st} (>0.40) between the landrace and the N. Am. cultivar populations for six of seven SNPs. However, a low F_{st} value was observed between the wild and landrace populations in this region of Gm19. This is consistent with the results reported by Tian et al. (2010) that determinacy in landraces was the result of a gene mutation and not selection from *G. soja*. A number of SNPs with high F_{st} between the wild and landrace populations were detected in regions previously reported to contain domestication QTL. These included the region at 4.00-7.00Mb of Gm05 containing a QTL conditioning pod number per plant (Zhang et al. 2007), plant height (Chen et al. 2007;

Kabelka et al. 2004) and days to maturity (Kabelka et al. 2004) and 2.8-7.1Mb, 41.1-42.4Mb and 46.1-49.1Mb of Gm19 (Bailey et al. 1997; Kang et al. 2009) where QTL conditioning pod shattering have been reported.

Application of genotyping data to the analysis of genome-wide association of seed weight

We identified 1,936 and 1,817 *G. max* accessions with seed weight ≥ 20 and ≤ 10 grams/100 seeds in the GRIN database, respectively. FastStructure analysis showed that the appropriate K was 5-15, and the Admixture analysis (Alexander et al. 2009) showed that the appropriate K was 12. As the accessions were from 12 maturity groups, the K=12 was chosen. A total of 30 loci were significantly associated with seed weight and the loci were concentrated in eight regions of chromosomes Gm01 at 32.3 Mb, Gm06 at 19.7 Mb, Gm11 at 28.2-30.0Mb, Gm13 at 38.4Mb, Gm14 at 28.5-34.2Mb, Gm17 at 2.4-2.5Mb and 8.7-8.8 Mb and Gm19 at 42.7-43.1Mb. (Figure S1). As shown in Table S11, QTL for seed weight were previously reported in similar genomic regions using recombinant inbred line populations (Gai et al. 2007; Hoeck et al. 2003; Hyten et al. 2004; Mian et al. 1996; Panthee et al. 2005; Specht et al. 2001; Zhang et al. 2004b).

However, because these were quantitative trait locus analyses of lines derived from single cross populations with the resulting high level of LD, the regions in which the reported seed weight QTL resided were poorly defined and in some cases the reported regions spanned as much as 17 Mb.

The association of the regions with seed size is also consistent with the observation that, with the exception of Gm1:32.3Mb and Gm13:38.4, the remaining regions each contained loci with high F_{st} (>0.70) between the wild soybean and landrace accessions.

DISCUSSION

The annual accessions in the USDA Soybean Germplasm Collection were assayed with the SNPs contained on the SoySNP50K (Song et al. 2013) BeadChip. The dataset will be instrumental in making large-scale, genome-wide association studies a reality for other laboratories and will help researchers narrow the genomic regions of the targeted QTL. Because the seeds of the soybean germplasm accessions and the DNA genotyping data generated from this study are publically available, researchers will be able to integrate their phenotypic data with the genome-wide SNP data to identify genetic factors that influence their traits of interest, such as biotic and abiotic stress resistance, as well as, agronomic and seed quality traits. In addition, this work provides a rich renewable genetic resource for soybean scientists and breeders to study the prevalence of haplotype variation and genetic diversity in soybean germplasm, as well as to identify regions and the specific genes associated with a diversity of traits and tag SNPs for selection of desired genotypes. In a recent report of an association analysis of soybean seed protein and oil content using the set of SNPs described in this report (Hwang et al. 2014), 17 genome positions associated with level of seed protein and 13 genome positions associated with seed oil content were identified. A total of 12 of the 17 regions containing protein QTL and 8 of the 13 regions with oil QTL had previously been reported based upon QTL analyses. It was concluded that the genome-wide association study not only identified many of the previously reported QTL controlling seed protein and oil, but also resulted in more narrowly defining the genomic regions containing the genes of interest than the regions reported based upon QTL analysis. This was also true for the genome-wide association analysis of seed weight that we report here. These narrower genomic regions will expedite the identification and cloning of the causal genes.

The high rate of redundant and highly similar accessions detected in the USDA Soybean Germplasm Collection is not surprising. Soybean accessions have been acquired from more than 80 countries worldwide and due to incomplete or nonexistent passport information, the same accession with different names have been acquired a number of times. In addition, in countries which are the center of origin of soybean such as China, soybean was commonly named by farmers according to soybean seed coat color, seed size and shape, pod color or the month of maturity and thus it is very common for the same accession to have different names in different areas of the country. As a limited number of agronomic or morphological traits are available to distinguish these accessions, profiling each accession in the USDA Soybean Germplasm Collection with a large number of molecular markers is essential to understand the level of repetitiveness, thus increasing the efficiency of germplasm preservation, characterization and promoting the more efficient utilization of the genetic resources in soybean breeding programs. This research provides the first in-depth analysis of the current soybean germplasm collection in the US.

The extent of linkage disequilibrium and the average haplotype block sizes were the lowest in the wild soybean population, were greater in the landrace population and were greater still in the N. Am. cultivar population. Studies indicate that long range LD and increased haplotype block size can arise by several different mechanisms including natural and artificial selection in the form of selective sweeps or background selection and population bottlenecks and founding effects from ancestrally conserved segments (Phillips et al. 2003). Domestication is the process of adapting a plant species to the production environment and to the needs of the consumer. This involves changes in the gene pool over generations. During domestication the plant species will differ from its wild counterparts by greatly diminished effective population size, and

consequently, increased LD. Modern breeding practices further shrink the effective population size by using a small fraction of accessions from the landrace gene pool such as occurred when the soybean was introduced into North America, e. g. Gizlice et al. (1994) identified a group of 14 genotypes which had contributed 80.5% of the allelic diversity present in N. Am. soybean cultivars released between 1947 and 1988 (Gizlice et al. 1994).

Subsequently, the extensive use of a small number of elite genotypes in breeding programs further reduces genetic variability. Thus, the extended regions of LD and increased haplotype block size are an indicator of reduced genomic diversity from wild to landrace populations and from landrace to the N. Am. cultivar populations, which was consistent with the previous report that genetic diversity of the landrace and cultivar populations was dramatically reduced by domestication and selection (Hyten et al. 2006), respectively.

We have provided the first high resolution haplotype maps based on the largest sample size and the largest number of loci reported in soybean thus far. With the aid of the haplotype block map, we can efficiently select and tag SNPs for optimized association analyses, explore the nature of fine-scale recombination and identify regions that may have been subject to natural selection, track the fragments that are transmitted from generation to generation and gain more insight into the organization of the soybean genome.

Knowledge of the positions of the regions associated with domestication and selection imposed in N. Am. soybean breeding programs is critical for the identification of genes controlling important agronomic traits. Previously, we reported regions related to the domestication and breeding improvement in N. Am. breeding programs using populations with 96 *G. soja*, 96 landraces and 96 elite cultivars (Song et al. 2013). However, due to limited sample size, the span

of the regions identified was still quite large. Now with the analysis of a vastly larger number of genotypes, the regions associated with domestication and intensive breeding were identified more narrowly and accurately. We can anticipate that the identification of such regions with strong signatures of selection will be the targets of investigation with the goal of identifying the impacted phenotype and the specific gene/genes that underlie the phenotypic changes.

Using the genotyping data, we identified major candidate regions on seven chromosomes that were significantly associated with seed weight. This is the first report that has narrowly delimited the regions controlling this important trait related to seed yield. Clearly, the candidate genes identified in these regions are of interest for further investigation.

ACKNOWLEDGEMENTS

We thank Dr. Donghe Xu at the Japan International Research Center for Agricultural Sciences, Tsukuba, Japan and Dr. Mun Sup Yoon at the Genetic Resources Division, National Institute of Agricultural Biotechnology, RDA, Suwon, South Korea for the identification of landrace genotypes from Japan and Korea. We thank Rob Parry and Chris Pooley for their technical support in assembling the necessary hardware and software required for the Illumina sequence analysis, David M. Grant at USDA-ARS, Ames, IA for depositing the genotyping dataset in SoyBase. This research was supported with funding from the United Soybean Board Project #8265. The support of the United Soybean Board is greatly appreciated.

AUTHOR CONTRIBUTIONS

D.L.H., Q.J. and P.B.C. provided project planning and coordination. R.L.N. prepared plant materials. C.V.Q. and E.W.F. performed DNA extraction. C.V.Q., G.J. and E.W.F. performed

molecular genotyping. Q.S., G.J. and C.V.Q. performed data analysis. Q.S and P.B.C. prepared the manuscript.

LITERATURE CITED

- Alexander, D.H., J. Novembre, and K. Lange, 2009 Fast model-based estimation of ancestry in unrelated individuals. *Genome Res* 19 (9):1655-1664.
- Bailey, M.A., M.A.R. Mian, T.E. Carter, D.A. Ashley, and H.R. Boerma, 1997 Pod dehiscence of soybean: identification of quantitative trait loci. *Journal of Heredity* 88 (2):152-154.
- Cardon, L.R., and G.R. Abecasis, 2003 Using haplotype blocks to map human complex trait loci. *TRENDS in Genetics* 19 (3):135-140.
- Carter, T.E., R.L. Nelson, C.H. Sneller, and Z. Cui, 2004 Genetic diversity in soybean. *Soybeans: Improvement, production, and uses* (American Society of Agronomy Monograph Series):303-416.
- Chen, Q., Z. Zhang, C. Liu, D. Xin, H. Qiu *et al.*, 2007 QTL Analysis of Major Agronomic Traits in Soybean *Ag. Sci. in China* 6 (4):399-405.
- Chien, Y.-L., H.-G. Hwu, C.S.J. Fann, C.-C. Chang, M.-T. Tsuang *et al.*, 2013 DRD2 haplotype associated with negative symptoms and sustained attention deficits in Han Chinese with schizophrenia in Taiwan. *Journal of human genetics* 58 (4):229-232.
- Daly, M.J., J.D. Rioux, S.F. Schaffner, T.J. Hudson, and E.S. Lander, 2001 High-resolution haplotype structure in the human genome. *Nature genetics* 29 (2):229-232.
- Excoffier, L., G. Laval, and S. Schneider, 2005 Arlequin (version 3.0): an integrated software package for population genetics data analysis. *Evol Bioinform Online* 1:47-50.
- Gabriel, S.B., S.F. Schaffner, H. Nguyen, J.M. Moore, J. Roy *et al.*, 2002 The structure of haplotype blocks in the human genome. *Science* 296 (5576):2225-2229.
- Gai, J., Y. Wang, X. Wu, and S. Chen, 2007 A comparative study on segregation analysis and QTL mapping of quantitative traits in plants-with a case in soybean. *Front. of Ag. in China*. 1 (1):1-7.
- Gizlice, Z., T. Carter, and J. Burton, 1994 Genetic base for North American public soybean cultivars released between 1947 and 1988. *Crop Sci* 34 (5):1143-1151.
- Hoeck, J.A., W.R. Fehr, R.C. Shoemaker, G.A. Welke, S.L. Johnson *et al.*, 2003 Molecular marker analysis of seed size in soybean. *Crop Sci*. 43 (1):68-74.
- Hwang, E.-Y., Q. Song, G. Jia, J.E. Specht, D.L. Hyten *et al.*, 2014 A genome-wide association study of seed protein and oil content in soybean. *BMC Genomics* 15 (1):1.
- Hymowitz, T., and J.R. Harlan, 1983 Introduction of soybean to North America by Samuel Bowen in 1765. *Economic Botany* 37 (4):371-379.
- Hyten, D.L., I.-Y. Choi, Q. Song, R.C. Shoemaker, R.L. Nelson *et al.*, 2007 Highly variable patterns of linkage disequilibrium in multiple soybean populations. *Genetics* 175 (4):1937-1944.
- Hyten, D.L., V.R. Pantalone, C.E. Sams, A.M. Saxton, D. Landau-Ellis *et al.*, 2004 Seed quality QTL in a prominent soybean population. *Theor. Appl. Genet.* 109 (3):552-561.
- Hyten, D.L., Q. Song, Y. Zhu, I.Y. Choi, R.L. Nelson *et al.*, 2006 Impacts of genetic bottlenecks on soybean genome diversity. *Proc Natl Acad Sci U S A* 103 (45):16666-16671.
- Kabelka, E.A., B.W. Diers, W.R. Fehr, A.R. LeRoy, I.C. Baianu *et al.*, 2004 Putative alleles for increased yield from soybean plant introductions. *Crop Sci*. 44 (3):784-791.

- Kang, S.-T., M. Kwak, H.-K. Kim, M.-G. Choung, W.-Y. Han *et al.*, 2009 Population-specific QTLs and their different epistatic interactions for pod dehiscence in soybean [*Glycine max* (L.) Merr.]. *Euphytica* 166 (1):15-24.
- Keim, P., T.C. Olson, and R.C. Shoemaker, 1988 A rapid protocol for isolating soybean DNA. *Soybean Genet. Newsl.* 15:150-152.
- Lam, H.M., X. Xu, X. Liu, W. Chen, G. Yang *et al.*, 2010 Resequencing of 31 wild and cultivated soybean genomes identifies patterns of genetic diversity and selection. *Nat Genet* 42 (12):1053-1059.
- Li, Y., R. Guan, Z. Liu, Y. Ma, L. Wang *et al.*, 2008 Genetic structure and diversity of cultivated soybean (*Glycine max* (L.) Merr.) landraces in China. *Theor and Appl Genet* 117 (6):857-871.
- Mian, M.A.R., M.A. Bailey, J.P. Tamulonis, E.R. Shipe, T.E. Carter *et al.*, 1996 Molecular markers associated with seed weight in two soybean populations. *Theor Appl Genet* 93 (7):1011-1016.
- Panthee, D.R., V.R. Pantalone, D.R. West, A.M. Saxton, and C.E. Sams, 2005 Quantitative trait loci for seed protein and oil concentration, and seed size in soybean. *Crop Sci* 45 (5):2015-2022.
- Phillips, M.S., R. Lawrence, R. Sachidanandam, A.P. Morris, D.J. Balding *et al.*, 2003 Chromosome-wide distribution of haplotype blocks and the role of recombination hot spots. *Nat Genet* 33 (3):382-387.
- Purcell, S., B. Neale, K. Todd-Brown, L. Thomas, M.A. Ferreira *et al.*, 2007 PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 81 (3):559-575.
- Raj, A., M. Stephens, and J.K. Pritchard, 2014 fastSTRUCTURE: variational inference of population structure in large SNP data sets. *Genetics* 197 (2):573-589.
- Song, Q., D.L. Hyten, G. Jia, C.V. Quigley, E.W. Fickus *et al.*, 2013 Development and evaluation of SoySNP50K, a high-density genotyping array for soybean. *PLoS One* 8 (1):e54985.
- Specht, J.E., K. Chase, M. Macrander, G.L. Graef, J. Chung *et al.*, 2001 Soybean response to water: A QTL analysis of drought tolerance *Crop Sci.* 4 (2):493-509.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei *et al.*, 2011 MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular biology and evolution* 28 (10):2731-2739.
- Tian, Z., X. Wang, R. Lee, Y. Li, J.E. Specht *et al.*, 2010 Artificial selection for determinate growth habit in soybean. *Proc Natl Acad Sci U S A* 107 (19):8563-8568.
- Zhang, D., H. Cheng, H. Wang, Z. Hengyou, C. Liu *et al.*, 2007 Identification of genomic regions determining flower and pod numbers development in soybean (*Glycine max* L) *J. Genet. and Genom* 37 (8):545-556.
- Zhang, K., Z.S. Qin, J.S. Liu, T. Chen, M.S. Waterman *et al.*, 2004a Haplotype block partitioning and tag SNP selection using genotype data and their applications to association studies. *Genome Research* 14 (5):908-916.
- Zhang, W.K., Y.J. Wang, G.Z. Luo, J.S. Zhang, C.Y. He *et al.*, 2004b QTL mapping of ten agronomic traits on the soybean (*Glycine max* L. Merr.) genetic map and their association with EST markers. *Theor Appl Genet* 108 (6):1131-1139.
- Zhu, Y.L., Q.J. Song, D.L. Hyten, C.P. Van Tassell, L.K. Matukumalli *et al.*, 2003 Single-nucleotide polymorphisms in soybean. *Genetics* 163 (3):1123-1134.

FIGURE LEGENDS

Figure 1. Dendrogram of wild and landrace genotypes from different countries

Figure 2. LD in euchromatic and heterochromatic regions. a. LD in euchromatic regions of wild, landrace, and N. Am. cultivar soybean populations. **b.** LD in heterochromatic regions of the wild, landrace, and N. Am. cultivar soybean populations

Figure 3. Distribution of haplotype block size. a. Haplotype block size in euchromatic regions of wild, landrace, and N. Am. cultivar populations. **b.** Haplotype block size in heterochromatic regions of the wild, landrace, and N. Am. cultivar populations

Table 1. Number of accessions in the USDA Soybean Germplasm Collection with similarity >99.9% based on SNP comparisons

| Similarity among accessions | <i>G. soja</i> | <i>G. max</i> |
|--|----------------|------------------|
| Accessions 100% similar to another accession | 95 | 1682 |
| Accessions >99.9% similar to another accession | 362 | 4306 |
| Proportion of identical accessions (%) | (95/1168)=8% | (1682/18480)=9% |
| Proportion of accessions with >99.9% similar (%) | (362/1168)=30% | (4306/18480)=23% |

Table 2. Number of haplotype blocks and their size in the wild, landrace and N. Am. cultivar soybean populations

| Population | Genome-wide | | | Euchromatic regions | | | | Heterochromatic regions | | | |
|------------------|----------------------|--------------------------|--------------|---------------------|--------------------------------------|-------------------------|---|-------------------------|--------------------------------------|-------------------------|---|
| | Number of accessions | Number of SNPs in blocks | Total blocks | Number of blocks | Total sequence length in blocks (kb) | Average block size (kb) | Proportion of euchromatic regions in blocks | Number of blocks | Total sequence length in blocks (kb) | Average block size (kb) | Proportion of heterochromatic regions in blocks |
| wild | 806 | 14343 | 4624 | 4331 | 46246 | 10.7 | 0.10 | 293 | 149958 | 511.8 | 0.31 |
| landrace | 5396 | 28111 | 5226 | 4777 | 189134 | 39.6 | 0.41 | 449 | 281061 | 626.0 | 0.57 |
| N. Am. cultivars | 562 | 24753 | 3093 | 2763 | 219872 | 79.6 | 0.48 | 330 | 222619 | 674.6 | 0.45 |

Table 3. Haplotype block structure in the wild, landrace and N. Am. cultivar soybean populations

| Population | Genome-wide | | Euchromatic regions | | | | Heterochromatic regions | | | |
|------------|----------------------|----------------------------|----------------------|----------------------------|--------------------------|----------------------|-------------------------|----------------------------|--------------------------|----------------------|
| | Number of haplotypes | Number of haplotypes/block | Number of haplotypes | Number of haplotypes/block | Number of SNPs in blocks | Number of SNPs/block | Number of haplotypes | Number of haplotypes/block | Number of SNPs in blocks | Number of SNPs/block |
| wild | 15890 | 3.4 | 14356 | 3.3 | 12417 | 2.9 | 1534 | 5.2 | 1984 | 6.8 |
| landrace | 19529 | 3.7 | 17617 | 3.7 | 23990 | 5.0 | 1912 | 4.3 | 4121 | 9.2 |
| N. Am. | 10933 | 3.5 | 9753 | 3.5 | 21435 | 7.8 | 1180 | 3.6 | 3361 | 10.2 |

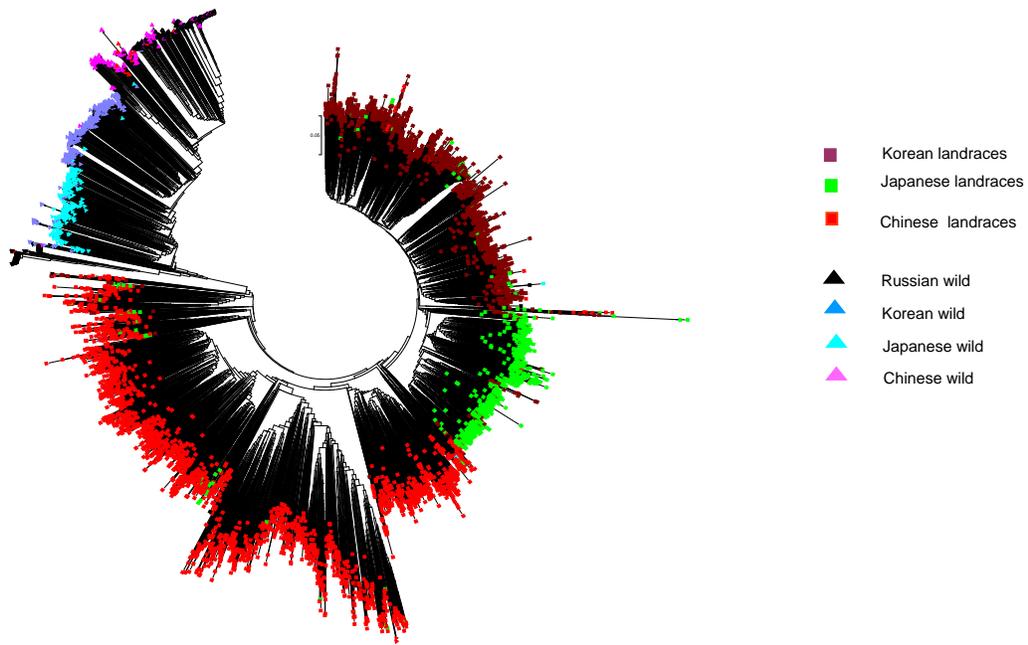


Figure 1

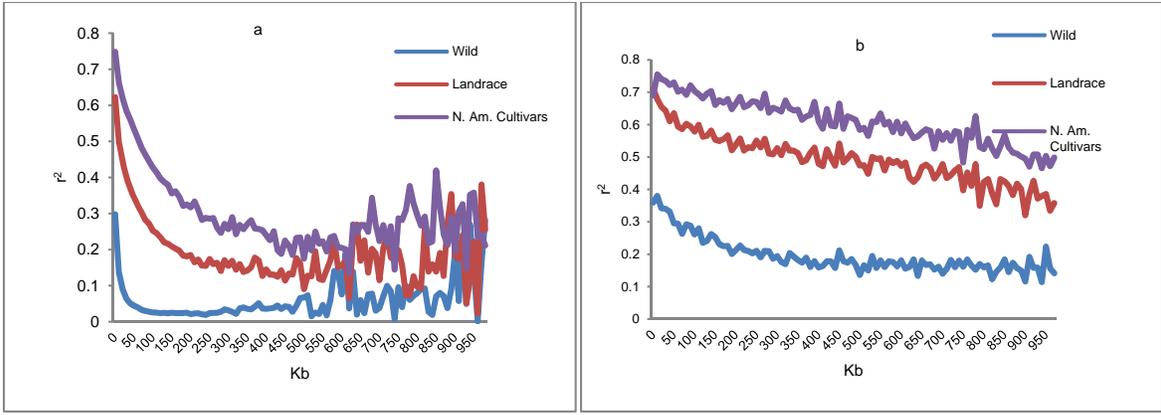


Figure 2.

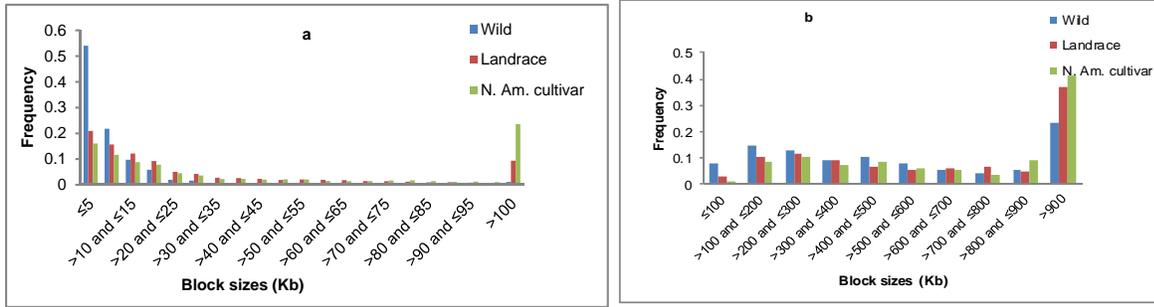


Figure 3.