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Aspen M. Workman

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, aspen.workman@ars.usda.gov

Aaron M. Dickey

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, aaron.dickey@usda.gov

Michael P. Heaton

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, mike.heaton@usda.gov

Michael L. Clawson

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, Mike.Clawson@usda.gov

Timothy P.L. Smith

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, tim.smith@ars.usda.gov

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Complete Genome Sequences of Two Genotype A2 Small Ruminant Lentiviruses Isolated from Infected U.S. Sheep

Aspen M. Workman, Aaron M. Dickey, Michael P. Heaton, Michael L. Clawson, Timothy P. L. Smith

USDA, ARS, U.S. Meat Animal Research Center, Clay Center, Nebraska, USA

ABSTRACT Two distinct subgroups of genotype A2 small ruminant lentiviruses (SRLVs) have been identified in the United States that infect sheep with specific host transmembrane protein 154 (*TMEM154*) diplotypes. Here, we report the first two complete genome sequences of SRLV strains infecting U.S. sheep belonging to genotype A2, subgroups 1 and 2.

Small ruminant lentiviruses (SRLVs) infect sheep, goats, and wild ruminants, causing an incurable multisystemic chronic disease (1). SRLVs are phylogenetically divided into five groups, A to E, which are further divided into different subtypes based on their *gag* and *pol* sequences (2, 3). In the United States, genotypes A2 and B1 infect sheep and goats (4). Viral tropism and host susceptibility to the different SRLV strains are influenced by host and viral genotypes. In sheep, amino acid sequence variation encoded by the transmembrane protein 154 (*TMEM154*) gene is associated with susceptibility to SRLV A2 infection (5–9). On the pathogen side, two distinct SRLV A2 subgroups have evolved to infect sheep with specific *TMEM154* diplotypes (8). The lack of complete genome sequences for representatives of these viral subgroups has hampered progress in the identification of viral elements correlated with host adaptation associated with *TMEM154* diplotypes.

Viral genomic RNA was sequenced from the lungs of clinically ill sheep at the U.S. Meat Animal Research Center. These virus strains (USMARC-200303013-1 and USMARC-199906011-2, from subgroups 1 and 2, respectively) were part of the original study describing the association of SRLV genetic subgroups with TMEM154 diplotypes (8). Encapsidated viral RNA was enriched in lung homogenates by nuclease treatment to degrade unprotected host and environmental nucleic acids. The remaining RNA was isolated using TRIzol LS (Life Technologies, Inc., Carlsbad, CA) and used to generate sequencing libraries according to the RNA Iso-Seq protocol (Pacific Biosciences, Menlo Park, CA), with one important modification. Fragment analysis revealed little RNA in the expected range of 9 kb for complete viral genomes. Therefore, poly(A) tails were added to the 3' ends of the RNA using a Poly(A) polymerase tailing kit (Epicentre, Madison, WI) to allow cDNA synthesis of subgenomic fragments using the SMARTer PCR cDNA synthesis kit (Clontech, Mountain View, CA). The resulting cDNA was amplified then size fractionated with ELF (Sage Science, Beverly, MA). The largest fragments (approximately 2 to 3 kb) were used to generate SMRTbell libraries using the PacBio template prep kit. The SMRTbell templates were then sequenced using Pacific Biosciences RSII sequencing technology.

Reads were error-corrected using the SMRT Analysis server version 2.3.0 and imported into the Geneious software suite (version 9.1.7; Biomatters, Auckland, New Zealand). Adapter sequences and the added poly(A) tails were trimmed from reads

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Address correspondence to Aspen M. Workman, aspen.workman@ars.usda.gov.

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using the BBDuk plugin (version 35.82) in Geneious. Reads greater than 1,000 nucleotides in length were assembled *de novo* with the Geneious assembler. All reads were then mapped to the *de novo* assembly, and the resulting consensus sequence was reported. The genome sizes of USMARC-200303013-1 and USMARC-199906011-2 were 9,206 and 9,185 nucleotides, with mean read lengths of 609 and 1,099 nucleotides, mapped read counts of 7,404 and 3,974, and mean read coverages of 617- and 721-fold, respectively. The complete genome sequences share 87% sequence identity and represent the first complete genome sequences for U.S. SRLV strains belonging to genotype A2.

Accession number(s). The sequences of USMARC-200303013-1 and USMARC-199906011-2 are deposited in GenBank/DDBJ/ENA under the accession numbers KY358787 and KY358788, respectively.

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REFERENCES

- 1. Blacklaws BA. 2012. Small ruminant lentiviruses: immunopathogenesis of visna-maedi and caprine arthritis and encephalitis virus. Comp Immunol Microbiol Infect Dis 35:259–269. https://doi.org/10.1016/j.cimid.2011.12
- Shah C, Böni J, Huder JB, Vogt HR, Mühlherr J, Zanoni R, Miserez R, Lutz H, Schüpbach J. 2004. Phylogenetic analysis and reclassification of caprine and ovine lentiviruses based on 104 new isolates: evidence for regular sheep-to-goat transmission and worldwide propagation through livestock trade. Virology 319:12–26. https://doi.org/10.1016/j.virol.2003.09 .047.
- 3. Shah C, Huder JB, Böni J, Schönmann M, Mühlherr J, Lutz H, Schüpbach J. 2004. Direct evidence for natural transmission of small-ruminant lentiviruses of subtype A4 from goats to sheep and vice versa. J Virol 78: 7518–7522. https://doi.org/10.1128/JVI.78.14.7518-7522.2004.
- Ramírez H, Reina R, Amorena B, de Andrés D, Martínez HA. 2013. Small ruminant lentiviruses: genetic variability, tropism and diagnosis. Viruses 5:1175–1207. https://doi.org/10.3390/v5041175.
- Heaton MP, Clawson ML, Chitko-Mckown CG, Leymaster KA, Smith TP, Harhay GP, White SN, Herrmann-Hoesing LM, Mousel MR, Lewis GS, Kalbfleisch TS, Keen JE, Laegreid WW. 2012. Reduced lentivirus suscepti-

- bility in sheep with TMEM154 mutations. PLoS Genet 8:e1002467. https://doi.org/10.1371/journal.pgen.1002467.
- Leymaster KA, Chitko-McKown CG, Clawson ML, Harhay GP, Heaton MP. 2013. Effects of TMEM154 haplotypes 1 and 3 on susceptibility to ovine progressive pneumonia virus following natural exposure in sheep. J Anim Sci 91:5114–5121. https://doi.org/10.2527/jas.2013-6663.
- Leymaster KA, Chitko-McKown CG, Heaton MP. 2015. Incidence of infection in 39-month-old ewes with TMEM154 diplotypes "1 1," "1 3," and "3 3" after natural exposure to ovine progressive pneumonia virus. J Anim Sci 93:41–45. https://doi.org/10.2527/jas.2014-8553.
- Sider LH, Heaton MP, Chitko-McKown CG, Harhay GP, Smith TP, Leymaster KA, Laegreid WW, Clawson ML. 2013. Small ruminant lentivirus genetic subgroups associate with sheep TMEM154 genotypes. Vet Res 44:64. https://doi.org/10.1186/1297-9716-44-64.
- Clawson ML, Redden R, Schuller G, Heaton MP, Workman A, Chitko-McKown CG, Smith TP, Leymaster KA. 2015. Genetic subgroup of small ruminant lentiviruses that infects sheep homozygous for TMEM154 frameshift deletion mutation A4Δ53. Vet Res 46:22. https://doi.org/10 .1186/s13567-015-0162-7.

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