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**THE GENUS *ARNOLDIOLA* (DIPTERA: CECIDOMYIIDAE) IN THE  
NEARCTIC REGION, WITH NEW SYNONYMIES AND COMBINATIONS**

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*Abstract.*—Six nominal species of gall midges (Diptera: Cecidomyiidae) originally assigned to four separate genera and recognized here as three valid species are transferred to *Arnoldiola* Strand. They are as follows: *Arnoldiola azaleae* (Felt 1907a), **new combination** (from *Oligotrophus* Latreille); *Arnoldiola caudata* (Felt 1915), **new combination** (from *Phytophaga* Rondani) and **new synonym** of *A. azaleae*; *Arnoldiola brevicornis* (Felt 1907a), **new combination** (from *Janetiella* Kieffer); *Arnoldiola tiliacei* (Felt 1907a), **new combination** (from *Janetiella*) and **new synonym** of *A. brevicornis*; *Arnoldiola castaneae* (Felt 1909), **new combination** (from *Rhopalomyia* Rübsaamen); and *Arnoldiola ligni* (Felt 1915), **new combination** (from *Janetiella*) and **new synonym** of *A. castaneae*. Diagnostic characters of the genus are outlined, and the newly combined species are described with some characters illustrated.

*Key Words:* gall midges, Holarctic, oak, chestnut

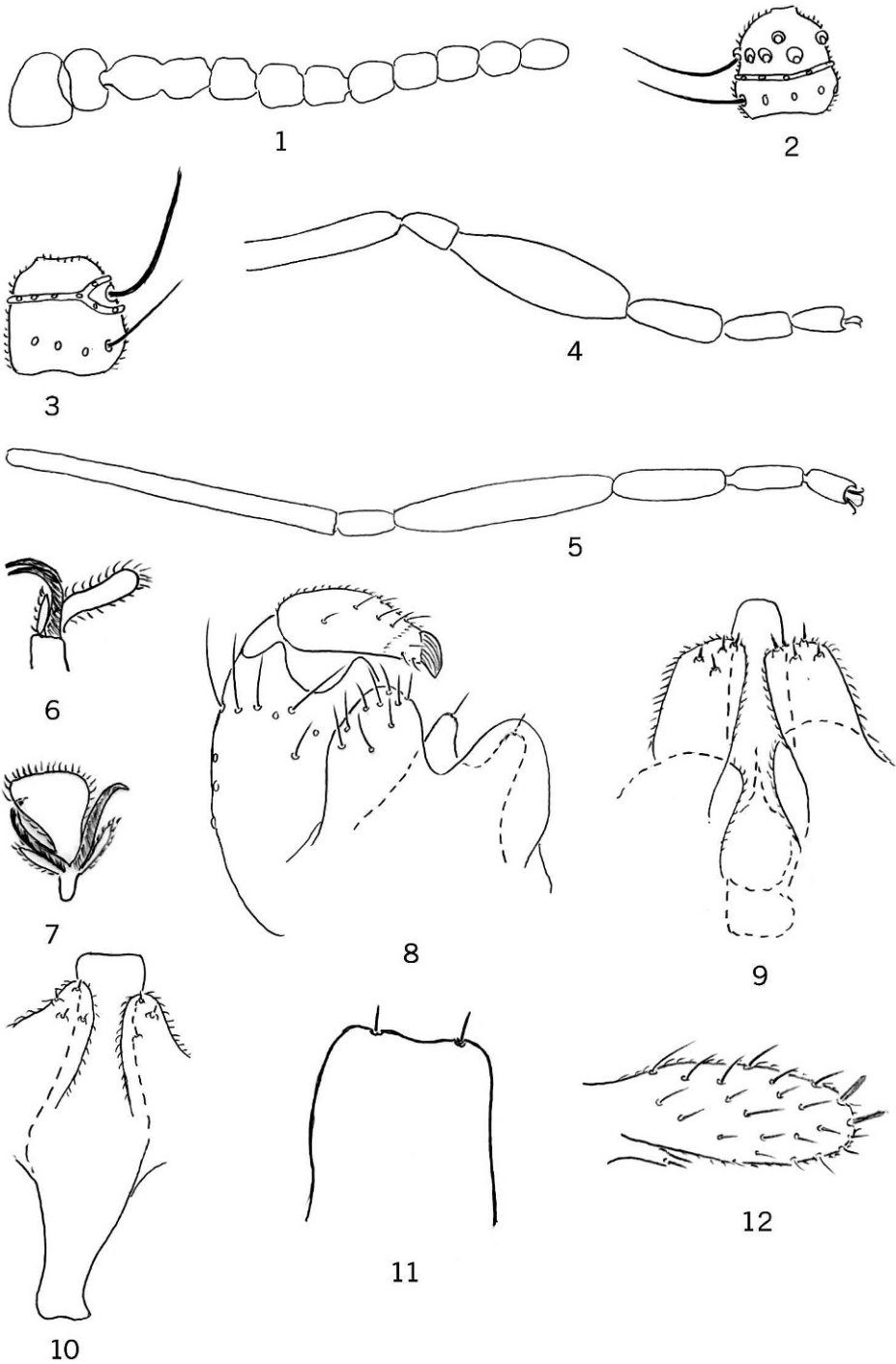
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*Arnoldiola* Strand has long been known from oaks in the Palearctic Region but was only recently noted in the Nearctic Region by Gagné and Riley (1999) with the discovery of *Arnoldiola atra* Gagné on live oak in Texas. The suggestion was made in that article that *A. atra* was a new introduction into the United States because its serious and widespread damage to oak buds in Texas had never previously been remarked upon. While constructing a key to genera of Cecidomyiidae in the Nearctic Region, I found six previously described species that also belong in *Arnoldiola*. Three were in *Janetiella* Kieffer, and the other three were “Unplaced Cecidomyiinae” of Gagné (2004). All were named by Felt and originally variously placed in

four separate genera. Only one of them, *Rhopalomyia castaneae* Felt, had been reared from definite plant damage. Their presence in North America and the fact that one was reared from *Castanea dentata* (Marsh.) Borkh. (Fagaceae) indicates that the genus is native across much of the Holarctic, and its host range includes chestnut as well as oak. *Arnoldiola* otherwise comprises 11 Palearctic species. Seven of those 11, including the type species, *Arnoldiola quercus* Binnie from Europe, are associated with galls on oak (Gagné 2004). I do not know whether those not connected with Fagaceae properly belong in the genus.

*Arnoldiola* was previously distinguished from other Dasineurini by the foreshortened antennal flagellomeres in both sexes (Figs. 1–3). Another consistent character noticed here, on all

\* Accepted by David R. Smith



Figs. 1–12. *Arnoldiola* spp. 1, Male antenna, *A. brevicornis*. 2, Third male flagellomere, dorsal, *A. brevicornis*. 3, Third female flagellomere, ventral, *A. ligni*. 4, Foretarsus and part of tibia, *A. caudata*. 5, Foretarsus and tibia, *A. brevicornis*. 6, Acropod, lateral, *A. brevicornis*. 7, Acropod, dorsal, *A. brevicornis*. 8, Male genitalia, only one gonopod shown, dorsal, *A. brevicornis*. 9, Aedeagus and mediobasal lobes of gonocoxites, ventral, *A. brevicornis*. 10, Aedeagus and mediobasal lobes of gonocoxites, ventral, *A. caudata*. 11, Male hypoproct, dorsal, *A. caudata*. 12, Female fused cerci and hypoproct, lateral, *A. ligni*.

Nearctic species and specimens of at least two Palearctic species, including the type species, is the elongate, cylindrical pulvilli that reach to midlength of the tarsal claws (Figs. 6–7). An additional character in males of the Nearctic species, at least, is the broad tarsi that are noticeably wider than the femora and tibiae (Figs. 4–5). The six nominal species newly combined here with *Arnoldiola* differ from the Palearctic species and *A. atra* Gagné from Texas in having simple rather than toothed tarsal claws. Whether gall midge genera have toothed or simple tarsal claws is usually a good generic character, although there are exceptions where other character combinations indicate its intrageneric variation (e.g., *Mayetiola* Kieffer, *Rabdophaga* Westwood, and now *Arnoldiola*).

#### MATERIALS AND METHODS

The type specimens of each of the species studied for this research are slide-mounted and part of the Felt Collection, currently on long-term loan to the Systematic Entomology Laboratory from the New York State Museum in Albany. Some of the eight individual specimens lack certain body parts or are improperly displayed but are exhibited adequately to support the conclusions of this article. Terminology for adult morphology follows McAlpine et al. (1981).

#### TAXONOMY

The three valid species, reduced from six nominal ones, that are treated here share the following characters, many that are also shared by *Dasineura* and other Dasineurini: eye facets circular, contiguous on the ventral two-thirds and dorsal fourth of the eye but separated by 1 to 1 1/2 facet diameters between those two parts and at the vertex (Gagné and Riley 1999, fig. 4); 10 cylindrical flagellomeres without necks in both sexes (Figs. 1–3); wing (Gagné and Riley 1999, fig. 11): C broken posteriad of its

junction with R5; R5 attaining C anterior to wing apex, Cu forked, M not traceable; male tarsi appreciably wider than tibiae and femora; acropods (Figs. 6–7): tarsal claws simple, empodia enlarging from narrow base to broad apex, slightly longer than claws, pulvilli cylindrical, 1/2 length of claws; first through seventh abdominal tergites (Gagné and Riley 1999, figs. 13–14) of male and first through sixth of female with single row of sparse posterior setae, no lateral setae, scales covering at least posterior half, and a pair of anterior trichoid sensilla; male eighth tergite with anterior pair of trichoid sensilla the only vestiture; female seventh and eighth tergites (Gagné and Riley 1999, fig. 13) rectangular, subequal, much narrower than sixth, not longitudinally divided, with setae posteriorly and with anterior pair of trichoid sensilla; male genitalia (Figs. 8–11): hypoproct weakly to deeply concave apically; gonostylus short, conical, mostly setulose, parameres broad at apex, shorter than aedeagus; ovipositor (Gagné and Riley 1999, fig. 13) protrusible, elongate, the fused cerci (Fig. 12) elongate, cylindrical, setulose and setose, with two wide, blunt-tipped setae near apex; hypoproct with two distal setae.

#### *Arnoldiola azaleae* (Felt), new combination

*azaleae* Felt 1907a: 26 (*Oligotrophus*); Felt 1915: 200, redescription; Gagné 2004: 274 (unplaced Cecidomyiinae).  
*caudata* Felt 1915: 199 (*Phytophaga*); Gagné 2004: 275 (unplaced Cecidomyiinae). **New combination, new synonym.**

The holotype male of *A. azaleae* was swept from an azalea on May 18, 1906 at Albany, NY. Of the four nominal species assembled in this paper that are known from the male sex, *A. caudata* was the only one for which Felt (1907a, 1915) noted the swollen tarsi. The name

*azaleae* is unfortunate because it implies that this species is somehow connected with azalea. *Arnoldiola caudata* is known from three males swept on May 9, 1910 from wild geranium, *Geranium maculatum*, in Albany, NY.

*Arnoldiola azaleae* and *A. caudata* are synonymized here because their genitalia are identical so far as can be determined. The genitalia of three of the four available specimens are not ideally displayed for comparison of all their parts, but visible on all four is a barely concave posterior margin of the hypoproct (Fig. 11). Unfortunately, none of the four specimens shows the mediobasal gonocoxal lobes very well. Those drawn here (Fig. 10) appear narrower than in *A. brevicornis*, but that could be an artifact of mounting. The conspicuously broad tarsi of this species differ from the more moderately enlarged tarsi of *A. brevicornis* (cf. Figs. 4–5). Additionally, the latter species has a deeply lobed instead of a barely concave hypoproct (cf. Figs. 8, 11).

*Arnoldiola brevicornis* (Felt),  
**new combination**

*brevicornis* Felt 1907a: 25 (*Oligotrophus*);  
Felt 1915: 219, redescription (*Janetiella*).

*tiliacei* Felt 1907a: 25 (*Oligotrophus*);  
Felt 1907b: 121, as *tiliaceus*, unjust.  
emend. of *tiliacei*; Felt 1915: 218,  
redescription (*Janetiella*). **New combination, new synonym.**

The holotype male of *A. brevicornis* was taken on goldenrod on June 14, 1906 in Nassau, NY. The holotype male of *A. tiliacei* was taken on basswood on May 23, 1906 in Westfield, NY. Felt did not remark upon the swollen tarsi on either nominal species.

*Arnoldiola brevicornis* and *A. tiliacei* are synonymized here because their genitalia (Figs. 8–9) are identical. This species differs from *A. azaleae* in having a moderately divided posterior margin of

the hypoproct (cf. Figs. 8, 11) and less inflated tarsi (Fig. 5).

*Arnoldiola castaneae* (Felt),  
**new combination.**

*castaneae* Felt 1909: 291 (*Rhopalomyia*);  
Felt 1915: 278, redescription; Gagné  
2004: 275 (unplaced Cecidomyiinae).  
*ligni* Felt 1915: 220 (*Janetiella*). **New combination, new synonym.**

The holotype female of *A. castaneae* was reared on June 13, 1908 from larvae collected at Stowe, Massachusetts the previous year from a burrlike gall (cf. Gagné 1989) on the basal part of the leaf petiole of American chestnut, *Castanea dentata* (Fagaceae). The holotype female of *A. ligni* was reared May 8, 1911 in Nassau, NY from decaying chestnut bark. The larva of this female presumably fell from leaves as it left the galls the previous year. The two specimens under this name are similar in every way, including in the shape of the fused cerci (Fig. 12). The abdomen is similar otherwise to that drawn for *A. atra* (Gagné and Riley 1999, fig. 13). *Arnoldiola castaneae* may be synonymous with either *A. azaleae* or *A. brevicornis*, but one cannot know without rearing both sexes from chestnut galls.

DISCUSSION

The species treated here will key to couplet 182 in Gagné (1981) where one will find a choice between *Janetiella* Kieffer and *Mayetiola*. *Arnoldiola* differs from those genera by the foreshortened, neckless antennal flagellomeres in both sexes, the pulvilli being half the length of the tarsal claws instead of one-third or less, and the male tarsi being appreciably wider than the tibiae or femora.

Swollen tarsi or other leg parts are uncommon in Cecidomyiidae. Other than in *Arnoldiola*, the only other example I can think of is *Trotteria* Kieffer, which has greatly swollen femora as

compared to the rest of the leg. I wondered if the pupal leg sheaths of *Arnoldiolo* were enlarged around the tarsi, but the leg sheaths of *A. atra* are of equal width throughout, which means that while the femora and tibiae lengthen and thin out following ecdysis, the tarsi do not or do so to a relatively lesser extent. This makes sense because, were only the tarsal part of the leg sheath relatively large, the tarsi might have difficulty being drawn through the thinner part of the leg sheath during ecdysis.

Pulvilli are not always noted in descriptions of Cecidomyiidae, perhaps because they are usually not prominent, only one-third or less the length of the claws and often obscured when the claws are drawn in side view. However, they can be a useful generic character as they are here.

Empodia are often described as longer than the claws, but in many cases the claws are splayed outward (cf. Fig. 7) and only appear shorter. With six legs on a good specimen, one hopes to have some acropods that will be well enough displayed to show the relative lengths well.

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