

2002

# Molecular Evidence for a Species Complex in the Genus *Aphelinus* (Hymenoptera: Aphelinidae), with Additional Data on Aphidiine Phylogeny (Hymenoptera: Braconidae)

Yi Chen

*Department of Entomology and Plant Pathology, Oklahoma State University, Stillwater, OK 74078-3033*

Kristopher L Giles

*Department of Entomology and Plant Pathology, Oklahoma State University, Stillwater, OK 74078-3033*

Matthew H. Greenstone

*Plant Science and Water Conservation Research Laboratory, USDA-ARS, 1301 N. Western, Stillwater, OK 74075*

Follow this and additional works at: <http://digitalcommons.unl.edu/entomologyother>



Part of the [Entomology Commons](#)

---

Chen, Yi; Giles, Kristopher L; and Greenstone, Matthew H., "Molecular Evidence for a Species Complex in the Genus *Aphelinus* (Hymenoptera: Aphelinidae), with Additional Data on Aphidiine Phylogeny (Hymenoptera: Braconidae)" (2002). *Entomology Papers from Other Sources*. 89.

<http://digitalcommons.unl.edu/entomologyother/89>

This Article is brought to you for free and open access by the Entomology Collections, Miscellaneous at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Entomology Papers from Other Sources by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

# Molecular Evidence for a Species Complex in the Genus *Aphelinus* (Hymenoptera: Aphelinidae), with Additional Data on Aphidiine Phylogeny (Hymenoptera: Braconidae)

YI CHEN,<sup>1</sup> KRISTOPHER L GILES,<sup>1</sup> AND MATTHEW H. GREENSTONE<sup>2,3</sup>

Ann. Entomol. Soc. Am. 95(1): 29–34 (2002)

**ABSTRACT** Mitochondrial 16s rDNA was sequenced from nine different populations of Aphelinidae and 10 of aphidiinae. Sequence divergences between populations within a species are low, ranging from 0 to 0.38%. Divergences among species within the same genus range from 0 to 8.71%. *Aphelinus asychis* Walker has a sequence divergence from the other *Aphelinus* spp. of 8.71%, which is even higher than the divergences among the aphidiine genera *Lysiphlebus*, *Aphidius* and *Diaeretiella*. Our 16s rDNA phylogeny for aphidiines is concordant with a previously published NADH1 dehydrogenase phylogeny. Our analysis identifies a complex comprising *A. albipodus* (Hayat & Fatima), *A. varipes* (Foerster), and *A. hordei* (Kurdjumov). The sequence divergences within this complex are low, ranging from 0 to 0.38% with an average of 0.19%, which suggests that the three species within the *A. varipes* complex diverged within the last  $\approx 90,000$  yr.

**KEY WORDS** *Aphelinus*, Aphelinidae, Braconidae, aphidiinae, 16s rDNA, phylogeny

THE MINUTE PARASITIDS of the genus *Aphelinus* are important biological control agents of aphids worldwide. During the last four decades, four species, *A. albipodus* (Hayat & Fatima), *A. asychis* (Walker), *A. varipes* (Foerster), and *A. hordei* (Kurdjumov), have been introduced into the United States to control several aphid species (van den Bosch et al. 1959, Jackson et al. 1970, Jackson and Eikenbary 1971, Hopper et al. 1998, Prokrym et al. 1998a, Zhu et al. 2000). Additional taxa, which may be synonymous with one or more of these species, have been reported from North America (Mackauer and Finlayson 1967, Lajeunesse and Johnson 1991, Yu 1992). Also, the *A. asychis* individuals released in the United States comprise a group of geographical isolates that may constitute a sibling species complex (Kazmer et al. 1996). These difficulties in determining taxonomic and geographic affinities of aphelinids exerting cereal aphid biological control make it difficult to evaluate the effectiveness of the biological control programs through which they were introduced (Hopper et al. 1998, Prokrym et al. 1998a).

The taxonomy of these species is also made difficult by their small size ( $\leq 1.5$  mm long) and the paucity

of clear morphological differences among them (Prokrym et al. 1998b). Several biochemical approaches, including allozyme electrophoresis (Walton et al. 1990), random amplified polymorphic DNA markers (Kazmer et al. 1995), microsatellite DNA variability (Vanlerberhe-Masutti and Chavigny 1997), and ITS-2 polymerase chain reaction (PCR) (Zhu and Greenstone 1999, Zhu et al. 2000), have been used to differentiate *Aphelinus* spp.

Ribosomes are a part of the translational machinery of the cell. Thus, the sequences and structures of rRNAs are conserved in the course of evolution due to their importance in the growth, function and survival of cells (Gutell 1992). Classical analysis of complete and partial sequences of rRNA has revealed that the primary structure of rRNAs consists of highly conserved regions interspersed with regions of moderate to low homology within related species (Gopo et al. 1988). The small size of 5S rRNA and extensive secondary structures in the 23S rRNA render these molecules unsuitable for sequencing and further analysis. However, the 16s rRNA has been found to be specific to a given species. The moderate size of the 16s rRNA, range of evolutionary rates across the sequence, and confirmed importance in systematics make 16s rRNA particularly useful for phylogenetic studies (Simon et al. 1994, Raghava et al. 2000).

As part of an ongoing program to develop molecular markers to separate and identify aphid endoparasitoids (Zhu and Greenstone 1999, Zhu et al. 2000), we have sequenced a segment of the mitochondrial 16s rDNA gene for these four *Aphelinus* species. For comparative purposes, we also sequenced a less problem-

This article reports the results of research only. Mention of a proprietary product does not constitute an endorsement or a recommendation by the USDA for its use.

<sup>1</sup> Department of Entomology and Plant Pathology, Oklahoma State University, Stillwater, OK 74078–3033.

<sup>2</sup> Plant Science and Water Conservation Research Laboratory, USDA-ARS, 1301 N. Western, Stillwater, OK 74075.

<sup>3</sup> Current address: American Institute of Biological Sciences, 2404 Northwood Lane, Edmond, OK 73013 (e-mail: mgreenstone@aibs.org).

Table 1. Taxa included in the study

Species name	abbreviation	Locality or reference
<i>Aphelinus asychis</i> Walker	aaska	Kazakhstan
<i>Aphelinus asychis</i> Walker	aasmp	Montpellier, France
<i>Aphelinus albipodus</i> Hayat & Fatima	aal	Tacheng, China
<i>Aphelinus varipes</i> (Foerster)	avalj	Lattes-Juvignac, France
<i>Aphelinus varipes</i> (Foerster)	avamp	Montpellier, France
<i>Aphelinus hordei</i> (Kurdjumov)	aho	Odessa, Ukraine
<i>Aphytis melinus</i> DeBach	ame	Downton & Austin 1994
<i>Encarsia formosa</i> Gahan	efo	Downton & Austin 1994
<sup>a</sup> <i>Pteromalus puparum</i> (L.)	ppu	Downton & Austin 1994
<i>Diaeretiella rapae</i> (M'Intosh)	draco	Colfax, WA, USA
<i>Diaeretiella rapae</i> (M'Intosh)	drauc	Riverside, CA, USA
<i>Lysiphlebus testaceipes</i> (Cresson)	ltesw	Stillwater, OK, USA
<i>Lysiphlebus testaceipes</i> (Cresson)	ltepu	Pullman, WA, USA
<i>Lysiphlebus testaceipes</i> (Cresson)	ltech	Chelan, WA, USA
<i>Lysiphlebus testaceipes</i> (Cresson)	lteel	Ellensburg, WA, USA
<i>Aphidius colemani</i> Viereck	acoch	Chile
<i>Aphidius colemani</i> Viereck	acook	Stillwater, OK, USA
<i>Aphidius ervi</i> Haliday	aer	Prosser, WA, USA
<i>Aphidius matricariae</i> Haliday	ama	Prosser, WA, USA
<i>Aphidius ervi</i> Haliday	aerjb	Belshaw et al. 2000
<i>Aphidius rosae</i> Haliday	aro	Downton et al. 1998
<i>Pseudephedrus chilensis</i> (Howard)	pch	Belshaw et al. 2000
<i>Dyscritulus planiceps</i> (Marshall)	dpl	Belshaw et al. 2000
<i>Ephedrus plagiator</i> (Nees)	epl	Belshaw et al. 2000
<i>Paralipsis enervis</i> (Nees)	pen	Belshaw et al. 2000
<i>Lipolexis gracilis</i>	lgr	Kambhampati et al. (GenBank accession No. AF174338)
<sup>a</sup> <i>Hormius</i> sp.	hsp	Belshaw et al. 2000

<sup>a</sup> *Pteromalus puparum* was used as outgroup for Aphelinidae, and *Hormius* sp. for Aphidiina.

atic group of aphid parasitoids of the braconid subfamily aphidiinae (GenBank accession numbers AF289130 to AF289148). Here we present phylogenies based on these sequences, and use estimates of relationships and divergence times among taxa to infer the systematic status of the aphelinid taxa.

### Materials and Methods

**Insects.** Most of the sequences used in this research were derived from parasitoids that were either received preserved in 95% ethanol or were removed from established colonies in the laboratory; additional sequences were derived from the literature. Colonies were reared by previously described protocols (Reed et al. 1991), and maintained in cages in a Conviron model I23 incubator (Controlled Environments, Pembina, ND) at 20°C and a photoperiod of 16:8 (L:D) h. To reduce the risk of contamination, only one parasitoid colony was maintained at one time. The collecting localities or literature sources of all populations used in this research are given in Table 1.

**DNA Extraction.** We modified the methods of Zhu and Greenstone (1999) to extract total DNA. Insects were placed individually in 1.5-ml microcentrifuge tubes and homogenized using a battery-powered homogenizer (Midwest Scientific, St. Louis, MO) in 100  $\mu$ l of isolation buffer containing 0.1 M NaCl, 0.2 M sucrose, 0.1 M Tris-HCl (pH 9.1), 0.05 M EDTA, 1% SDS, and 20  $\mu$ g/ml RNase A. The homogenate was vortexed briefly and incubated for 30 min at 65°C. The supernatant was transferred to a new tube and extracted once with one volume of chloroform/isoamyl

alcohol (24:1). One-tenth volume of 3.0 M sodium acetate and two volumes of ice-cold 100% EtOH were added to the tube. DNA was then pelleted by centrifugation and resuspended in 200  $\mu$ l of distilled water.

**PCR Amplification, Purification, and Sequencing of 16s rDNA.** Portions of the mitochondrial 16s rDNA gene were amplified by PCR using primers designed by Downton and Austin (1994). The primers used were 16SWb (5'-CACCTGTTTATCAAAAA-CAT-3'), which anneals to nucleotides 13924–13943, and 16SWa (5'-CGTCGATTTGAACTCAAATC-3'), which anneals to nucleotides 13392–13411 of the honey bee mitochondrial genome (Crozier and Crozier 1993). Where amplifications were unsuccessful under a variety of conditions, the primer 16S.Sh (5'-AGATTTTAAAAGTTCGAACAG-3'), which anneals to nucleotides 13417–13436 of the honey bee genome, was used in place of 16SWa. The PCR product from the 16s rDNA gene was thus in the range of 530–550 bp.

PCR reactions (25  $\mu$ l) contained 10 mM Tris-HCl pH 9.0, 1.5 mM MgCl<sub>2</sub>, 1.0  $\mu$ M of each primer, 50 mM KCl, 0.1 mM of each dNTP, 0.05 U/ $\mu$ l of *Taq* DNA polymerase (Promega, Madison, WI), and 2  $\mu$ l of DNA template, and were performed in a PTC-100 thermocycler (MJ Research, Watertown, MA). DNA was denatured for 3 min at 94°C, followed by 35 amplification cycles comprising 30 s denaturing at 94°C, 30 s annealing at 50°C, and 1 min extension at 72°C. DNA was finally extended for 2 min at 72°C after amplification.

**Table 2.** 16s rDNA sequence divergence (%) (above the diagonal) and number of differences (below the diagonal) based on pairwise comparisons among aphid parasitoid populations

	ltepu	ltech	lteel	ltesw	drauc	draco	acoch	acook	aer	ama	aaska	aasmp	avalj	avamp	aho	aalca
1. ltepu		0	0	0.38	4.58	4.58	5.17	5.17	5.56	5.75	24.38	24.38	25.24	25.24	25.05	25.05
2. ltech	0		0	0.38	4.58	4.58	5.17	5.17	5.56	5.75	24.38	24.38	25.24	25.24	25.05	25.05
3. lteel	0	0		0.38	4.58	4.58	5.17	5.17	5.56	5.75	24.38	24.38	25.24	25.24	25.05	25.05
4. ltesw	2	2	2		4.96	4.96	5.53	5.53	5.94	6.13	24.38	24.38	25.19	25.19	25.38	25.38
5. drauc	24	24	24	26		0	3.26	3.26	2.68	3.26	24.00	24.00	25.05	25.05	24.86	24.86
6. draco	24	24	24	26	0		3.26	3.26	2.68	3.26	24.00	24.00	25.05	25.05	24.86	24.86
7. acoch	27	27	27	29	17	17		0	3.64	4.41	24.57	24.57	25.05	25.05	24.86	24.86
8. acook	27	27	27	29	17	17	0		3.64	4.41	24.57	24.57	25.05	25.05	24.86	24.86
9. aer	29	29	29	31	14	14	19	19		1.72	24.57	24.57	26.01	26.01	25.82	25.82
10. ama	30	30	30	32	17	17	23	23	9		25.05	25.05	26.40	26.40	26.20	26.20
11. aaska	127	127	127	127	126	126	128	128	128	130		0.19	8.71	8.71	8.71	8.71
12. aasmp	127	127	127	127	126	126	128	128	128	130	1		8.71	8.71	8.71	8.71
13. avalj	131	131	131	132	130	130	130	130	135	137	46	46		0	0.38	0.19
14. avamp	131	131	131	132	130	130	130	130	135	137	46	46	0		0.38	0.19
15. aho	130	130	130	133	129	129	129	129	134	136	46	46	1	1		0
16. aal	130	130	130	133	129	129	129	129	134	136	46	46	1	1	0	

PCR products were separated on a 1% low melting point agarose gel. DNA fragments were sliced from the gel and extracted using a wizard PCR Preps DNA Purification System (Promega). Purified DNA fragments were directly sequenced from both directions using an automated sequencer located at the Recombinant DNA/Protein Resource Facility, Oklahoma State University, Stillwater.

**Phylogenetic Analysis.** The amplified 16s rDNA sequences were confirmed by similarity searching of the GenBank database using the Fasta and Blast programs in the GCG Wisconsin Package. We aligned 16s rDNA sequences by using SeqLab and Pileup of the GCG Wisconsin Package (Unix version 10, Genetics Computer Group, Madison, WI), and confirmed alignment by visual inspection. MEGA program version 1.01 (Kumar et al. 1993) was used to estimate evolutionary distances and compute basic statistical quantities.

Phylogenetic analyses were done using neighbor-joining, parsimony, and maximum likelihood methods with the software PAUP (Swofford 1997). *Pteromalus puparum* (L.) (Hymenoptera: Pteromalidae) was used as the outgroup for Aphelinidae, and *Hormius* sp. (Hymenoptera: Braconidae) for aphidiinae. To find minimum-length phylogenetic trees, heuristic parsimony searches (Hillis et al. 1996) were performed using 100 replicates of random additions with the tree bisection and reconnection option for branch swapping. The stepwise addition option was used to create an initial tree for the heuristic search. Random number seed was reset for random resolution of polytomies during the swapping procedure. Maximum likelihood trees using nucleotide data were generated under the Hasegawa-Kishino-Yano substitution model (Hasegawa et al. 1985) using observed base frequencies. Transition/transversion ratios estimated by PAUP were used in maximum likelihood analyses. Heuristic searches were used with 10 times of random additions and the tree bisection and reconnection option. To assess the support of clades in the phylogenetic trees, the bootstrap test (Felsenstein 1985) was performed

with 1000 replications. The trees were visualized by Treeview (version 1.6.1; Page 1996).

**Voucher Specimens.** Voucher specimens of *A. hordei* and all of the aphidiine species have been deposited in the USDA-ARS-PSWCRL Cereal Insect Genetics Resource Library, Stillwater, OK. Voucher information for the other aphelinid species was reported previously (Zhu and Greenstone 1999).

## Results

**Sequence Data and Divergence.** The 16s rDNA PCR products ranged from 524 to 531 bp; the aligned sequences were 534 bp long due to the introduction of gaps among sequences. Of 534 total characters analyzed, 363 (67.98%) are constant, and the number of parsimony-informative characters is 165 (30.90%). For the *Aphelinus* spp., 340 (63.43%) characters are constant and the number of parsimony-informative characters is 84 (15.67%); for the aphidiines, 164 characters are constant (30.83%), and the number of parsimony-informative characters is 137 (25.75%).

Sequence divergences differ greatly between different pairs of taxa, ranging from 0 to 26.4% (Table 2). Divergences within species are generally low. For four populations of *Lysiphlebus testaceipes* (Cresson), the sequence divergence ranges from 0 to 0.38%. No sequence differences were found within populations of *Diaretiella rapae* (M'Intosh), *Aphidius colemani* Vierck, and *A. varipes*. The sequence divergence between the two populations of *A. asychis* is 0.19%. Sequence divergence among congeneric species is generally higher than within species, ranging from 0 to 8.71%. The divergence among the three *Aphidius* species ranges from 1.72 to 4.41%. The sequence of *A. hordei* differs from that of *A. varipes* by 0.38% and from that of *A. albipodus* by 0.19%.

**Phylogenetic Relationships.** For the aphelinids, six equally parsimonious trees were constructed and from these a consensus tree, using *P. puparum* as the outgroup, was generated (Fig. 1). All *Aphelinus* species

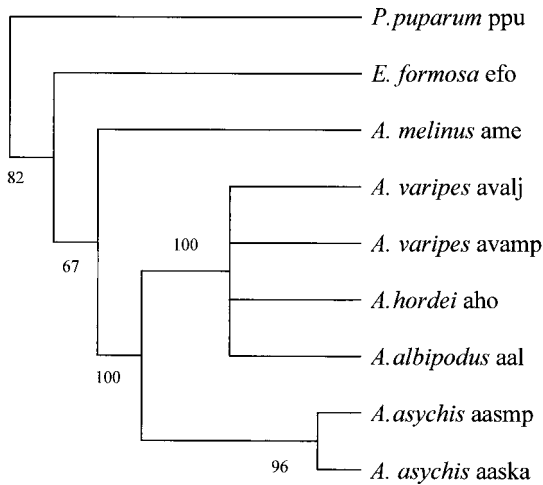


Fig. 1. Strict consensus of six equally parsimonious trees resulting from maximum parsimony analysis for aphelinids. Of 536 total characters, 340 characters are constant; 84 are parsimony-informative. Tree length = 249, consistency index (CI) = 0.8394, retention index (RI) = 0.7546, rescaled consistency index (RC) = 0.6334. Numbers at branches are bootstrap values with 1,000 bootstrap replications. Abbreviations as in Table 2.

form a monophyletic group with 100% bootstrap value support. Within *Aphelinus*, two clades are obvious, one with two *A. asychis* strains and the other containing the other three species, both with 100% bootstrap value support. These three species are shown as monophyletic.

For aphidiine wasps, a single most parsimonious tree, using *Hormius* sp. as the outgroup, resulted from the parsimony analysis (Fig. 2). All species and strains of *Lysiphlebus*, *Diaeretiella* and *Aphidius* form a monophyletic group. *Diaeretiella* and *Aphidius* are more closely related to each other than to *Lysiphlebus*. *Aphidius* is paraphyletic due to the inclusion of *Diaeretiella*.

For both aphelinids and aphidiines, trees obtained from neighbor-joining and maximum likelihood analyses were very similar to those from the parsimony analysis (data not shown). This indicates strong support for the clades.

### Discussion

Among the parasitoid genera studied here, sequence divergences are high, ranging from 2.68 to 26.4%. *Aphelinus* has the highest sequence divergence, >20% from the other genera. The sequence of *A. asychis* is very different from those of its congeners, diverging from them by 8.71%, which is even higher than the divergence among the aphidiid genera *Lysiphlebus*, *Aphidius*, and *Diaeretiella*, which range from 2.68 to 6.13%. The 16s rDNA sequences of *A. varipes*, *A. hordei*, and *A. albipodus* are sufficiently similar to suggest that they belong either to the same species or to a species complex distinguishable from *A. asychis*.

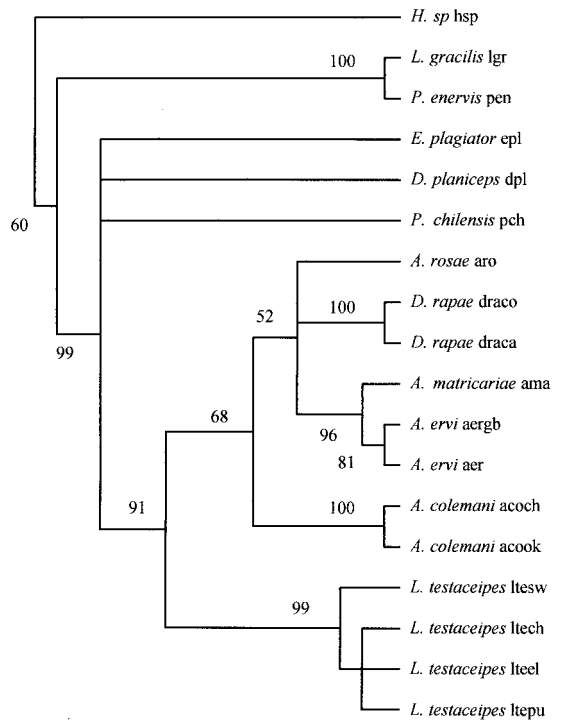


Fig. 2. Single most parsimonious tree yielded by maximum parsimony analysis for aphidiine wasps. Of 532 total characters, 164 are constant, 137 are parsimony-informative. Tree length = 644, CI = 0.7966, RI = 0.6042, RC = 0.4813. Numbers and abbreviations as in Fig. 1.

Here we conservatively consign all of the individuals from the *A. albipodus*, *A. varipes*, and *A. hordei* populations studied here to an *A. varipes* species complex.

Our aphidiine phylogeny supports the NADH 1 dehydrogenase gene phylogeny of Smith et al. (1999), in which *Diaeretiella* also falls inside *Aphidius*. *Aphidius ervi* Haliday and *A. matricariae* Haliday are closely related to one other, and *A. colemani* is a basal taxon within the *Aphidius* and *Diaeretiella* complex. A 28S D2 phylogeny (Belshaw et al. 2000) indicated that the genus *Lysiphlebus* is polyphyletic.

If we assume a rate of divergence for mtDNA of 2% per million years (Myr) as in *Drosophila* (DeSalle et al. 1987), we find that the average sequence divergence between *A. asychis* and the members of the *A. varipes* complex corresponds to a time of 4.36 Myr. Using the same rate, the average sequence divergence within the *A. varipes* complex, 0.19%, corresponds to only 90,000 yr. The lack of sequence variation in 16s rDNA between *A. hordei* and *A. albipodus* may indicate that they belong to the same species.

### Acknowledgments

We thank Dan Gonzáles, Keith Hopper, Doug Jones, Keith Pike, Goddy Prinsloo, Rick Turcotte, and Marcella Waggoner for parasitoid stocks or alcoholic specimens; John Cushman and Brian Jones for technical assistance; and Robert Belshaw,



Doug Bergman, Greta Binford, Jiang Haobo, Michael Schauff, and Jim Woolley for comments on the manuscript. Y.C. was supported by an Oklahoma State University Postdoctoral Research Fellowship. This work was approved for publication by the director of the Oklahoma Agricultural Experiment Station, Stillwater, OK.

### References Cited

- Belshaw, R., M. Dowton, D. L. Quicke, and A. D. Austin. 2000. Estimating ancestral geographical distributions: a Gondwanan origin for aphid parasitoids? *Proc. R. Soc. Lond. B Biol. Sci.* 267: 491–496.
- Crozier, R. H., and Y. C. Crozier. 1993. The mitochondrial genome of the honeybee *Apis mellifera*: complete sequence and genome organization. *Genetics* 133: 97–117.
- DeSalle, R., T. Freedman, E. M. Prager, and A. C. Wilson. 1987. Tempo and mode of sequence evolution in mitochondrial DNA of Hawaiian *Drosophila*. *J. Mol. Evol.* 26: 157–164.
- Dowton, M., and A. D. Austin. 1994. Molecular phylogeny of the insect order Hymenoptera: apocritan relationships. *Proc. Natl. Acad. Sci. U.S.A.* 91: 9911–9915.
- Dowton, M., A. D. Austin, and M. F. Aotolin. 1998. Evolutionary relationship among the Braconidae (Hymenoptera: Ichneumonoidea) inferred from partial 16s rDNA gene sequences. *Insect Mol. Biol.* 7: 129–150.
- Felsenstein, J. 1985. Confidence intervals on phylogenies: an approach using the bootstrap. *Evolution* 39: 783–791.
- Gopo, J. M., R. Melis, E. Filipiska, R. Meneveri, and J. Filipiski. 1988. Development of a *Salmonella*-specific biotinylated DNA probe for rapid routine identification of *Salmonella*. *Mol. Cell Probes* 2: 270–271.
- Gutell, R. R. 1992. Evolutionary characteristics of 16s rRNA and 23S rRNA structures, pp. 243–309. *In* H. Hartman and K. Matsuno [eds.], *The origin and evolution of the cell*. World Scientific Publishing, Singapore.
- Hasegawa, M., H. Kishino, and T. Yano. 1985. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J. Mol. Evol.* 22: 160–174.
- Hillis, D. M., C. Moritz, and B. K. Mable (eds.). 1996. *Molecular systematics*, 2nd ed. Sinauer, Sunderland, MA.
- Hopper, K. R., D. Coutinot, K. Chen, D. J. Kazmer, G. Mercadier, S. E. Halbert, R. H. Miller, K. S. Pike, and L. K. Tanigoshi. 1998. Exploration for natural enemies to control *Diuraphis noxia* (Homoptera: Aphididae) in the United States, pp. 166–182. *In* S. S. Quisenberry and F. B. Peairs [eds.], *A response model for an introduced pest—the Russian wheat aphid*. Thomas Say Publications, Entomological Society of America, Lanham, MD.
- Jackson, H. B., L. W. Coles, E. A. Wood, Jr., and R. D. Eikenbary. 1970. Parasites reared from the greenbug and corn leaf aphid in Oklahoma in 1968. *J. Econ. Entomol.* 63: 733–736.
- Jackson, H. B., and R. D. Eikenbary. 1971. Bionomics of *Aphelinus asychis* (Hymenoptera: Eulophidae) an introduced parasite of the sorghum greenbug. *Ann. Entomol. Soc. Am.* 64: 81–85.
- Kazmer, D. J., K. R. Hopper, D. M. Coutinot, and D. G. Heckel. 1995. Suitability of random amplified polymorphic DNA for genetic markers in the aphid parasitoid, *Aphelinus asychis* Walker. *Biol. Control* 5: 503–512.
- Kazmer, D. J., K. Maiden, N. Ramaulde, D. M. Coutinot, and K. R. Hopper. 1996. Reproductive compatibility, mating behavior, and random amplified polymorphic DNA variability in some *Aphelinus asychis* (Hymenoptera: Aphelinidae) derived from the Old World. *Ann. Entomol. Soc. Am.* 89: 212–220.
- Kumar, S., K. Tamura, and M. Nei. 1993. MEGA: Molecular evolutionary genetics analysis, version 1.0. The Pennsylvania State University, University Park, PA.
- Lajeunesse, S. E., and G. D. Johnson. 1991. New North American host records for *Aphelinus* sp. nr. *Varipes* (Foerster) (Hymenoptera: Aphelinidae): the western wheat aphid, *Diuraphis tritici* (Gillette), and the Russian wheat aphid, *Diuraphis noxia* (Mordvilko) (Homoptera: Aphididae). *Can. Entomol.* 123: 413–415.
- Mackauer, M., and T. Finlayson. 1967. The hymenopterous parasites (Hymenoptera: Aphidiidae and Aphelinidae) of the pea aphid in eastern North America. *Can. Entomol.* 99: 1051–1082.
- Page, R.D.M. 1996. TREEVIEW: an application to display phylogenetic trees on personal computers. *Comput. Appl. Biosci.* 12: 357–358.
- Prokrym, D. R., K. S. Pike, and D. J. Johnson. 1998a. Biological control of *Diuraphis noxia* (Homoptera: Aphididae): implementation and evaluation of natural enemies, pp. 183–208. *In* S. S. Quisenberry and F. B. Peairs [eds.], *A response model for an introduced pest—the Russian wheat aphid*. Thomas Say Publications, Entomological Society of America, Lanham, MD.
- Prokrym, D. R., M. E. Schauff, and J. B. Wooley. 1998b. Minutes of taxonomy workshop on Russian wheat aphid parasitoids, Washington, DC, 13–14 January 1998. U.S. Department of Agriculture, APHIS Niles Plant Protection Center.
- Raghava, G.P.S., R. J. Solanki, V. Soni, and P. Agrawal. 2000. Fingerprinting method for phylogenetic classification and identification of microorganisms based on variation in 16s rRNA gene sequences. *BioTechniques* 29: 108–116.
- Reed, D. K., J. A. Webster, B. G. Jones, and J. D. Burd. 1991. Tritrophic relationships of Russian wheat aphid (Homoptera: Aphididae), a hymenopterous parasitoid (*Diaretiella rapae* McIntosh), and resistant and susceptible small grains. *Biol. Control* 1: 35–41.
- Simon, C., F. Frati, A. T. Beckenbach, B. Crespi, H. Liu, and P. Flook. 1994. Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Ann. Entomol. Soc. Am.* 87: 651–701.
- Smith, P. T., S. Kambhampati, W. Volkl, and M. Mackauer. 1999. A phylogeny of aphid parasitoids (Hymenoptera: Braconidae: Aphidiinae) inferred from mitochondrial NADH 1 dehydrogenase gene sequence. *Mol. Phylogenet. Evol.* 11: 236–245.
- Swofford, D. L. 1997. PAUP: Phylogenetic analysis using parsimony (Version 4.0d.54). Smithsonian Institution, Washington, DC.
- Van den Bosch, R. B., E. I. Schlinger, E. J. Dietrick, J. C. Hall, and B. Puttler. 1959. The colonization and establishment of imported parasites of the spotted alfalfa aphid in California. *J. Econ. Entomol.* 52: 136–141.
- Vanlerberhe-Masutti, F., and P. Chavigny. 1997. Characterization of a microsatellite locus in the parasitoid wasp *Aphelinus abdominalis* (Hymenoptera: Aphelinidae). *Bull. Entomol. Res.* 87: 313–318.
- Walton, M. P., H. D. Loxdale, and L. A. Williams. 1990. Electrophoretic keys for the identification of parasitoids (Hymenoptera: Aphelinidae) attacking *Stobion* (F.) (Hemiptera: Aphididae). *Biol. J. Linn. Soc.* 40: 333–346.
- Yu, D. S. 1992. Effects of photoperiod and temperature on diapause of two *Aphelinus* sp. (Hymenoptera: Aphelinidae) parasitizing the Russian wheat aphid. *Can. Entomol.* 124: 853–860.
- Zhu, Y., and M. H. Greenstone. 1999. Polymerase chain reaction techniques for distinguishing three species and

- two strains of *Aphelinus* (Hymenoptera: Aphelinidae) from *Diuraphis noxia* and *Schizaphis graminum* (Homoptera: Aphididae). *Ann. Entomol. Soc. Am.* 92: 71–79.
- Zhu, Y.-C., J. D. Burd, N. C. Elliott, and M. H. Greenstone. 2000. Specific ribosomal DNA markers for early PCR detection of *Aphelinus hordei* (Hymenoptera: Aphelinidae) and *Aphidius colemani* (Hymenoptera: Aphidiidae) from *Diuraphis noxia* (Homoptera: Aphididae). *Ann. Entomol. Soc. Am.* 93: 486–491.
- Received for publication 26 February 2001; accepted 31 August 2001.*
-