Plant MicroRNAs Display Differential 3' Truncation and Tailing Modifications That Are ARGONAUTE1 Dependent and Conserved Across Species

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LARGE-SCALE BIOLOGY ARTICLE

Plant MicroRNAs Display Differential 3’ Truncation and Tailing Modifications That Are ARGONAUTE1 Dependent and Conserved Across Species

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Plant small RNAs are 3’ methylated by the methyltransferase HUA1 ENHANCER1 (HEN1). In plant hen1 mutants, 3’ modifications of small RNAs, including oligo-uridylation (tailing), are associated with accelerated degradation of microRNAs (miRNAs). By sequencing small RNAs of the wild type and hen1 mutants from Arabidopsis thaliana, rice (Oryza sativa), and maize (Zea mays), we found 3’ truncation prior to tailing is widespread in these mutants. Moreover, the patterns of miRNA truncation and tailing differ substantially among miRNA families but are conserved across species. The same patterns are also observable in wild-type libraries from a broad range of species, only at lower abundances. ARGONAUTE (AGO1), even with defective slicer activity, can bind these truncated and tailed variants of miRNAs. An ago1 mutation in hen1 suppressed such 3’ modifications, indicating that they occur while miRNAs are in association with AGO1, either during or after RNA-induced silencing complex assembly. Our results showed AGO1-bound miRNAs are actively 3’ truncated and tailed, possibly reflecting the activity of cofactors acting in conserved patterns in miRNA degradation.

INTRODUCTION

The stability and function of small RNAs (sRNAs) are affected by various modifications, such as methylation, uridylation, adenylation, and RNA editing (Kim et al., 2010; Ji and Chen, 2012). Plant microRNAs (miRNAs) pair with their targets near-perfectly and are 3’ protected by methylation (Voinnet, 2009); by contrast, animal miRNAs are unmethylated and typically partially complementary to their mRNA targets via the short seed region at the 5’ end (Bartel, 2009) and can be truncated and tailed when extensively matched by a target RNA (Ameres et al., 2010).

HEN1 ENHANCER1 (HEN1), first identified in a screen for floral mutants in Arabidopsis thaliana, was later characterized as a methyltransferase that adds a 2’-O-methyl group to the 3’ terminal nucleotide of miRNAs and small interfering RNAs (siRNAs) (Li et al., 2005; Yu et al., 2005). Structural analysis determined that HEN1 specifically recognizes the sRNA duplex as a substrate (Huang et al., 2009). In hen1 mutant backgrounds, miRNAs examined by RNA gel blot analysis exhibit unusual length heterogeneity and the abundance of most miRNAs is considerably reduced (Li et al., 2005; Yu et al., 2005). Comparisons performed by cloning and sequencing individual miRNAs in wild-type and hen1-1 backgrounds showed the addition of predominantly uridine (U) nucleotides at the 3’ end of some miRNAs in a hen1-1 mutant (Li et al., 2005). The U-rich tail of unmethylated miRNAs in Arabidopsis hen1 mutants is added by nucleotidyld transferase HEN1 SUPPRESSOR1 (HESO1), whose mutation can partially rescue the phenotype of hen1 mutants by increasing miRNA accumulation (Ren et al., 2012; Zhao et al., 2012b).

A family of SMALL RNA DEGRADING NUCLEASE (SDN) proteins was identified in Arabidopsis that degrades mature miRNAs (Ramachandran and Chen, 2008). Work on plant HEN1 has been primarily performed in Arabidopsis, the characterization of wavy leaf1 (wafl) mutants in rice (Oryza sativa), the ortholog of Arabidopsis...
Figure 1. miRNAs Are Differentially Truncated and Tailed in hen1 Mutants.
HEN1 (Abe et al., 2010), and a newly identified hen1 mutant in maize (Zea mays) represent an opportunity to expand our understanding of HEN1 function into monocots. In this study, we used deep sequencing to characterize the extent of 3’ modification of sRNAs in both the wild type and hen1 mutants of Arabidopsis, rice, and maize and found that, in contrast with the previous understanding that all miRNAs are simply shortened or tailed in hen1, the pattern of 3’ modifications varied substantially among miRNA families, and both truncation and tailing are AGO1 dependent.

RESULTS

Sequencing of sRNAs from Plant hen1 Mutants

To date, miRNAs in hen1 mutants were mainly investigated via RNA gel blot hybridization, which is low throughput and cannot reveal the changes in miRNA sequence composition. Cloning followed by Sanger sequencing of miR173 and miR167 in At-hen1-1 revealed truncation and one to five nucleotides of uridylation at the 3’ end of miRNAs (Li et al., 2005). With the tremendous advances in high-throughput sequencing technologies, it is possible systematically to study sRNA population in hen1 mutants with unprecedented depth. Several hen1 mutant alleles have been identified in both Arabidopsis and rice (Chen et al., 2002; Abe et al., 2010; Yu et al., 2010). In this analysis, we analyzed three hen1 mutant alleles in Arabidopsis, three hen1 alleles in rice, and a newly identified hen1 mutant in maize. This includes a new mutant allele of the rice HEN1 gene (Os-hen1-3) from the Korean (POSTEC) rice T-DNA mutant population (Jeong et al., 2002). This mutant, with a T-DNA insertion that disrupts the Os-HEN1 open reading frame 97 bp upstream of the stop codon, has a similar phenotype to the wavy leaf mutants (see Supplemental Figure 1 online). We also identified the hen1 mutant allele in maize (Zm-hen1-1) from the UniformMu population with an insertion in the first exon. In the native W22 background, these plants displayed a shoot-meristemless phenotype like many rice waf1 plants, and we therefore analyzed seedlings from a backcross to B73, which displayed a weaker phenotype (see Supplemental Figure 2 online). We sequenced by Illumina technology sRNAs from all of these plant materials (see Supplemental Table 1 online).

In addition to reduced sRNA levels in hen1 mutants (Li et al., 2005; Yu et al., 2005), we observed that the size distribution of sRNAs in hen1 differed from the wild type in all three species, with reduced 21- and 24-nucleotide peaks, accompanied by an increase in the relative proportion of other sRNA sizes (see Supplemental Figure 3 online). This size shift could be due to sRNA truncation, polyuridylation, or a combination of both. To analyze the sRNA derivatives, we devised aninformatics pipeline as follows (Zhai and Meyers, 2013): First, each non-genome-matched sRNA read was divided into two parts, viz. the longest 5’ genome-matched component head portion and a 3’ tail portion (for the reads that perfectly mapped to the genome we assumed there was no tail); next, focusing first on miRNAs, we aligned the head portion of the reads to each miRBase-annotated miRNA (allowing only perfect matches) to determine the extent of tailing (addition of nontemplated 3’ nucleotides) and truncation (shortening of the miRNA from the 3’ end, usually prior to tailing). This calculated the degree and characteristics of both truncation and tailing for annotated Arabidopsis, rice, and maize miRNAs.

miRNAs Are Differentially 3’ Modified

Consistent with the previous reports of miRNAs in hen1 mutants (Li et al., 2005; Yu et al., 2005; Yu et al., 2010), we were able to...
identify a high level of tailing for many miRNAs, predominantly via uridylation. However, contrary to the previous impression that miRNAs in hen1 mutants are simply tailed or slightly shortened, we found that the degree of 3′ modifications varied substantially among different miRNA families (Figure 1A). Additionally, we observed heavy truncation prior to uridylation for many miRNAs (see Supplemental Data Set 1 online). Because in many cases this restored a length similar to the wild-type miRNA (see below), the combination of heavy truncation and uridylation cannot be revealed by hybridization-based methods like RNA gel blots. The

Figure 2. Arabidopsis miR158 Is Partially Unmethylated in the Wild Type and Subject to Truncation and Uridylation.

(A) Levels of miR158 truncation and tailing are similar in hen1-8 and the wild type (WT) and are reduced by the heso1-1 mutation in the background of either a functional HEN1 allele or in a hen1-1 background. See Figure 1 for the interpretation of matrix. nt, nucleotides.

(B) The length of miR158 tail is shifted to smaller sizes in the heso1-1 mutant compared with Columbia (Col), but the composition is unchanged. The tails of the miR158-derived reads were grouped by their length (position on x axis); the count for the composition of each type of ribonucleotide was as indicated (y axis).

(C) miR158 is partially unmethylated in both Columbia and Ler wild types and thus subject to truncation and tailing. “β” indicates β-elimination treatment, after which unmethylated sRNAs run approximately two nucleotides faster than methylated sRNAs. Red arrows indicate the unmethylated miR158 after β-elimination in both Columbia and Ler.

(D) The expression level of miR158 target, AT3G03580, is decreased in a heso1-1 single mutant relative to the wild-type Columbia-0 level, as measured by quantitative RT-PCR, and error bars represent the estimated SD from three replicated experiments.

(E) The proportion of the most abundant miRNAs comprised by miR158 derivatives in replicated wild-type and hen1 Arabidopsis sRNA libraries: miR158 derivatives are relatively enriched in hen1 mutants compared with the wild type. The y axis shows the mean values for the proportion of miR158 derivatives from three replicated libraries for each genotype, and error bars represent the estimated SD.
unique patterns described below for different miRNA families were highly reproducible in both hen1-1 and hen1-8, mutant alleles isolated in two different Arabidopsis accessions (Figures 1B and 1C; see Supplemental Data Set 1 online) and are even conserved across plant species (Figure 1B; see Supplemental Figure 4 and Supplemental Data Set 2 online), suggesting that structural, sequence-dependent, or other as yet unknown effects specify the 3’ modifications resulting from reduced HEN1 levels.

miR165/6 (21 nucleotides) family members play important roles in plant development by targeting type III HD-Zip genes REVOLUTA, PHABULOSA, and PHAVOLUTA to regulate leaf polarity and stem cell identity (Mallory et al., 2004; Williams et al., 2005). In hen1 mutants, we observed that miR166 is first truncated to ~17 nucleotides without any tailing and then is uridylated to ~21 nucleotides, its original length (Figure 1B). Our pipeline defined the truncation to ~17 nucleotides, but it is possible that the truncation was to ~15 nucleotides (16th and 17th nucleotides of miR166a are both uridine). Since plant miRNAs are generally 21 nucleotides, we wondered if the tailing back to 21 nucleotides is a universal feature. To test this, we analyzed miR163, which is unique because of its unusual 24-nucleotide length, due to two asymmetric bulges on the miRNA strand of the duplex (Kurihara and Watanabe, 2004). In Arabidopsis hen1 mutants, miR163 has the longest truncation among all miRNAs; it is truncated by eight nucleotides to a predominant length of 16 nucleotides before uridylation, and then it is tailed back to its original length at ~24 nucleotides (Figures 1C and 1D). This ability to regain the original miRNA size after truncation and tailing implies a memory of the original length. These two examples also suggest that the truncation activity, possibly performed by the SDN family of exonucleases (Ramachandran and Chen, 2008), in cases like miR166 and miR163 could be upstream of the uridylation activity, as tailing may not occur until the miRNA is shortened to a certain length. Other patterns were also apparent, for example, miR156, which targets SPL genes to regulate developmental time (Poethig, 2009), and miR172, which represses the translation of AP2-domain transcription factors.
(Chen, 2009), were tailed but with little truncation; the Dicer-like1-targeting miR162 (Xie et al., 2003) was only slightly tailed or truncated (somewhat protected) (see Supplemental Figure 4 online). Thus, these patterns are highly conserved among different hen1 alleles from different plant species.

3’ Modified miRNAs in Wild-Type Plants

Prior analyses suggested that plant miRNAs are fully methylated in vivo by HEN1, which protects them from 3’ uridylation (Li et al., 2005; Yu et al., 2005). While our sequencing results showed that miRNAs are predominately protected in the wild type from truncation and tailing, we found some exceptions. Even in an Arabidopsis wild-type background with functional HEN1, miR158 was both truncated and tailed at substantial levels, levels somewhat similar to this miRNA in the hen1 mutants (Figure 2A). HEN1 SUPPRESSOR1 (HESO1) is a nucleotidyldtransferase that can uridylate unmethylated miRNAs and promote their degradation (Ren et al., 2012; Zhao et al., 2012b). Arabidopsis HESO1 was identified because its loss of function can suppress the phenotype of hen1-8 by reducing levels of uridylation and increasing miRNA accumulation in general. HESO1 can only add nucleotides (predominantly uridine) to the 3’ terminus of unmethylated RNA, as 2’-O-methylation blocks its activity (Zhao et al., 2012b). In the heso1-1 mutant, we found that the level of tailing of miR158 is dramatically reduced (Figure 2A), and the length of the miR158 tail was shifted to smaller sizes because of reduced uridylation (Figure 2B). However, miR158 is not unique, as miR319a is also tailed in wild-type Arabidopsis and its modification is suppressed in heso1-1 (Zhao et al., 2012b).

The fact that miR158 is uridylated in the wild type and its tailing pattern is impacted by heso1-1 suggests that miR158 might be partially unmethylated in the wild type. To assess miR158 methylation in the wild type, we performed a β-elimination experiment, which increases the mobility of only unmethylated sRNAs by approximately two nucleotides in RNA gel blots (Aufelder et al., 1998; Hutvágner et al., 2001). Consistent with our hypothesis, a portion of miR158 in the wild type ran two nucleotides faster, the same as the unmethylated miR158 in hen1 mutants (Figure 2C). We concluded that in the wild type Arabidopsis miR158 is characterized by incomplete 2’-O-methylation and consequently is truncated and uridylated. Since uridylation leads to degradation of miRNAs, we asked whether miR158 activity is modulated by its incomplete methylation and uridylation. Indeed, the expression level of the miR158 target, AT3G03580, is decreased in the heso1-1 mutant (Figure 2D), suggesting that miR158 function is partially suppressed in the wild type via destabilization. To assess the possibility of destabilization, we examined the proportion of the most abundant miRNAs comprised by miR158 derivatives in replicated wild-type and hen1 Arabidopsis sRNA libraries (see Supplemental Table 1 online). This showed that miR158 levels comprise ~14 to 17% of the total in the wild type (Columbia-0 and Landsberg erecta [Ler]) but 41% in both hen1-1 and hen1-8 (Figure 2E). This enrichment suggests that in a hen1 background in which all other miRNAs are destabilized, miR158 is relatively less impacted by the loss of HEN1.

The level of 3’ modifications observed in miR158 and miR319 led us to look more carefully at published sequencing data in wild-type plants for many other species, including monocots (maize, Brachypodium, and rice), other eudicots (soybean [Glycine max], Medicago, and tomato [Solanum lycopersicum]), and even the basal land plant Selaginella. We picked the three most representative sequence variants of the truncated and tailed forms of miR166 in the Arabidopsis hen1 mutant to search in wild-type sRNA libraries, and we found them present in all the species examined (see Supplemental Table 2 online). Next, we...
examined 3’ truncation and tailing patterns of miR166 variants from these wild-type species, removing from consideration the canonical miR166 to emphasize the truncated and tailed variants (Figure 3A). Across the broad range of species examined, the patterns of truncation and tailing are strikingly similar, with slightly reduced truncation and tailing observed only in tomato (Figure 3B). This suggests that the truncation and tailing of miR166 is ancient and highly conserved across plant species and only becomes more visible in a hen1 mutant background. Thus, wild-type plants show truncation and tailing of miRNAs enhanced in hen1 (e.g., miR166), and some miRNAs are naturally 3’ modified at moderate levels (e.g., miR158 and miR319).

miR171a Is Uridylated from 21 to 22 Nucleotides in hen1, Triggering Phased, Secondary siRNA Production

Many miRNAs are truncated and uridylated to exhibit heterogeneous sizes in hen1 mutants, which includes 22 nucleotides, a length perhaps sufficient to trigger at their targets the production of secondary siRNAs (Chen et al., 2010; Cuperus et al., 2010). These secondary siRNAs are distributed in fixed intervals (a phase of usually 21 nucleotides) due to consistent and precise miRNA cleavage. Given that most Arabidopsis miRNAs are 21 nucleotides in the wild-type background, it is possible that some or many miRNA targets produce tasiRNAs in the hen1 mutant because the miRNA variants that are tailed to 22 nucleotides. We performed a genome-wide analysis of sRNA phasing (Zhai et al., 2011) with the hen1 mutant sRNA libraries. Only the two loci AT2G45160 and AT3G60630, both of which regulate the development of shoot meristem (Schulze et al., 2010), produced phased siRNAs in hen1 mutants but not the wild type (Figure 4A; see Supplemental Figure 5 online). These genes are both the predicted targets of miR170/171a (see Supplemental Figure 5 online). Both miR170 and miR171a were primarily tailed to and stabilized at 22 nucleotides in hen1 (Figure 4B; see Supplemental Data Set 1 online). However, 22-nucleotide variants of other miRNAs that are 21 or 20 nucleotides in the wild type did not gain the ability to trigger phased siRNAs at their targets (data not shown). Thus, in hen1, an sRNA length of 22 nucleotides is necessary but not sufficient to trigger phased siRNA production. Manavella et al. (2012) previously described that both (1) RNA-induced Silencing Complex (RISC) re-programming by asymmetric structure of miRNA/miRNA* duplex can enable them to trigger secondary siRNAs, and (2) miR170/171 lack such asymmetry on their duplex but were found in ARGONAUTE7 immunoprecipitation (AGO7-IP) sRNAs, which might permit it to initiate secondary siRNA production, like the AGO7-bound miR390. Curiously, in the double mutant heso1-1 hen1-8, the 22-nucleotide variant of miR171a was unaffected (Figure 4B); thus, the single 3’ nucleotide addition must be added by a different enzyme. We checked the miR171c/d family...

Figure 5. AGO1 Binding and Involvement in 3’ Truncation and Tailing of Arabidopsis miRNAs.

(A) AGO1 binding of miRNA variants shown by RNA gel blots using input and AGO1 IP-derived sRNAs from Columbia (Col), hen1-8, and hen1-8 heso1-1. Top panel: The truncated and uridylated miR158 variants. Bottom panel: The canonical 21-nucleotide (nt) miR171a and its primarily 22-nucleotide variant.

(B) Measurement of truncated and tailed miR166a variants in Arabidopsis in either hen1-2 (left) or a double mutant of ago1-11 and hen1-2 (right). In the double mutant, truncation and tailing is suppressed.

(C) Proportion of 3’ modified variants of two Arabidopsis miRNAs in single and double mutants of ago1-11 and hen1-2 (as labeled).
members, which are 3’ shifted by three nucleotides relative to miR170/171a and show different patterns of 3’ modifications (e.g., not consistently 22 nucleotides; see Supplemental Data Set 1 online).

**Mutations in AGO1 Suppress the Truncation and Tailing of miRNAs**

Since most plant miRNAs are bound by AGO1, we next wondered about the role of AGO1 in 3’ modifications of miRNAs, asking the following: (1) Are 3’ modified miRNAs competent to be bound by AGO1? (2) If they are bound, does AGO1 influence 3’ modifications? In *Drosophila melanogaster*, extensive matching between Argonaute1-bound miRNA and its target destabilize miRNA through 3’ truncation and tailing (Ameres et al., 2010). By RNA gel blots, immunoprecipitation (IP) of AGO1 showed that it bound the variants of all sizes for miR158 and miR171a in an *Arabidopsis hen1* background, with the binding of these variants unaffected in the *heso1-1 hen1-8* double mutant (Figure 5A). Similar observations were made for miR165/6 and miR167 (Zhao et al., 2012a). Sequenced sRNAs from AGO1 IP further confirm the binding of 3’ modified sRNAs in *hen1* and *hen1 heso1* for many other miRNAs (see Supplemental Figure 6A online). To answer whether AGO1-bound miRNAs were actively modified while in association with AGO1 or these miRNAs were already modified prior to AGO1 loading, we sequenced sRNAs from *ago1-1* (a weak allele), *hen1-2*, and an *ago1-11 hen1-2* double mutant (see Supplemental Table 1 online). Truncation and tailing of many miRNAs in *hen1* was suppressed by *ago1* mutants, for different alleles of both genes (Figures 5B and 5C; see Supplemental Figure 6C online), suggesting that 3’ modifications of unmethylated miRNAs occur after AGO1 loading. Consistent with this, AGO1 and HESO1 colocalize (G. Ren, M. Xie, C. Vinovskis, and B. Yu, unpublished data). We also examined immunoprecipitated miRNAs from an AGO1 slicer-defective mutant (Carbonell et al., 2012) and observed little impact on miR158 tailing, indicating that AGO1 slicer activity is not required to modulate 3’ modifications (see Supplemental Figure 6B online). Such 3’ modifications occurring without target cleavage are reminiscent of *D. melanogaster* miRNA truncation and tailing that occurs when extensively matched by a synthetic target mRNA (Ameres et al., 2010). In contrast with the *heso1 hen1* double mutant, which is impacted only in 3’ tailing (Zhao et al., 2012b), in *ago1-11 hen1-2*, both truncation and tailing are reduced (Figure 5B), suggesting that AGO1 not only interacts with HESO1 but may have a role in modulating 3’ exonucleolytic activity.

**Sequence Determinants of miRNA 3’ Modifications**

A key question for our study is the nature of the determinants impacting the 3’ modifications. The sRNA sequence presumably has a role, which could reflect individual nucleotides or structural characteristics of the miRNA duplex. We sought to investigate such impacts using miRNA family members with similar but distinguishable sequence. The 5’-most nucleotide of a miRNA is a primary determinant for the AGO onto which the miRNA will be loaded (Mi et al., 2008). The related miR158a and miR158b differ only by the 5’ nucleotide, yet have identical 3’ modifications (Figure 6A). Meanwhile, miR170/171a are closely related to miR171b/c, with a conserved core sequence, but a processing shift of three nucleotides alters the 3’ modifications (Figure 6B). Together, these data suggest that the 5’ and middle part of the miRNA are not essential for shaping the 3’ truncation and tailing pattern, leaving the 3’-end sequence as a more promising candidate. Indeed, miR166 and miR169, which share similar sequences at the 3’ end but different in their 5’ and

![Figure 6. Impacts of Sequence on miRNA Truncation and Tailing (in Arabidopsis hen1-1).](image-url)

Three sets of miRNAs were selected to test whether the 5’ end, middle region, or 3’ end of the miRNA influences 3’ truncation and tailing. In each case, an alignment above highlights the difference of the selected miRNAs, and the panels below show the pattern of 3’ truncation and tailing in the *Arabidopsis hen1-1* mutant. The sets of miRNAs and the three regions of the miRNA are as follows: miR158 family (5’ end) (A), miR171 family (middle part and 3’ end) (B), and miR166 and miR169 (3’ end) (C). WT, the wild type.
middle regions, are almost identical in their truncation and tailing pattern (Figure 6C), indicating that the 3' end sequence may substantially influence the patterns of miRNA 3' modification.

**siRNAs Are Truncated and Uridylated in hen1 Mutants**

Previous studies on individual sRNAs have shown that 24-nucleotide heterochromatic siRNAs (hc-siRNAs) are uridylated and decreased in abundance in hen1 mutants (Li et al., 2005), and their uridylation is also impacted by a heso1 mutation, confirmed in the heso1-1 hen1-8 background (Zhao et al., 2012b). To systematically study the impact of the hen1 mutation on hc-siRNAs, we mapped the genome-matched components of sRNAs (the head portion) to annotated transposable elements to enrich for 24-nucleotide hc-siRNAs and filter out miRNAs and other types of sRNAs (such as trans-acting siRNAs). As expected, in the wild type, almost all the sRNAs that mapped to the transposable elements were predominately 24 nucleotides in length; in both hen1-1 and hen1-8 mutants, the proportion of 23- and 25-nucleotide variants increased relative to 24-nucleotide siRNAs, probably reflecting both truncation and tailing of these hc-siRNAs (see Supplemental Figure 7 online). Although the abundance of hc-siRNAs is drastically altered in the mutant background (Yu et al., 2010), the overall size distribution of hc-siRNAs in hen1 was only moderately more dispersed from the 24-nucleotide peak (see Supplemental Figure 7 online), suggesting a more stringent size constraint on the 24-nucleotide hc-siRNAs than the AGO1-bound miRNAs. This constraint could result from properties of AGO4, the AGO protein that binds most hc-siRNAs and acts in transcriptional silencing (Zilberman et al., 2004; Qi et al., 2006). We also examined tasiRNAs from Arabidopsis and observed that two abundant TAS3 sRNAs are highly truncated and tailed in hen1 mutants, but with different patterns (see Supplemental Data Set 1 online). Thus, hen1 mutations broadly affect sRNAs via increased 3' modifications.

**DISCUSSION**

The Process of Plant miRNA 3' Truncation and Tailing

When and where do these 3' sRNA modifications occur? Our data showing that ago1 mutations suppress both truncation and tailing indicate that the 3' modifications take place in association with AGO1, either during the process of AGO1 loading or in the assembled RISC. Other data support this, including a recent demonstration of direct AGO1–HESO1 interaction (G. Ren, M. Xie, C. Vinovskis, and B. Yu, unpublished data) and a demonstration that near-perfect miRNA-mRNA target pairing can trigger 3’ modifications of D. melanogaster miRNAs (Ameres et al., 2010).

To achieve differential truncation and tailing, the machinery responsible for sRNA modifications must be able to distinguish different miRNAs after their loading into AGO1. The distinct truncation and tailing patterns of miR166 and miR163, and especially the fact that they are uridylated back to their original length (21 and 24 nucleotides, respectively) (Figure 1D), show that the truncation and tailing pattern is specified by the miRNA sequence. Our analysis of similar miRNAs and their 3’ modifications suggest that it is the 3’ sequence that influences the patterns of miRNA 3’ modification. Structural analyses of a binary complex of a bacterial Argonaute bound to a 5’-phosphorylated...
DNA guide strand and target RNA reveal that base-pairing between guide strand and target occurs from positions 2 to 16 of the guide DNA strand (Wang et al., 2009), which may explain why many miRNAs in hen1 are truncated to −16 nucleotides before being tailend. Meanwhile, although animal miRNAs generally match poorly to their targets, it was proposed that differential miRNA-target pairing could affect a variety of allosteric changes on the Argonaute protein or RISC complex (Elkayam et al., 2012). Perhaps the RISC complex is programmed by the 3′ end of the loaded miRNA to direct specific patterns of 3′ modifications; consistent with this, a recent report illustrated that an asymmetric bulge on the miRNA/miRNA∗ duplex can program RISC during the loading step to enable the function of triggering effectors can recognize specific DNA sequences with single nucleotide resolution through their repeat variable dinucleotides (Deng et al., 2012; Mak et al., 2012). Two different transcription activator-like effectors, Talc of Xanthomonas oryzae pv oryzicola and Tal9a of X. oryzae pv oryzae, are encoded by two distinct Xanthomonas strains yet share a common target: the promoter of the rice HEN1 gene. These bacterial proteins target different regions of the same promoter, and both effectors dramatically enhance HEN1 expression during infection (Moscou and Bogdanove, 2009). The reason for the induced overexpression of HEN1 is as yet unknown, but perhaps this produces higher levels of HEN1 protein that may alter the relative abundance of miRNAs or other sRNAs significant to host responses. Perhaps Xanthomonas is thus exploiting HEN1 and sRNA methylation to enhance the susceptibility of rice to the pathogen, as numerous sRNA-related cellular components are known to have a role in plant defenses (Katiyar-Agarwal and Jin, 2010). We believe that sRNA truncation and tailing, highly pronounced in the hen1 mutants, is likely to have an important endogenous function in wild-type plants.

METHODS

Plant Materials

Immature inflorescence tissues, including inflorescence meristem, and early stage floral buds (up to stage 11/12) were collected from Arabidopsis thaliana grown in a growth chamber with 16 h of light for 5 weeks. For the seedling materials from the wild type, and the ago1-11 and hen1-2 single mutants, as well as the ago1-11 hen1-2 double mutants, seeds were sterilized and plated on half-strength Murashige and Skoog medium containing 1% (w/v) Suc and 1% (w/v) agar. Plates were first kept in 4°C for 3 d and then moved to growth chamber and grown under continuous light. Whole-plant tissue samples were harvested 12 d after germination.

Rice plants containing the Os-hen1 alleles waf1-1 and waf1-2 were grown in a growth chamber with 12 h of light for 5 weeks and genotyped by PCR to identify homozygous plants. The Os-hen1-3 allele was identified from the Kyung Hee University T-DNA collection (Jeong et al., 2002) as line PFG_2C-00348; seeds were obtained and homozygous lines identified by PCR-based genotyping. For library construction, homozygous mutant seedlings at the 5-week stage were collected and the entire plants used for RNA extraction.

The Zm-hen1-1 allele was isolated in W22 background with a Mu insertion (mu1007900) within the first exon of HEN1. This insertion was verified by PCR analysis and introgressed into B73. Materials were obtained from the homozygous F2 progeny derived from the W22 × B73 cross.

RNA Extraction and Sequencing of sRNAs

Total RNA from the materials described above was isolated using Trizol reagents from Invitrogen. sRNA libraries were constructed using the Illumina TruSeq Small RNA Sample Preparation Kit (RS-200-0012) and sequenced on either an Illumina GAIIx or HiSeq2000 instrument at the Delaware Biotechnology Institute.

RNA Gel Blotting, IP of AGO1, and Real-Time RT–PCR

RNA isolation and RNA gel blotting to detect sRNAs were performed as previously described (Park et al., 2002). Antisense DNA oligonucleotides that were 5′ end labeled (32P) were used to detect miRNAs. Sodium periodate treatment and β elimination were done as previously described.
The probe for detecting miR171a was 5’-GATATTGG-CGCCGGCTCAATCA-3’ (21 nucleotides); the probe for detecting miR158 was 5’-GCTTTGCTTACATTGGGA-3’ (19 nucleotides). IP was performed as previously described (Li et al., 2011). The AGO1 mouse polyclonal antibody was generated in the Cao lab (Institute of Genetics and Developmental Biology, Chinese Academy of Sciences). Reverse transcription and real-time PCR were performed as previously described (Kim et al., 2011) with UBIQUITIN5 used as an internal control. Primers used to examine the expression of the miR158 target, AT3G03580, were described previously (Li et al., 2010).

**Bioinformatics Analysis of sRNAs**

For sRNA reads that were not genome matched, one nucleotide was chopped off from the 3’ end in successive rounds until the remaining 5’ sequence was perfectly mapped to the Arabidopsis genome as previously described (Zhai and Meyers, 2013). Thus, any non-genome-matched sRNA read could be split into two parts: a 5’ head and a 3’ tail. The head portion of each read was aligned to all annotated Arabidopsis miRNAs in miRBase version 17 (Griffiths-Jones et al., 2008) for their origin, thus determining the extent of tailing (addition of nonconventional 3’ nucleotides) and truncation (shortening of the miRNA from the 3’ end). The head portion of reads was also mapped to the annotated transposable elements in TAIR (version 10; used only for its repeat data) to filter the transposable element–associated sRNAs used in Supplemental Figure 7 online, which were mostly ~24 nucleotides in the wild-type libraries. Genome-wide phasing analysis of sRNAs was performed as described previously (Zhai et al., 2011).

**Accession Numbers**

The GenBank Gene Expression Omnibus accession number for the sRNA data described in Supplemental Table 1 online is GSE35562 (also found at http://smallrna.udel.edu/). Data for Columbia, hsa0-1, hen1-8, and hsa01-1 hen1-8 libraries are from Zhao et al. (2012b). IP data from the wild type and catalytic mutants of AGO1 are from Carbonell et al. (2012).

**Supplemental Data**

The following materials are available in the online version of this article.

Supplemental Figure 1. Characterization of New hen1 Allele in Rice.

Supplemental Figure 2. Characterization of New hen1 Allele in Maize.

Supplemental Figure 3. Size Distributions for Small RNA Sequencing Data.

Supplemental Figure 4. miRNAs are Differentially Truncated and Unmodified in hen1 Mutants.

Supplemental Figure 5. miR171a Targets Produce 21 nt siRNAs in hen1 Mutants.

Supplemental Figure 6. In Arabidopsis, AGO1 Binds to 3’ Truncated and Tailed miRNAs.

Supplemental Figure 7. Size Distributions of Transposon-associated Small RNAs.

Supplemental Table 1. Summary of Small RNA Libraries from Plant hen1 Mutants.

Supplemental Table 2. Truncated and Tailed Variants of miR166 in Diverse Plant Species.

Supplemental Data Set 1. Truncation and Tailing Patterns of Conserved miRNAs in Arabidopsis hen1 Mutants.

Supplemental Data Set 2. Truncation and Tailing Patterns of Conserved miRNAs in Rice hen1 Mutants.

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**AUTHOR CONTRIBUTIONS**

J.Z. and B.C.M. designed research, performed research, contributed new computational methods, analyzed data. M.P. contributed new computational methods. S.A.S., S.H., K.P., S.A., L.J., M.X., X.C., B.Y., M.T., B.Y., and X.M. designed and performed research. All authors contributed to writing the article.

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