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Jong W. Rhim

Mokpo National University, Korea

Aristippos Gennadios

Banner Pharmacaps Inc., aris.gennadios@catalent.com

Curtis Weller

University of Nebraska-Lincoln, cweller1@unl.edu

Milford Hanna

University of Nebraska-Lincoln, mhanna1@unl.edu

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Sodium dodecyl sulfate treatment improves properties of cast films from soy protein isolate

Jong W. Rhim,¹ Aristippos Gennadios,²
Curtis L. Weller,³ and Milford A. Hanna³

1. Department of Food Engineering, Mokpo National University, 61 Dorim-ri, Chungkye-myon, Muan-gun, Chonnam 534-729, Republic of Korea
2. Product Development, Materials Science Group, Research and Development, Banner Pharmacaps Inc., 4125 Premier Drive, High Point, NC 27265-8144, USA
3. Industrial Agricultural Products Center and Department of Biological Systems Engineering, University of Nebraska-Lincoln, Lincoln, NE 68583-0730, USA

Corresponding author – A. Gennadios, tel 336 812-8700, fax 336 812-9091, email agennadios@banpharm.com

Abstract

The manufacture of edible/biodegradable films or coatings can potentially add value to soy protein. This study was conducted to determine the effect of sodium dodecyl sulfate (SDS) on selected physical properties of glycerin-plasticized soy protein isolate (SPI) films. Films were cast from heated (70 °C for 20 min), alkaline (pH 10) aqueous solutions of SPI (5 g/100 ml water), glycerin (50% w/w of SPI), and SDS (0, 5, 10, 20, 30, or 40% w/w of SPI). Tensile strength (TS), elongation at break (E), moisture content (MC), total soluble matter (TSM), water vapor permeability (WVP), and color values (L , a , and b) were determined after conditioning film specimens at 25 °C and 50% relative humidity (RH) for 2 days. SDS reduced ($P < 0.05$) film TS by as much as 43% for films with 40% SDS (6.2 vs. 10.9 MPa for control SPI films). In contrast, film E increased ($P < 0.05$) notably with addition of SDS even at 5%. Films with SDS had smaller ($P < 0.05$) MC and larger ($P < 0.05$) TSM values than control SPI films. Films containing 10% or more SDS had lower WVP values than control SPI films by as much as 50%. Increased yellowness, evidenced by greater ($P < 0.05$) $+ b$ color values, was noted for films with high amounts (20, 30, or 40%) of SDS. Changes in tensile, solubility, and water vapor barrier properties of SPI films due to the addition of SDS were largely attributed to disruption of hydrophobic associations among neighboring protein molecules as the non-polar portions of the SDS molecules attached onto hydrophobic amino acid residues within the film structure. It was demonstrated that adding anionic surfactant SDS to film-forming solutions prior to casting could greatly modify the properties of SPI films. In particular, SDS improved the water vapor barrier ability and the extendibility of SPI films, both desirable attributes when assessing the potential of such films for packaging applications.

Keywords: protein films, soy protein, sodium dodecyl sulfate, water vapor permeability

1. Introduction

The manufacture of edible/biodegradable films or coatings can potentially add value to soy protein. Soy protein-based films formed on the surface of heated soy-milk have traditionally been prepared in the Far East (Gennadios et al., 1994a). Also, free-standing soy protein films can be formed by casting (Brandenburg et al., 1993; Gennadios et al., 1993), extrusion (Naga et al., 1996), spinning in a coagulating buffer (Rampon et al., 1999), or thermal compaction (Cunningham et al., 1999; Ogale et al., 2000). Plasticizers, typically glycerin and/or sorbitol are added to soy protein films to impart flexibility (Gennadios et al., 1994a).

Various physical (Gennadios et al., 1996, 1998a; Rangavajhyala et al., 1997; Rhim et al., 1999a, 2000), chemical (Ghorpade et al., 1995; Rhim et al., 1998, 1999b, 2000; Were et al., 1999), or enzymatic (Motoki et al., 1987; Stuchell and Krochta, 1994; Yildirim and Hettiarachchy, 1997) treatments have been used to modify the properties of soy protein films. Such treatments mainly promote cross-linking within the protein film network. Ionic surfactants, such as sodium dodecyl sulfate (SDS), are powerful denaturing and dissociating agents for proteins (Graveland et al., 1979; Cheftel et al., 1985). Strength reduction, and even re-solubilization, of protein gels in SDS buffers has been reported (Kitabatake and Doi, 1993; McClements et al., 1993; Kiosseoglou et al., 1999). Therefore, incorporation of SDS into protein-based film-forming solutions would be expected to affect the structure and properties of cast protein films. Fairley et al. (1996) reported that SDS showed potential for use as an adjunct to polyols (i.e. glycerol and sorbitol) for the plasticization of whey protein isolate films. Our objective was to investigate the effects of SDS on selected physical properties (i.e. tensile strength, elongation at break, total soluble matter, water vapor permeability, and color) of cast soy protein isolate (SPI) films.

2. Materials and methods

2.1. Film preparation

Film-forming solutions were prepared by slowly dissolving 5 g of SPI (minimum 90% protein content on dry basis, SUPRO 620, Protein Technologies International, St. Louis, MO) in a constantly stirred mixture of distilled water (100 ml) and glycerin (2.5 g). Glycerin was added as a plasticizer to overcome film brittleness and to obtain freestanding films. The solution pH was adjusted to 10 ± 0.1 with 1 M sodium hydroxide. Alkaline conditions favor SPI film formation, presumably by aiding protein dispersion in film-forming solutions (Okamoto, 1978; Gennadios et al., 1993). After heating the solutions for 20 min at 70 °C in a constant temperature water bath, they were strained through eight-layered cheese cloth (grade 40, Fisher Scientific, Pittsburgh, PA) to remove any small lumps (minuscule amounts), and cast onto leveled, Teflon®-coated glass plates (21×35 cm). Film thickness was controlled by casting the same amount (80 ml) of film-forming solution per plate. The castings were dried at ambient conditions (25 °C) for about 20 h. Dried films were peeled from the plates and specimens, for property testing, were cut (2.54×10 cm for tensile testing; 2×2 cm for moisture content and total soluble matter; and 7×7 cm for water vapor permeability and color measurements). In addition to 'control' films prepared in this manner, films also were prepared by adding SDS (Fisher Scientific) at 5, 10, 20, 30, or 40% w/w of SPI into the film-forming solutions prior to pH adjustment.

2.2. Film thickness and conditioning

Film thickness was measured to the nearest 2.54 μm with a hand-held micrometer (B.C. Ames Co, Waltham, MA). Five thickness measurements were taken on each tensile testing specimen along the length of the rectangular strip and the mean value was used in tensile strength calculations. Similarly, five measurements were taken on

each water vapor permeability specimen, one at the center and four around the perimeter, and the mean values were used in calculating water vapor permeability. All film specimens were conditioned for 2 days in an environmental chamber (Model RC-5492, Parameter Generation & Control, Inc, Black Mountain, NC) at 50% relative humidity (RH) and 25 °C before testing.

2.3. Tensile testing

Film tensile strength (TS) and percentage elongation at break (E) were determined with an Instron Universal Testing Machine (model 5566, Instron Engineering Corp, Canton, MA). Initial grip separation was set at 50 mm and cross-head speed was set at 500 mm/min. TS was calculated by dividing the maximum load by the initial cross-sectional area of the specimen (ASTM, 1995a). E was calculated by dividing the extension at rupture of the specimen by the initial gage length of the specimen (50 mm) and multiplying by 100 (ASTM, 1995a). TS and E measurements for each type of film were replicated three times with individually prepared films as the replicated experimental units and each replicate being the mean of two tested sampling units (specimens) taken from the same film.

2.4. Moisture content

Film samples were weighed (± 0.0001 g) into aluminum dishes and dried in an air-circulating oven at 105 °C for 24 h. Moisture content (MC) was determined as the percentage of initial film weight lost during drying and reported on a wet basis. Triplicate measurements of MC were obtained for each type of film with individually prepared films as replicated experimental units and three specimens tested from each film.

2.5. Total soluble matter

Total soluble matter (TSM) was determined as the percentage of film dry matter solubilized after 24 h immersion in distilled water. Proteins are susceptible to heat-induced cross-linking (Cheftel et al., 1985). Heat

treatments have increased the water resistance of soy protein films (Gennadios et al., 1996; Rangavajhyala et al., 1997; Rhim et al., 2000). Therefore, to avoid heat curing, film specimens were not dried in an oven to determine their initial dry matter prior to solubility testing (Rhim et al., 1998). Instead, the initial dry matter of conditioned films was determined on different specimens (two from each film) by drying in an air-circulating oven (105 °C for 24 h).

2.6. Water vapor permeability

Water vapor permeability (WVP) ($\text{g mm}/\text{m}^2 \text{ h kPa}$) was calculated from:

$$\text{WVP} = \frac{\text{WVTR} \cdot L}{\Delta p}$$

where WVTR was the measured water vapor transmission rate ($\text{g}/\text{m}^2 \text{ h}$) through the film specimen; L was the mean film thickness (mm); and Δp was the partial water vapor pressure difference (kPa) across the two sides of the film specimen.

WVTR was determined gravimetrically using a modified ASTM Method E 96-95 (ASTM, 1995b). Film specimens were mounted horizontally on poly(methylmethacrylate) cups filled with distilled water up to 1 cm from the bottom side of the film. The design of the cups was described by Gennadios et al. (1994b). The cups were placed in an environmental chamber at 25 °C and 50% RH. A fan was operated within the chamber (air velocity of 198 m/min) to remove permeating water vapors from the surface of the cups. The cups were weighed every hour for a period of 6 h. Steady state was reached within 1 h. The slopes of the steady state (linear) portion of weight loss versus time curves were used to calculate WVTR. Due to the low water vapor resistance of the hydrophilic SPI films, actual RH values at the film surfaces facing the water during testing were less than the theoretical value of 100%. Actual RH values at the film surfaces facing the water and film WVP values were calculated after accounting for the resistance of the stagnant air layer between the film surfaces facing the water and the water surface in the cups (Gennadios et al., 1994b;

McHugh et al., 1994). The mean of the initial and final stagnant air gap heights was used in the calculations. For each type of film, WVP measurements were replicated three times with individually prepared films as the replicated experimental units and each replicate being the mean of two tested sampling units (specimens) taken from the same film.

2.7. Color measurements

Color values of the films were measured with a portable colorimeter (CR-300 Minolta Chroma Meter, Minolta Camera Co., Osaka, Japan). Film specimens were placed on a white standard plate (calibration plate CR-A43; $L = 96.86$, $a = -0.02$, and $b = 1.99$) and the L , a , and b color values were measured. L values range from 0 (black) to 100 (white); a values range from -80 (greenness) to 100 (redness); and b values range from -80 (blueness) to 70 (yellowness). Five measurements were taken on each specimen, one at the center and four around the perimeter. Color measurements for each type of film were replicated three times with individually prepared films as the replicated experimental units.

2.8. Statistical analysis

Statistics on a completely randomized experimental design were determined using the General Linear Models procedure in SAS software (SAS, 1993). Significantly ($P < 0.05$) different means were separated with Duncan's multiple range test.

3. Results and discussion

3.1. Tensile properties

The addition of SDS substantially reduced ($P < 0.05$) the TS of SPI films by as much as 43% for films with 40% SDS (Table 1). In general, the formation of SPI film and gel structures involves covalent disulfide bonds as well as hydrophobic interactions and hydrogen bonds (Fukushima and Van Buren, 1970; Farnum et al., 1976; Doi and Kitabatake, 1989). SDS does not cleave disulfide bonds, but it prevents hydrophobic associations among protein molecules (Fukushima and Van Buren, 1970; Kitabatake and Doi, 1993). This is accomplished through attachment of the hydrophobic (non-polar) portions of the SDS molecules onto hydrophobic amino acid residues (Krull and Wall, 1969). Therefore, it is likely that the reduced TS of SDS-containing SPI films was related to formation of "weaker" structures due to lack of hydrophobic interactions among neighboring protein molecular chains. In addition to its hydrophobic portion, the SDS molecule also carries an anionic (polar) portion. Since the pH of the film-forming solutions was markedly greater than the approximate isoelectric point of soy protein (pH 4.5), protein chains were mainly negative charged. The attached SDS molecules further increased the negative charges along protein molecular chains. This highly charged state of the protein molecules could have further contributed to low film TS values as strong repulsive forces could have decreased the occurrence of molecular associations within the protein matrix.

Table 1. Tensile strength (TS), elongation at break (E), moisture content (MC), and total soluble matter (TSM) of cast films from soy protein isolate (SPI) treated with different levels of sodium dodecyl sulfate (SDS)

SDS (% w/w of SPI)	TS (MPa) ^a	E (%) ^a	MC (% wb) ^a	TSM (%) ^a
0	10.9 ± 0.8 <i>a</i>	50 ± 6 <i>c</i>	23.0 ± 1.2 <i>a</i>	25.3 ± 0.4 <i>c</i>
5	8.4 ± 0.6 <i>b</i>	156 ± 11 <i>b</i>	22.3 ± 0.4 <i>b</i>	25.3 ± 4.2 <i>c</i>
10	7.3 ± 0.3 <i>bc</i>	192 ± 15 <i>a</i>	22.0 ± 0.8 <i>b</i>	32.1 ± 2.0 <i>b</i>
20	7.1 ± 0.6 <i>bc</i>	209 ± 25 <i>a</i>	21.2 ± 0.2 <i>c</i>	36.0 ± 3.0 <i>ab</i>
30	6.4 ± 0.7 <i>c</i>	190 ± 15 <i>a</i>	21.4 ± 0.5 <i>c</i>	35.8 ± 3.4 <i>ab</i>
40	6.2 ± 0.2 <i>c</i>	186 ± 12 <i>ab</i>	17.2 ± 0.3 <i>d</i>	37.0 ± 0.7 <i>a</i>

a. Means of three replicates ± standard deviations. Means in the same column followed by the same letter are not significantly ($P > 0.05$) different by Duncan's multiple range test.

In contrast to TS, film *E* values increased ($P < 0.05$) notably with the addition of SDS (Table 1). For example, films with 5% SDS had three times greater *E* than control films. This can be attributed to SDS-induced unfolding of the protein chains. In general, by disrupting hydrophobic interactions in proteins, SDS separates polypeptide chains into 'rod-shaped' units (Graveland et al., 1979). Therefore, the greater *E* could have resulted from a more linear orientation of protein molecules in SPI films containing SDS. Similarly, Fairley et al. (1996) reported that SDS had a moderate plasticization effect on whey protein isolate films when used in combination with glycerol, as in our SPI films, or sorbitol, although SDS could not plasticize films on its own.

3.2. Moisture content and total soluble matter

The MC of the SPI films, after conditioning at 25 °C and 50% RH for 2 days, decreased ($P < 0.05$) with the addition of SDS (Table 1). This was likely partially due to the hydrophobic portions of the SDS molecules. Also, the amount of glycerol per gram of film decreased as the amount of added SDS increased. Since glycerol is highly hydrophilic and a strong humectant, the smaller glycerol content per gram of film resulted in smaller amounts of bound water, and smaller MC values. Therefore, the presence of SDS can possibly reduce

the susceptibility of protein films to RH changes.

TSM can be viewed as a measure of the water resistance and the integrity of a film. Films with 10% or more SDS had notably greater ($P < 0.05$) TSM values than control SPI films (Table 1). For example, films with 40% SDS had greater TSM than control films by about 46%. The larger solubility of the SDS-containing films was probably linked to the "weaker" structure of such films as evidenced by their smaller TS values compared with control SPI films.

3.3. Water vapor permeability

Films containing 10% or more SDS had smaller ($P < 0.05$) WVP values than control SPI films (Table 2). The mean WVP value of films with 40% SDS was almost one-half that of the control films. Similar to MC values, this was likely due to the hydrophobic portions of the SDS molecules, which decreased the rate of sorption and diffusion of water molecules through the film structures. Poor water vapor barrier ability is generally considered a major limitation to the use of protein films as packagings (Gennadios et al., 1994a; Krochta and De Mulder-Johnston, 1997). Therefore, the reduction in film WVP accompanying SDS addition is desirable for the functionality of SPI films. Gennadios et al. (1998b) reported that incorporation of saturated fatty acids such as lauric acid, myristic acid, and palmitic acid also re-

Table 2. Water vapor permeability (WVP) of cast films from soy protein isolate (SPI) treated with different levels of sodium dodecyl sulfate (SDS)

SDS (% w/w of SPI)	Thickness (μm)	WVP ($\text{g mm/m}^2 \text{ h kPa}$) ^a	RH inside cup (%) ^{a,b}
0	72 \pm 2	10.1 \pm 0.5 <i>a</i>	66.2 \pm 0.5 <i>f</i>
5	84 \pm 3	10.2 \pm 0.8 <i>a</i>	67.9 \pm 0.7 <i>e</i>
10	84 \pm 1	8.6 \pm 0.3 <i>bc</i>	69.9 \pm 0.3 <i>d</i>
20	86 \pm 8	7.8 \pm 0.9 <i>a</i>	71.4 \pm 0.3 <i>c</i>
30	79 \pm 2	6.4 \pm 0.1 <i>d</i>	72.8 \pm 0.2 <i>b</i>
40	83 \pm 8	5.6 \pm 0.4 <i>d</i>	75.2 \pm 0.5 <i>a</i>

a. Means of three replicates \pm standard deviations. Means in the same column followed by the same letter are not significantly ($P > 0.05$) different by Duncan's multiple range test.

b. Actual RH values at film surfaces facing the water calculated as described by Gennadios et al. (1994b) to account for resistance of stagnant air layer between film and water surface in testing cups. RH outside cups was 50%.

duced the WVP of glycerin-plasticized SPI films. However, such films had poor structural integrity as evidenced by their lower TS and *E* values compared with control SPI films. In contrast, SDS successfully reduced WVP while also increasing film extendibility. Nevertheless, the WVP values of SDS-containing SPI films were still two to four orders of magnitude greater than the WVP values reported for polyethylene, polypropylene, and polyester type polymeric films (Briston, 1988).

3.4. Color

The effects of SDS addition on *L* and *a* color values of SPI films were practically inconsequential (Table 3). However, films with large amounts (20, 30, or 40%) of SDS had greater ($P < 0.05$) + *b* color values (increased yellowness) (Table 3). Increased yellowness (greater *b* color values) of SPI films as a result of heat curing (Gennadios et al., 1996), UV radiation (Gennadios et al., 1998a), or cross-linking with dialdehyde starch (Rhim et al., 1998) has been reported. In general, edible/biodegradable films should be as close to colorless as possible to simulate the appearance of common polymeric films. However, the increased yellowness of SPI films with large amounts of SDS, which was attributed to the inherent color of SDS, was not considered visually detrimental.

Table 3. *L*, *a*, and *b* color values of cast films from soy protein isolate (SPI) treated with different levels of sodium dodecyl sulfate (SDS)

SDS (% w/w of SPI)	<i>L</i> ^a	<i>a</i> ^a	<i>b</i> ^a
0	90.04 ± 0.28 <i>a</i>	-3.32 ± 0.15 <i>d</i>	15.95 ± 0.83 <i>c</i>
5	92.48 ± 0.18 <i>b</i>	-3.02 ± 0.05 <i>bc</i>	16.25 ± 0.41 <i>c</i>
10	92.68 ± 0.09 <i>ab</i>	-3.51 ± 0.10 <i>e</i>	17.13 ± 0.53 <i>c</i>
20	91.06 ± 0.27 <i>c</i>	-2.98 ± 0.08 <i>ab</i>	19.31 ± 0.87 <i>b</i>
30	90.62 ± 0.08 <i>d</i>	-2.84 ± 0.01 <i>a</i>	19.36 ± 0.19 <i>b</i>
40	90.45 ± 0.32 <i>d</i>	-3.17 ± 0.03 <i>cd</i>	22.14 ± 0.74 <i>a</i>

a. Means of three replicates ± standard deviations. Means in the same column followed by the same letter are not significantly ($P > 0.05$) different by Duncan's multiple range test.

4. Conclusions

Functional properties of glycerin-plasticized SPI films can be modified by incorporation of SDS. Specifically, SDS appears to increase substantially film extendibility while simultaneously improving the moisture barrier ability of the films. These are both desirable improvements in considering such films for packaging applications. However, these improvements are realized at the expense of reduced film tensile strength.

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