

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Faculty Publications from the Harold W. Manter
Laboratory of Parasitology

Parasitology, Harold W. Manter Laboratory of

4-2003

Survey for Coccidia and Haemosporidia in the Lesser Prairie Chicken (*Typanuchus pallidicinctus*) from New Mexico with the Description of a New *Eimeria* Species

B. H. Smith

University of New Mexico, bhsmith@unm.edu

Donald Duszynski

University of New Mexico, eimeria@unm.edu

K. Johnson

University of New Mexico

Follow this and additional works at: <http://digitalcommons.unl.edu/parasitologyfacpubs>



Part of the [Parasitology Commons](#)

Smith, B. H.; Duszynski, Donald; and Johnson, K., "Survey for Coccidia and Haemosporidia in the Lesser Prairie Chicken (*Typanuchus pallidicinctus*) from New Mexico with the Description of a New *Eimeria* Species" (2003). *Faculty Publications from the Harold W. Manter Laboratory of Parasitology*. 194.
<http://digitalcommons.unl.edu/parasitologyfacpubs/194>

This Article is brought to you for free and open access by the Parasitology, Harold W. Manter Laboratory of at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Faculty Publications from the Harold W. Manter Laboratory of Parasitology by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

SURVEY FOR COCCIDIA AND HAEMOSPORIDIA IN THE LESSER PRAIRIE-CHICKEN (*TYMPANUCHUS PALLIDICINCTUS*) FROM NEW MEXICO WITH DESCRIPTION OF A NEW *EIMERIA* SPECIES

B. H. Smith,^{1,2} D. W. Duszynski,¹ and K. Johnson¹

¹ Department of Biology, The University of New Mexico, Albuquerque, New Mexico 87131, USA

² Corresponding author (email: bhsmith@unm.edu)

ABSTRACT: Blood films and fecal samples of the lesser prairie-chicken (*Tympanuchus pallidicinctus*) were examined for parasites when we surveyed specimens captured during a radio-tracking study conducted in Chaves County, New Mexico (USA). All birds were captured on the Caprock Wildlife Habitat Management Area, administered by the Bureau of Land Management. Samples were collected in late March, April, and early May 1998–2000. Oocysts were detected in five of 64 (8%) birds sampled and, upon sporulation, were determined to be an *Eimeria* species. This is the first eimerian reported from the lesser prairie-chicken and is described here as a new species. Sporulated oocysts are ellipsoidal, 27.1×22.7 (22–32×18–26) μm, with micropyle absent, but oocyst residuum and polar granule present. Sporocysts are ovoidal, 11.9×7.8 (10–14×6–10); a Stieda body, and sporocyst residuum are present, as is a small, indistinct substieda body. Inspection of blood smears revealed four cases of *Plasmodium* infection of 32 (13%) individuals sampled. The characteristics of this plasmodiid are consistent with the description of *Plasmodium (Giovannolaia) pedioecetii*, previously found in *T. pallidicinctus* (Stabler, 1978).

Key words: *Eimeria tympanuchi*, lesser prairie-chicken, new species, *Plasmodium (Giovannolaia) pedioecetii*, *Tympanuchus pallidicinctus*.

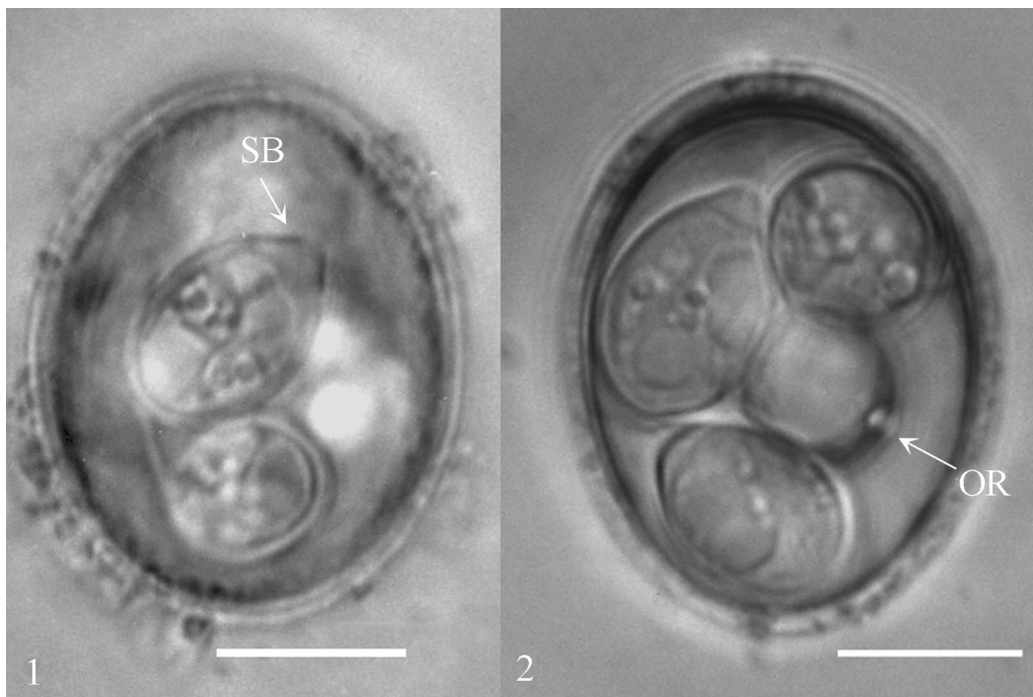
INTRODUCTION

The lesser prairie-chicken *Tympanuchus pallidicinctus* (Phasianidae: Tetraoninae) is a lekking grouse species found in the dry short grass prairie of Colorado, Kansas, New Mexico, Oklahoma, and Texas (USA). In New Mexico, the shinnery oak (*Quercus havardii*) shrublands of the Staked Plain are the principal breeding grounds. A 4 yr trapping and banding study of *T. pallidicinctus* was conducted in Chaves County, eastern New Mexico. In the last 3 yr of the study, blood and fecal samples were collected in the field. This population of prairie-chickens inhabits rangeland managed for wildlife, grazing, and oil and gas development. Recent declines in *T. pallidicinctus* populations are a consequence of habitat alteration and drought (Giesen, 1998). Chemical control of shinnery oak has further reduced available nesting and brood-rearing habitat. The species is a candidate for federal protection as threatened by the United States Fish and Wildlife Service. This study found the eastern New Mexico population of *T. pallidicinctus* positive for *Eimeria*

and *Plasmodium* species. Parasite load is not currently perceived as a significant contributor to lesser prairie-chicken mortality. However, if remaining lesser prairie-chicken populations become concentrated into smaller areas of acceptable habitat, parasite transmission rates could rise (Dobson and May, 1986).

MATERIALS AND METHODS

Lesser prairie-chickens were trapped in the wild on the Bureau of Land Management administered Caprock Wildlife Habitat Management Area, Chaves County, New Mexico (33°28'22"N, 103°47'44"W). Members of four leks were sampled for blood and fecal parasites (Johnson and Smith, 1998, 1999). All birds screened for parasites were trapped within 3,200 ha of contiguous habitat. This constitutes a small percentage of the extant New Mexico population. We trapped birds in circular, welded-wire, walk-in traps connected by chicken-wire drift fences (Toepfer et al., 1987). Samples were taken in late March, April, and early May, from 1998–2000. Average capture time was 6:30 AM (0500–08:15 AM, n=56), with only one bird captured after 8:00 AM. Each bird was temporarily detained in a small cardboard box. Fecal samples were collected from holding boxes, or during the banding and release process, and transported to the University of New



FIGURES 1, 2. Photomicrographs of sporulated oocysts of *Eimeria tympanuchi*. SB=Stieda body, OR=oocyst residuum. Bar=10 μ m.

Mexico (UNM; Albuquerque, New Mexico). Morphologic data including weight, tarsus length, wing chord, culmen depth, and length of pinna were taken on each donor bird, as well as age, sex, date, time, and location of capture. The unique identity of each bird was kept via a numbered aluminum leg band provided by the New Mexico Department of Game and Fish.

Fecal samples were placed in separate vials containing 2% aqueous (w/v) potassium dichromate ($K_2Cr_2O_7$). In the laboratory each sample was incubated at room temperature and processed following the methods outlined in Duszynski and Wilber (1997). All samples were examined and oocysts measured and photographed within 1 yr of collection. Photosyntheses of sporulated oocysts are deposited in the United States National Parasite Collection (USNPC), Beltsville, Maryland (USA) following Bandoni and Duszynski (1988) and Duszynski (1999).

We drew 10–50 μ l of blood from captured birds using brachial venipuncture. Smears of whole blood were made in the field. Each slide was fixed in 100% methyl alcohol to preserve the cells, and later were stained with Giemsa's blood stain. Smears were scanned under oil immersion (1,000 \times) to detect the presence of blood parasites. Blood films were scanned for

20 min (≥ 150 fields/film) for hematozoa, including *Plasmodium*, *Haemoproteus*, *Leucocytozoon*, trypanosomes, and microfilarie. Prevalence of parasitemias was determined from four randomly selected counts of 500 erythrocytes ($n=2,000$) per positive slide (Godfrey et al., 1987). Positive slides were read extensively in order to best describe life stage characteristics. All measurements of *Eimeria* and *Plasmodium* species are in micrometers with means followed by ranges in parentheses.

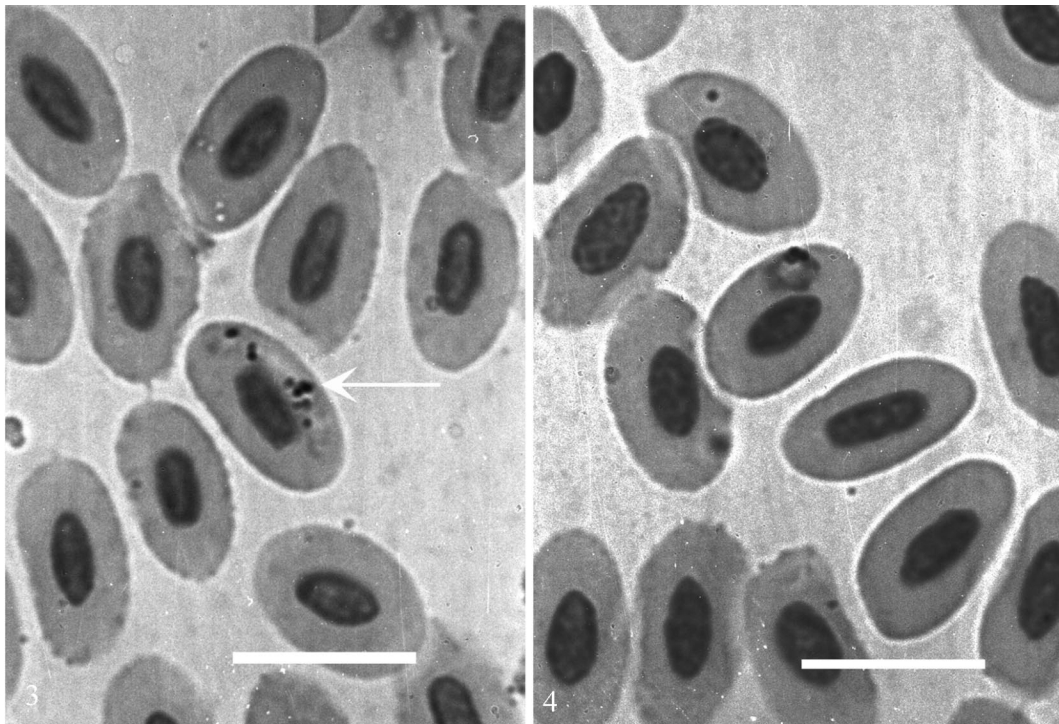
RESULTS

The birds we examined were infected with one *Eimeria* species, described here as new, and with one malarial species, *Plasmodium* (*Giovannolaia*) *pedioecetii* first described by Shillinger (1942), emended by Stabler et al. (1973).

Eimeria tympanuchi n. sp. (Figs. 1, 2, 5)

Description

Sporulated oocysts ($n=56$) ellipsoidal, 27.1 \times 22.7 (22–32 \times 18–26); shape index (SI=length/width)=1.2 (1.0–1.5); wall smooth in optical cross-section, 1.8 (1.0–



FIGURES 3, 4. Photomicrographs of *Plasmodium* (*Giovannolaia*) *pedioecetii* macrogametocyte (3, arrow) and ring stage (4). Bar=10 μ m.

2.5) thick, with two layers; outer, lightly striated, $\sim 2/3$ of total thickness; micropyle absent; usually one small, refractile polar granule present; two to five oocyst residual bodies present as round, opaque spheres. Sporocysts ($n=56$) ovoidal, 11.9×7.8 ($10-14 \times 6-10$) with L:W ratio 1.5 (1.2–2.0); Stieda body present at pointed end of sporocyst, with evidence that a small, indistinct substieda body also may be present immediately below it; sporocyst residuum present, comprised of small uniformly shaped spheres in a cluster, approximately two, partially obscuring sporozoites; sporozoites with two refractile bodies, a larger one at the blunt end and a second, smaller body at pointed end with a nucleus visible between them.

Taxonomic summary

Type host: *Tympanuchus pallidicinctus* (Ridgway, 1885), lesser prairie-chicken (Phasianidae: Tetraoninae).

Type locality: USA, New Mexico, Chaves County, $33^{\circ}28'22''N$, $103^{\circ}47'44''W$.

Prevalence: 5/64 (8%).

Site of infection: Unknown, oocysts recovered from feces.

Material deposited: Photosyntypes of sporulated oocysts in the USNPC, No. 092386.00.

Etymology: The specific name is derived from the generic name of the host.

Remarks: There are now 15 *Eimeria* species described from grouse and ptarmigan (Phasianidae: Tetraoninae) (Table 1). The sporulated oocysts of *E. tympanuchi* are distinguished from the others by the presence of a distinct oocyst residuum, lacking a micropyle, and the presence of both Stieda and substieda bodies (see Table 1).

Plasmodium (*Giovannolaia*) *pedioecetii* (Shillinger, 1942) Stabler et al., 1973 (Figs. 3, 4)

Plasmodium was detected in five of 34 (17%) samples. The samples come from 32

TABLE 1. Mensural characters of sporulated oocysts of *Eimeria* species known from the subfamily Tetraoninae as given in the original description (NG = measurements, structures not given/mentioned) and/or in subsequent re-descriptions of sporulated oocysts.

Host/ <i>Eimeria</i> spp.	Oocyst		Sporocyst		Qualitative wall structures			Author(s)
	Length×width (range)	L/W (range)	Length×width (range)	L/W (range)	Thickness (μm)	Present ¹		
<i>Bonasa umbellus</i>								
<i>E. angusta</i>	32.5×17.1 (28–37×15–19)	1.9 (1.7–2.2)	14.8×6.1 (12–17×5–8)	2.4 (2.0–2.9)	1.5–2.0	M, PC, SB, SR	Allen, 1934; Todd and Lepp, 1970	
<i>E. bonasae</i>	21.6×20.6 (18–25×18–23)	1.05 (1.0–1.2)	12.8×7.1 (10–14×6–8)	1.8 (1.6–2.0)	1.5	PG, SB, SR	Allen, 1934; Todd and Lepp, 1970	
<i>Centrocercus urophasianus</i>								
<i>E. centroceri</i>	22.6×17.1 (17–25×13–18)	1.3 NG	11.8×7.6 (11–13×7–8)	1.6 NG	?	M(?), PG, SB, SR	Simon, 1939; Honess and Post, 1955	
<i>E. pattersoni</i>	20.2×13.5 (18–23×12–15)	1.5 NG	NG	NG	?	M(?)	Honess and Post, 1955	
<i>Lagopus leucurus</i>								
<i>E. leucuri</i>	26.6×17.7 (22–31.5×15–20)	1.5 NG	15.4×6.7 (13–18×6–7)	2.3 NG	1.5	M, MC, PC, SB, SR	Stabler et al., 1979	
<i>E. oreocetes</i>	26.0×22.6 (23–29×20–26)	1.2 NG	14.6×8.8 (13–16×8–10)	1.6 NG	2.0	PC, SB, SSB, SR	Stabler et al., 1979	
<i>Lagopus mutus</i>								
<i>E. brinkmanni</i>	28.6×18.8 (26–30×18–20)	1.5 (1.4–1.6)	13×7	1.9 NG	NG	PC, SB	Levine, 1953	
<i>E. fanthami</i>	28.3×18.8 (27–29×18–20)	1.5 (1.4–1.6)	NG	NG	NG	PG, SB	Levine, 1953	
<i>E. lagapodi</i>	24×15 NG	1.6 NG	12×12 NG	1.0 NG	NG	M(?)	Galli-Valerio, 1929	
<i>Tetrao tetrix</i>								
<i>E. lyruri</i>	29.6×15.3 (22–37×12–20)	1.9 NG	9.8×3.6 NG	2.7 NG	NG	PC, SR	Galli-Valerio, 1927; Yakimoff and Gousseff, 1936	
<i>E. nadsoni</i>	24.9×21.3 (21–29×17–24)	1.2 NG	12.2×10.9 NG	1.1 NG	1.5–2.0	PC, SB(?)	Yakimoff and Gousseff, 1936	
<i>E. tetricis</i>	31×15 (30–31×15–15)	2.1 NG	NG	NG	1.2	M, MC(?), SB, SR	Haase, 1939	

TABLE 1. Continued.

Host/ <i>Eimeria</i> spp.	Oocyst		Sporocyst		Qualitative wall structures		Author(s)
	Length×width (range)	L/W (range)	Length×width (range)	L/W (range)	Thickness	Present ¹	
<i>T. urogallus</i>							
<i>E. ventriosa</i>	32×22 (31–33×20–23)	1.4 NG	NG	NG	1.0	M, PC, SB, SR	Haase, 1939
<i>E. yakisevi</i>	20 (24–19.5)	NG	10.8×5.1 NG	2.1 NG	NG		Yakimoff and Gousseff, 1936; Hardcastle, 1943
<i>Tympanuchus pallidicinctus</i>							
<i>E. tympanuchi</i>	27.1×22.7 (22–32×18–26)	1.2 (1.0–1.5)	11.9×7.8 (10–14×6–10)	1.5 (1.2–2.0)	1.8	OR, PC, SR, SB, SSB	This study

¹ Oocyst and sporocyst qualitative structures present in each species. Abbreviations: M=micropyle; MC=micropyle cap; OR=oocyst residuum; PC=1 or more polar granules; SB=Stieda body; SSB=substieda body; SR=sporocyst residuum. If not stated in the original description or the re-description, this structure is presumed to be absent although this is not always certain in some of the older work.

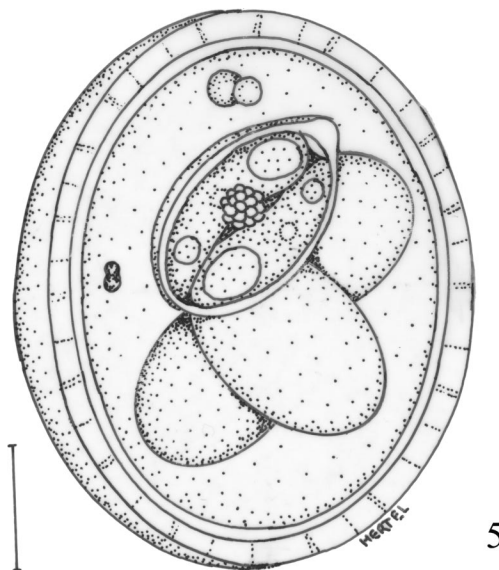


FIGURE 5. Line drawing of a sporulated oocyst of *Eimeria tympanuchi*. Bar=6 μ m.

subjects, because two birds were captured and screened in successive years. One individual was found positive in successive years, the only infected bird either recaptured or resighted between years. Therefore, four of 32 possible hosts were infected (13%). Intensity of each case (0.005–0.007, $n=5$) suggests hosts persisted with chronic infections.

Trophozoites either were ameboid (78%) with one to five ($\bar{x}=1.5$) small pigment granules or were ring-shaped (22%), with central vacuoles and one to four aggregated pigment granules (Fig. 4); 70% of all trophozoites seen were subpolar or polar.

Gametocytes ($n=29$) appeared immature and measured 7.5×1.9 ($6.25-10 \times 1.25-2.5$); 25/29 (86%) were found on the lateral edge of the nucleus without apparent alteration of nuclear position or cell wall. Pigment granules, 3.4 (1–11), were often on the edge of developing gametocytes. Mature gametocytes (Fig. 3) were elongate and narrow, with irregular edges and with pigment granules throughout their cytoplasm. The ends of the mature gametocyte usually curled around the polar ends of the nucleus without distortion

of erythrocyte shape. Only one mature segmenter was seen with nine merozoites visible in a tight cluster. It did not distort the erythrocyte's shape or the position of its nucleus.

The stage of the life cycle most evident was ameboid trophozoites in erythrocytes (60%). The gamont life stages were often immature (66%), hence the observation of mean length and width falls beneath the stated means of Stabler (1978). Mature gametocyte dimensions and morphology are consistent with *P. (Giovannolaia) pedioecetii*. Length/width measurements of a subset of nine mature gametocytes, 9.5×2.1, almost exactly reflects the means reported by Stabler (1978), 9.2×2.0. Gametocytes from the present study, however, were more prone to lateral orientation (85%) than those reported by Stabler (33.8%), in 1978.

DISCUSSION

There are currently 15 *Eimeria* spp. described from the subfamily Tetraoninae, including the new species described here. The systematics of grouse and ptarmigan were recently amended, and this paper follows the American Ornithologists' Union (1998) classification of species within Tetraoninae. The Tetraoninae has six genera including *Tympanuchus*, which has three species: *T. cupido*, *T. pallidicinctus*, and *T. phasianellus*. The lack of clear genetic differences within this species complex (Ellsworth et al., 1996) and proximity of historic range, suggested a comparison of existing eimerians of closely related species. In a comparison of the characters of sporulated oocysts from Tetraonid hosts, the oocyst residuum of *E. tympanuchi* is novel. There is no report of an oocyst residuum present in any of the *Eimeria* species found in hosts of the subfamily Tetraoninae.

Light infections of the hematozoan *Plasmodium (Giovannolaia) pedioecetii* were found. This species was previously described from lesser prairie-chickens of New Mexico and Texas by Stabler (1978).

In this study there was a low prevalence of schizonts and mature gamonts. In a study of *P. (Giovannolaia) pedioecetii* from galliformes, Stabler and Kitzmiller (1976) reported a daily peak in merozoite production from 8:00 AM to noon. The earlier average capture time of birds sampled in our study (0500–08:15 AM) may explain the lack of segmenters and large number of underdeveloped gametocytes.

ACKNOWLEDGMENTS

We thank the Bureau of Land Management, Roswell, New Mexico Field Office for funding the majority of this study. Grants from T&E, Inc. and the New Mexico Department of Game and Fish Share With Wildlife program allowed us to complete the 2000 field season. We thank the following for trapping work: M. Berry, D. Bilyeu, M. Kline, B. Long, J. Montgomery, M. Radke, and C. Westwood and I. Asmundsson, L. Couch, K. Decker, J. Hnida, A. Lynch, and M. Ryan for assistance in the laboratory. We are especially grateful to L. Hertel for the line drawing.

LITERATURE CITED

- ALLEN, E. A. 1934. *Eimeria angusta* sp. nov. and *Eimeria bonasae* sp. nov. from grouse, with a key to the species of *Eimeria* in birds. Transactions of the American Microscopical Society 53: 1–5.
- AMERICAN ORNITHOLOGISTS' UNION. 1998. Checklist of North American birds, 7th Edition. American Ornithologists' Union, Washington, D.C., 829 pp.
- BANDONI, S., AND D. W. DUSZYNSKI. 1988. A plea for improved presentation of type material for coccidia. Journal of Parasitology 74: 519–523.
- DOBSON, A. P., AND R. M. MAY. 1986. Disease and conservation. In Conservation biology: The science of scarcity and diversity. M. E. Soule (ed.). Sinauer Press, Sunderland, Massachusetts, pp. 345–365.
- DUSZYNSKI, D. W. 1999. Revisiting the Code: Clarifying name-bearing types for photomicrographs. Journal of Parasitology 85: 769–770.
- , AND P. G. WILBER. 1997. A guideline for the preparation of species descriptions in the Eimeriidae. Journal of Parasitology 83: 333–336.
- ELLSWORTH, D. L., R. L. HONEYCUTT, AND N. J. SILVY. 1996. Systematics of grouse and ptarmigan determined by nucleotide sequences of the mitochondrial cytochrome-b gene. The Auk 113: 811–822.
- GALLI-VALERIO, B. 1927. Notes de parasitologie et de technique parasitologique. Zentralblatt für

- Bakteriologie. I. Abteilung Originale 103: 177–182.
- . 1929. Notes de parasitologie. Zentralblatt für Bakteriologie. I. Abteilung Originale 112: 54–59.
- GIESEN, K. M. 1998. Lesser prairie-chicken (*Tympanuchus pallidicinctus*) In The birds of North America, Number 364. A. Poole and F. Gill (eds.). The Birds of North America, Inc., Philadelphia, Pennsylvania, 17 pp.
- GODFREY, JR., R. D., A. M. FEDYNICH, AND D. B. PENCE. 1987. Quantification of hematozoa in blood smears. Journal of Wildlife Diseases 23: 558–565.
- HAASE, A. 1939. Untersuchungen über die bei deutschen Wildhühnern vorkommenden *Eimeria*-Arten. Archiv für Protistenkunde 92: 329–383.
- HARDCASTLE, A. B. 1943. A check-list and host index of the species of the protozoan genus *Eimeria*. Proceedings of the Helminthological Society of Washington 10: 35–69.
- HONESS, R. F., AND G. POST. 1955. *Eimeria* of grouse (Family: Tetraonidae), with a description of *Eimeria pattersoni* n. sp. from the sage grouse. Wyoming Game & Fish Commission, Bulletin 8, Article 2: 5–11.
- JOHNSON, K., AND B. H. SMITH. 1998. Radio telemetry study of lesser prairie-chicken habitat use in the Caprock Wildlife Habitat Management Area. New Mexico Natural Heritage Program technical report to the Bureau of Land Management, 17 pp.
- , AND ———. 1999. Lesser prairie-chicken habitat use on the Sand Ranch and population status in the Caprock Wildlife Habitat Management Area, 1999. New Mexico Natural Heritage Program technical report to the Bureau of Land Management, 19 pp. and maps.
- LEVINE, N. D. 1953. A review of the coccidia from the avian orders Galliformes, Anseriformes and Charadriiformes, with descriptions of three new species. American Midland Naturalist 49: 696–719.
- SCHILLINGER, J. E. 1942. Diseases of wildlife and their relationship to domestic livestock. Yearbook of agriculture, USDA, US Government Printing Office, pp. 1217–1225.
- SIMON, F. 1939. *Eimeria centrocerci* n. sp. du *Centrocerus urophasianus* (coq du Bruyère). Annales de Parasitologie Humaine et Comparée, Paris 17: 137–138.
- STABLER, R. M. 1978. *Plasmodium (Giovannolaia) pedioecetii* from the lesser prairie-chicken, *Tympanuchus pallidicinctus*. Journal of Parasitology 64: 1125–1126.
- , AND N. J. KITZMILLER. 1976. *Plasmodium (Giovannolaia) pedioecettii* from gallinaceous birds of Colorado. Journal of Parasitology 62: 539–544.
- , ———, AND C. E. BRAUN. 1973. *Plasmodium* in a Darwin's tinamou from Colorado. Journal of Parasitology 59: 395.
- , A. G. HASKINS, N. J. KITZMILLER, O. W. OLSEN, AND C. E. BRAUN. 1979. Two new species of coccidia, *Eimeria leucuri* and *E. oreocetes* (Protozoa: Eimeriidae), in grouse from Colorado. Journal of Parasitology 65: 272–274.
- TODD, JR., K. S., AND D. L. LEPP. 1970. Redescription of *Eimeria angusta* Allen, 1934 and *E. bonasae* Allen, 1934 (Protozoa: Eimeriidae) from the ruffed grouse *Bonasa umbellus*. Journal of Protozoology 17: 427–429.
- TOEPFER, J. E., J. A. NEWELL, AND J. MONARCH. 1987. A method for trapping prairie grouse hens on display grounds. Montana Agriculture Experiment Station, Contribution #2144: 21–23.
- YAKIMOFF, W. L., AND W. F. GOUSSEFF. 1936. A propos des coccidies des oiseaux sauvages. Annales de Parasitologie Humaine et Comparée 14: 449–456.

Received for publication 13 May 2002.