

2004

Primaquine Therapy for Malaria

J. Kevin Baird

US Naval Medical Research Center Detachment, Lima, Peru, jkevinbaird@yahoo.com

Stephen L. Hoffman

Sanaria, Rockville, Maryland

Follow this and additional works at: <http://digitalcommons.unl.edu/publichealthresources>

Baird, J. Kevin and Hoffman, Stephen L., "Primaquine Therapy for Malaria" (2004). *Public Health Resources*. 329.
<http://digitalcommons.unl.edu/publichealthresources/329>

This Article is brought to you for free and open access by the Public Health Resources at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Public Health Resources by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Primaquine Therapy for Malaria

J. Kevin Baird and Stephen L. Hoffman

¹US Naval Medical Research Center Detachment, Lima, Peru, and ²Sanaria, Rockville, Maryland

Primaquine is the only available drug for preventing relapse of malaria, and confusion surrounds its use. This review examines the wide range of clinical applications of primaquine described in the medical literature between 1946 and 2004. The risk of relapse of *Plasmodium vivax* malaria without primaquine therapy ranged from 5% to 80% or more, depending largely upon geographic location. Supervision of therapy profoundly impacts the risk of relapse, and almost all reports of malaria resistant to primaquine are associated with lack of such supervision. We nonetheless suspect that there is widespread resistance to the standard course of primaquine therapy, which is 15 mg primaquine base daily for 14 days. Clinical evidence confirms that a course of 15 mg daily for just 5 days, a regimen widely used in areas where malaria is endemic, has no discernible efficacy. This review supports a recommendation for a regimen of 0.5 mg/kg primaquine daily for 14 days, on the basis of superior efficacy and good tolerability and safety in nonpregnant persons without glucose-6-phosphate dehydrogenase deficiency.

Malaria causes an acute, debilitating febrile syndrome that ends in death for 1.5–2.7 million of the ~500 million infected annually [1]. Just 100 years ago, malaria infected millions of people in North America, Europe, Australia, and other subtropical and temperate regions [2]. Chloroquine, primaquine, and dichlorodiphenyltrichloroethane (DDT) helped eradicate malaria from temperate latitudes and control it in the tropics [3]. Those gains have deteriorated substantially, and outbreaks occur even in the United States [4–7].

Primaquine, introduced in 1950, prevents relapse and sterilizes infectious sexual plasmodia, but confusion surrounds its use. Among the several widely used regimens, none has been adequately evaluated. Tolerance of primaquine by *Plasmodium vivax* occurs in Southeast Asia and Oceania, but the risk of therapeutic failure has been rarely documented anywhere. Poor adherence to pri-

maquine therapy and resistance to companion drugs like chloroquine, compounds the confusion. Worse still, abbreviated regimens of primaquine without proven clinical efficacy are also widely used. Finally, available evidence refutes primaquine's reputation for being toxic and poorly tolerated. This review examines these issues.

BIOLOGY

The protozoa that cause malaria belong to the apicomplexid coccidian family Plasmodiidae, genus *Plasmodium*. The genus contains 172 species, but only 4 routinely infect humans: *Plasmodium falciparum*, *P. vivax*, *Plasmodium malariae*, and *Plasmodium ovale*. Other plasmodia infect mammals, birds, and reptiles, and these rarely infect humans [8]. The plasmodia follow a similar life cycle (figure 1). The most important distinction between the 4 different *Plasmodium* species that bears on therapy is that 2 can cause relapse—parasitemia originating from hypnozoites in the liver that occurs from 16 days to several years after the primary infection. Only *P. vivax* and *P. ovale* form hypnozoites. Latent blood stages of the parasite account for the chronicity of malaria due to *P. malariae*, and malaria due to *P. falciparum* typically exhibits no chronic latency.

P. vivax is pantropical, but it is largely absent from Africa. Black Africans lack an erythrocyte surface pro-

Received 15 January 2004; accepted 15 June 2004; electronically published 12 October 2004.

The views and opinions expressed herein are those of the authors and do not purport to reflect those of the US Navy, the US Department of Defense, or Sanaria, Inc.

Reprints or correspondence: Capt. J. Kevin Baird, US Naval Medical Research Center Detachment, American Embassy Lima, APO AA 34031, USA (bairdk@nmrc.navy.mil).

Clinical Infectious Diseases 2004;39:1336–45

© 2004 by the Infectious Diseases Society of America. All rights reserved.
1058-4838/2004/3909-0009\$15.00

This document is a U.S. government work and is not subject to copyright in the United States.

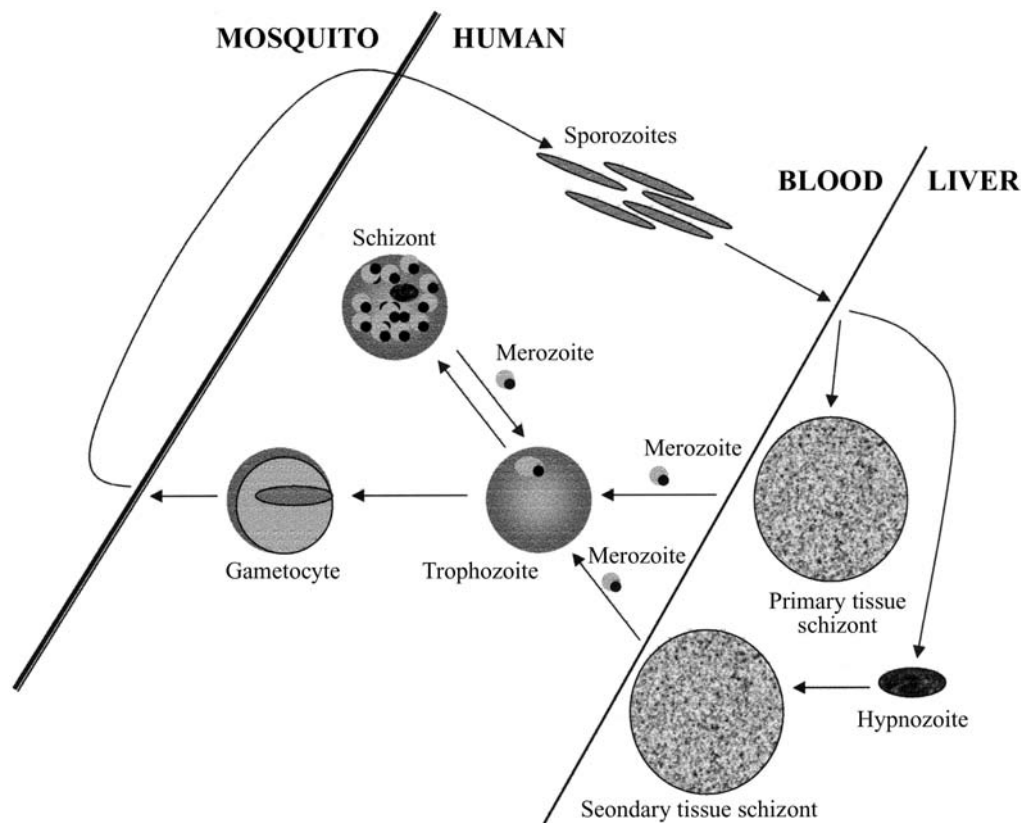


Figure 1. Life cycle of *Plasmodium* species in human beings. *Plasmodium vivax* and *Plasmodium ovale* produce both primary tissue schizonts and the hypnozoite that leads to a secondary tissue schizont, whereas *Plasmodium falciparum* and *Plasmodium malariae* produce only a primary tissue schizont. Parasitemia arising from hypnozoites is called a “relapse,” and *P. vivax* and *P. ovale* malaria are called the “relapsing malarias.” Primaquine treatment is often called “anti-relapse” therapy.

tein called Duffy factor that *P. vivax* merozoites require for invasion. Vivax malaria is especially common in India, Indochina, and the Philippine, Indonesian and New Guinean archipelagos. In the New World, vivax malaria occurs from northern Mexico to northern Argentina. Endemic *P. ovale* malaria occurs only in West Africa, The Philippines, Eastern Indonesia, and Papua New Guinea.

RELAPSE

Primaquine prevents relapse of malaria [9]. The pattern and probability of relapse in the absence of primaquine therapy varies by geographic origin. *P. vivax* malaria in temperate regions relapses at long intervals (>6 months) [10]. Among 1021 soldiers infected with *P. vivax* in Korea, 32% had relapse [11–14]. In India, the 12-month relapse rate ranged from 9% to 19% [15, 16]. All 180 subjects challenged with the North American St. Elizabeth strain of *P. vivax* had relapse 6–12 months later [17]. Experimental challenge notwithstanding, in temperate regions, the odds of relapse of *P. vivax* malaria are approximately 1 in 4 (table 1).

Infection with tropical strains of *P. vivax* is associated with a higher probability of relapse, relapse that occurs sooner, and, typically, multiple relapses. Among 54 American soldiers infected in the Pacific, all experienced relapse within 3 months [19]. Similar cohorts including a total of 562 subjects had a 5-month relapse rate of 72% [20, 21]. Of 213 subjects experimentally challenged with the Chesson strain of *P. vivax* from New Guinea [27], 99% had relapse within 8 months, most within 1 month after the primary parasitemia [17, 18]. Among 333 subjects infected with *P. vivax* from the tropical Pacific, the median time of relapse was day 22 after onset of the primary parasitemia [28]. No relapse occurred before day 16.

The probability of relapse is high (>1 in 5), regardless of where the infection is acquired, and potential exposure to infection indicates use of terminal prophylaxis (i.e., presumptive primaquine therapy). Indeed, failure to prescribe or comply with post-exposure prophylaxis accounts for most cases of vivax malaria in travelers [29]. Schwartz et al. [30] found that late onset (>2 months after exposure) occurred in 62% of 1321 vivax malaria cases, and in 63% of these cases, there was adequate adherence

Table 1. Summary of reports of relapse of *Plasmodium vivax* malaria not treated with primaquine therapy.

Reference(s)	Geographic location	<i>P. vivax</i> strain	No. of patients	Duration of follow-up	Percentage of patients with relapse
[17, 18]	New Guinea	Chesson	213	2 w to >8 mo	99
[19]	New Guinea	Wild	54	2 w to 3 mo	100
[20, 21]	Pacific	Wild	562	2–5 mo	72
[12–14]	Korea	Wild	1021	>4 mo	32
[16]	India	Wild	264	2–8 mo	19
[15]	India	Wild	5528	12 mo	11
[22]	India	Wild	222	12 mo	9
[23]	Pakistan	Wild	250	12 mo	52
[24]	Thailand	Wild	342	2 mo	63
[25]	Ethiopia	Wild	29	3 mo	50
[17]	North America	St. Elizabeth	180	12 mo	100
[26]	Global	Wild	68	NR	25

NOTE. Mo, months; NR, not reported; w, weeks.

to prescribed suppressive prophylaxis (i.e., against blood stages of the parasite). Most cases of vivax malaria among travelers are relapses and are preventable with primaquine, the only currently available drug for therapy to prevent relapse.

PRIMAQUINE THERAPY

Development. War in the Pacific in 1941 created an urgent strategic need in the United States for a drug to prevent relapse of malaria. Studies during and after World War II focused on 8-aminoquinolines, because, in the 1920s, pamaquine (the prototypical 8-aminoquinoline) had proven effective but too toxic. Thousands of compounds were screened in animals, and 21 went to clinical trials. Isopentaquine and primaquine proved superior [31]. Primaquine became available to American troops during the Korean War.

A total dose of 200 mg primaquine base (all doses of primaquine in this article refer to base, exclusive of weight of the typical diphosphate salt formulation) achieved cure, and a dose of 15 mg was well tolerated, so a 14-day regimen was adopted. According to Schmidt et al. [32], “the use of this regimen should not be construed as synonymous with necessity” (p. 1127). Provided an adequate total dose was delivered, schedule did not impact efficacy. A single 45-mg dose administered once per week for 8 weeks (360 mg) was as effective as 30 mg daily for 14 days (420 mg) or 60 mg daily for 7 days (420 mg) [33] and was more effective than 15 mg daily for 14 days (210 mg) [34].

Pharmacokinetics. Primaquine is rapidly absorbed in the gastrointestinal tract and concentrated in the liver, brain, heart, lungs, and skeletal muscle. It crosses the placenta. The mean volume of distribution is 3 L/kg. It peaks in plasma within 1–3 h, at ~70 mg/mL. It is rapidly excreted in urine, with a plasma half-life of 4–9 h. Its metabolism is complex and poorly un-

derstood [35]. Among the many known or suspected metabolites, none has been definitively linked to activity against the *Plasmodium* parasite.

Standard therapy. Primaquine therapy is given after the diagnosis of *P. vivax* or *P. ovale* malaria and should coincide with blood schizonticidal therapy. Others recommend commencing primaquine therapy after blood schizonticidal therapy, on the basis of immunosuppressive activity observed in vitro [36–38]. In vivo studies have failed to corroborate those findings [39]. Primaquine appears to be more effective when given concurrently with blood schizonticides [40–42] (table 2). Indonesians who took 30 mg daily for 1 year showed no effects on cellular immunity to tetanus toxoid [43] or on susceptibility to malaria [44].

A few studies during the past 25 years gauged therapeutic efficacy of standard primaquine therapy [45–55]. Reports up to 1977 confirmed the excellent efficacy of supervised therapy against *P. vivax* infection in regions other than New Guinea and Thailand (table 3). Of 1344 patients given supervised therapy, only 14 (1%) experienced relapse, whereas, of 2061 patients given unsupervised therapy, 449 (22%) had relapse (relative risk, 0.05; 95% CI, 0.03–0.09; $P < .0001$). Similarly, the relative risk of relapse with supervised therapy among 469 soldiers returned from Vietnam was 0.23 (95% CI, 0.12–0.46; $P < .0001$). Relapse after unsupervised therapy [56, 58–67] does not prove resistance.

Nonetheless, primaquine does not remain universally effective. During the past 10 years, we heard from clinicians around the world complaining of more frequent failures of primaquine therapy. Even though no unambiguous evidence of resistance yet exists, experience teaches us to heed such warnings. We routinely advise against using the standard regimen and instead

Table 2. Summary of relapse rates after standard primaquine therapy in early reports of infection with the Chesson strain of *Plasmodium vivax*.

Reference	Primaquine dosage	Concurrent blood schizonticidal therapy	No. of patients	Duration of follow-up, months	Percentage of patients with relapse
[41]	15 mg/day for 14 days	None	19	12	79
[18]	15 mg/day for 14 days	Quinine	24	12	21
[34]	15 mg/day for 14 days	Chloroquine	79	12	27
[18]	22.5 mg/day for 14 days	None	5	12	80
[18]	22.5 mg/day for 14 days	Quinine	31	12	3
[18]	30 mg/day for 14 days	Quinine	5	12	0
[33]	60 mg/day for 7 days	Chloroquine	11	1–14	0
[34]	45 mg/week for 8 weeks	Chloroquine	71	12	8

recommend the alternative of 0.5 mg/kg daily for 14 days. This now concurs with recommendations by the Centers for Disease Control and Prevention (unpublished data, CDC).

Failure of standard therapy. Evidence for failure of the standard primaquine regimen emerged from early experimental challenge with the Chesson strain of *P. vivax* isolated from an American soldier infected in New Guinea in 1944 [27]. Table 2 summarizes that work of 50 years ago. Relapse occurred in 26 of 103 subjects treated with chloroquine or quinine and the standard primaquine regimen of 15 mg, whereas only 1 of 36 subjects treated with primaquine regimens of 22.5 or 30 mg daily for 14 days had relapse. The relative risk associated with the higher dose was 0.11 (95% CI, 0.02–0.78; $P < .005$). More recent trials in Thailand demonstrated similar findings: 7 (18%) of 81 patients given a standard dosage of chloroquine and a 15-mg primaquine regimen had relapse within 6 months, whereas only 1 of 86 treated with a 22.5-mg regimen had relapse [46]. In the other study [47], 10 (17%) of 60 patients given standard primaquine therapy had relapse. The relative risk of relapse associated with the 22.5-mg regimen, compared with the 15-mg regimen, was 0.1 (95% CI, 0.01–0.71; $P < .005$). A trial found that a regimen of 30 mg daily for 14 days combined with atovaquone and proguanil (Malarone; GlaxoSmithKline) was efficacious against *P. vivax* infection in 46 Thai patients, of whom 35 were followed up for 12 weeks, and there were just 2 cases of recurrent parasitemia (94% efficacy) [68]. Jelinek et al. [59] demonstrated that infection acquired on the island of New Guinea had a 12-fold higher risk of relapse after primaquine therapy. Only 3 of 44 who did not travel to New Guinea experienced relapse, whereas 4 of 5 persons who did travel to New Guinea experienced relapse (OR, 11.7; 95% CI, 1.6–100; $P < .001$). Duarte et al. [69] report the only recent study of therapeutic response to standard primaquine therapy among patients with vivax malaria in the New World: 7 (14%) of 50 patients given supervised therapy had relapse within 6 months.

Vivax malaria should be treated with a primaquine regimen

of ≥ 22.5 mg daily for 14 days (we favor 30 mg), or a total dose of ≥ 315 mg for as long as 8 weeks [29, 42, 43, 48, 49]. Schwartz and colleagues [70] demonstrated a high risk of relapse after standard “adult-dose” therapy (15 mg daily for 14 days) among Israeli patients weighing >80 kg. Medical officers of the New Zealand armed forces described the same phenomenon among troops who returned from East Timor (unpublished data). Patients heavier than 70 kg should receive at least 0.5 mg/kg daily.

For patients who are pregnant or who have well-documented failure of recommended primaquine therapy, no currently available alternative therapies exist. The risk of relapse in these patients should be managed with suppressive therapy with chloroquine or mefloquine for at least 4 weeks, preferably 8 weeks. These patients certainly should be counseled regarding their risk of relapse beyond 8 weeks.

Resistance to primaquine. Resistance to primaquine by blood stages of the *Plasmodium* parasite [71] is of little clinical consequence. Resistance in tissue stages dominates public health concern, and the absence of such resistance after 50 years seems incredible. There may be compelling physical, chemical, or biological reasons; for example, short plasma half-life or sterilization of gametocytes. Alternatively, clinical evidence of resistance to primaquine may be present but difficult to detect. We favor this explanation.

Proof of resistance requires addressing important confounding factors. Use of directly observed therapy administered by reliable people addresses the most important of these. Patients exposed to risk of infection after treatment should be excluded from analysis. Resistance to chloroquine by *P. vivax* [72–75] must be considered; recurrent parasitemia may be recrudescence of a chloroquine-resistant strain, rather than relapse due to a primaquine-resistant strain. Clinical trials designed to detect resistance to primaquine should use an effective blood schizonticide with a short plasma half-life, such as quinine, which will rule out the possibilities of suppression of early

Table 3. Summary of reports of relapse of *Plasmodium vivax* malaria after treatment with a standard regimen of primaquine therapy (15 mg per day for 14 days).

Type of therapy, geographic location	Year	No. of patients	Duration of follow-up, months	Percentage of patients with relapse	Reference(s)
Supervised					
Vietnam	1974	218	12	4	[45]
Thailand	1994	141	1–18	18	[46, 47]
Solomon Islands	1977	10	12	0	[48]
Korea	1953	914	4–20	<1	[49, 50]
Nicaragua	1953	145	4	0	[51]
Central America	1974	57	9–36	4	[52]
Unsupervised					
Vietnam	1970	251	12	18	[53, 54]
Brazil	1991	1347	3–12	25	[55]
Somalia	1997	60	NR	43	[29]
Global	1995	57	>18	13	[59]
Global	1987	175	NR	10	[26]
Global	1990	132	NR	9	[57]

NOTE. Mo, months; NR, not reported.

relapse by lingering traces of the drug in patients (i.e., false-negative responses) and recrudescence due to resistance to chloroquine (i.e., false positive responses). Table 4 summarizes key confounders of the diagnosis of infection with primaquine-resistant *P. vivax*. Collins and Jeffery [76] have reviewed resistance to and tolerance of primaquine in *P. vivax*.

Five-day regimen. A primaquine regimen of 15 mg daily for 5 days to prevent relapse of *P. vivax* malaria is national policy in many countries where the disease is endemic. In 1954, Singh et al. [77] found no cases of relapse among 50 patients treated with pyrimethamine and the 5-day regimen. Basavaraj [78] corroborated this in 1960, as did Mendoza in Mexico (World Health organization document WHO/MAL/527.65, cited by Contacos et al. [79]). Cedillos et al. [80] found reductions in the number of clinical episodes in communities where the 5-day regimen plus standard amodiaquine was given. Some studies showed low relapse rates (<10%) among large numbers of patients in India given the 5-day regimen [22, 81], but relapse rates for patients not treated with primaquine may be this low. Randomized and controlled studies are needed to prove the importance of this point.

Contacos et al. [79] treated 5 volunteers exposed to a Pakistani strain of *P. vivax* with the 5-day regimen, and all had relapse within 181 days. Miller et al. [52] also tried this regimen against the Salvador II strain of *P. vivax*, and the patient had relapse at day 223. Singh et al. [82] recorded a relapse rate of 11% among 995 patients treated with the 5-day regimen, and the relapse rate among 222 patients not treated with primaquine was 9%. Rowland and Durrani [23] reported *P. vivax* relapse rates of 52% and 51%, respectively, among 500 Pakistani patients randomized to receive treatment either with chloroquine

alone or with the 5-day, 15-mg regimen combined with chloroquine. Gogtay et al. [83] and Yadav and Ghosh [84] reported essentially similar findings from India. Villalobos-Salcedo and colleagues [85] compared 60-day relapse rates among patients in Amazonia treated with chloroquine and 15 mg of primaquine daily given for 5 or 14 days: the odds of relapse among subjects who received the 5-day regimen was 5.3 (95% CI, 0.9–40; $P < .03$) (tables 5 and 6).

ACTIVITY AGAINST BLOOD-STAGE PARASITES AND *P. OVALE*

Blood schizonticidal therapy. Primaquine is not recommended as a stand-alone blood schizonticide, but its effect upon blood-stage parasites should be understood. Primaquine monotherapy was shown to be effective against *P. vivax* parasitemia in Thailand by Pukrittayakamee et al. [88], who administered 15 mg daily for 14 days to 30 patients, and parasitemia was cleared in all. Wilairatana et al. [89] treated 23 patients infected with *P. vivax* in Thailand with 30 mg of primaquine daily for 14 days, and all patients remained a parasitemic at day 28. These studies demonstrate the apparently potent blood schizonticidal activity of therapeutic doses of primaquine against *P. vivax*.

Therapeutic doses of primaquine do not affect the asexual blood stages of *P. falciparum*. In eastern Indonesia, we found no difference in activity of primaquine against *P. falciparum* in 25 subjects given chloroquine plus primaquine (30 mg daily for 28 days) and in 28 given chloroquine plus a placebo of primaquine [90]. In 1955, Arnold et al. [91] described complete therapeutic failure of a primaquine regimen of 30 mg daily against *P. falciparum* (Panama P-F-6 strain) in 6 volunteers.

Table 4. Key confounders of the diagnosis of infection with primaquine-resistant *Plasmodium vivax*.

Confounder	Ambiguity	Solution
Noncompliance to prescribed therapy	Adequacy of dose	Supervise of therapy
Emesis of dose	Adequacy of dose	Supervise of therapy and readminister vomited doses
Parasite tolerance	Resistance vs. tolerance	Determine baseline susceptibility or use 30-mg regimen
Chloroquine resistance	Recrudescence vs. relapse	Relapse can be discounted if recurrence is <1 month after the end of therapy; or use an alternative therapy (e.g. quinine)
Reinfection	Reinfection vs. relapse	Evaluate repatriated traveler or, if necessary, evacuate to an area where malaria is not endemic

Transmission-blocking therapy. A single dose of 45 mg of primaquine is routinely prescribed for *P. falciparum* malaria in areas where it is endemic, to reduce the risk of transmission. In experimentally challenged volunteers, primaquine markedly reduced the number of circulating gametocytes and sterilized those remaining [92, 93]. This was an important finding, because effective blood schizonticidal treatment may leave surviving gametocytes [94]. This result is especially problematic when slow-acting antimalarials are used, because the brief period of primaquine gametocytocidal activity precedes elimination of trophozoites that may differentiate to gametocytes. In practice, a single dose of primaquine may accomplish little [92, 93, 95]. Kaneko et al. [96] evaluated this regimen in a village in Sumatra and documented reduced numbers of gametocyte in patients treated with primaquine, but they did not assess transmission to mosquitoes. Standard primaquine therapy against *P. vivax* resulted in rapid (4–20 h) and complete loss of transmission to mosquitoes in 5 patients infected with *P. vivax* in Brazil [97].

***P. ovale* infection.** This parasite occurs in West Africa, The Philippines, eastern Indonesia, and New Guinea. The foci in the Pacific have exceedingly low but consistent frequencies of infection [98]. Diagnosis of *P. ovale* infection should be supported by agreement among expert microscopists or by molecular biological evidence [99, 100]. Primaquine therapy for *P. ovale* infection is as for *P. vivax* infection. Therapeutic failure of primaquine against *P. ovale* has been reported [101], but only in patients who did not receive directly observed therapy.

TOLERABILITY AND TOXICITY

Gastrointestinal upset. We do not accept the view that primaquine is toxic and poorly tolerated. Compared with other antimalarials, it has good tolerability and safety in people considered good candidates to receive it. Lethal doses of 8-aminoquinolines in animals exhibited pronounced hepatotoxicity [31], and lesser doses showed hematological and gastrointestinal effects, primarily epigastric discomfort [17, 102]. However, studies of therapeutic doses demonstrate good tolerability and safety. Clayman et al. [103] observed abdominal distress in human subjects who had fasted and received a single dose of

primaquine: it was reported by 5% of subjects who received a 15-mg dose, by 10% who received a 30-mg dose, 35% who received a 60-mg dose, and 100% who received a 90-mg dose. The drug was tolerated without complaint in subjects who had eaten, even at the highest doses administered.

The 15-mg dose of primaquine consistently shows good tolerability [12, 14, 18], and few complaints occur at higher doses. Clyde and McCarthy [33] administered 60 mg daily for 7 days to 11 men and described the adverse effects as “negligible in 9 men, and 2 others consisted of moderate abdominal cramps and nausea toward the end of the course” (p. 563). Baird et al. [42] administered two 60-mg and one 30-mg dose concurrent with chloroquine therapy to 22 subjects in Indonesia; physical complaints were no more frequent than among 23 other subjects receiving chloroquine and a placebo of primaquine. Among 5 subjects given a 14-day, 30-mg regimen, Edgecomb et al. [18] observed mild transient adverse effects in only 1 subject. In a trial of a regimen of 30 mg daily for prophylaxis in Indonesia, 43 men continued taking the regimen for 1 full year and had no more complaints than did 42 subjects who received placebo [104]. Kenyan children tolerated the same regimen for 12 weeks [105], as did another Indonesian group taking 30 mg every other day for 16–19 weeks [106].

Methemoglobinemia. Primaquine consistently elevates the methemoglobin level (typically <5 g%, and >12 g% is rare) [31]. Fletcher et al. [107] reported elevation to 6.1 g/L (level at baseline, 1.6 g/L) after a regimen of 15 mg daily for 14 days—essentially the same as the level of 5.8 g% observed in Indonesian subjects who received 30 mg daily for 52 weeks [105]. Methemoglobin levels of <20% are tolerated without symptoms or signs [108], and we are not aware of patients receiving primaquine who required treatment for methemoglobinemia. An inborn deficiency of methemoglobin reductase greatly increases the capacity for primaquine and other agents to induce clinically relevant methemoglobinemia [109].

Glucose-6-phosphate dehydrogenase (G6PD) deficiency and primaquine-induced hemolytic anemia. Primaquine causes acute hemolysis in people with inborn G6PD deficiency. Among G6PD-deficient Africans, treatment causes destruction primarily of senescent erythrocytes and is therefore mild and self-

Table 5. Rates of relapse of *Plasmodium vivax* malaria after primaquine therapy at a dosage of 15 mg daily for 5 days.

Reference	Geographic location	Supervised therapy	No. of patients	Duration of follow-up, months	Percentage of patients with relapse
[52]	Central America	Yes	1	9	100
[79]	Pakistan	Yes	5	7–11	100
[23]	Pakistan	No	250	12	51
[22]	India	Yes	725	12	7
[81]	India	No	995	8	10
[82]	India	No	6393	12	1
[83]	India	Yes	62	6	27
[86]	India	No	15,240	12	16
[87]	India	No	5541	12	9

limited, with recovery (with reticulocytosis) even if primaquine therapy is continued [34]. The hemolytic effects in G6PD-deficient subjects were either less severe or altogether absent using either the 45-mg or 60-mg weekly dose for 8 weeks, compared with the 15-mg regimen [34].

Many G6PD variants have been identified [110]. Some are not associated with any hemolytic sensitivity, and others are associated with life-threatening hemolytic episodes, such as the Mediterranean B- variant [111]. Primaquine causes largely unpredictable degrees of severity of hemolysis among patients with other variants, which may occur in any ethnic group. G6PD variants associated with mild primaquine sensitivity, such as African A-, typically show 10%–20% of normal G6PD activity, whereas variants associated with severe sensitivity, such as Mediterranean A-, show less than 5% of the normal activity. Commercially available qualitative tests for G6PD deficiency make diagnosis relatively easy [112, 113]. The inability to rou-

tinely assess G6PD status of a fetus in utero explains why pregnancy is a contraindication for primaquine therapy.

G6PD catalyzes the rate-limiting step in the hexose monophosphate shunt, which drives reduction of glutathione. Diminished G6PD activity thus limits defenses against oxidative damage. However, primaquine-induced hemolysis involves more subtle effects than oxidative attack, because potent stimulation of the hexose monophosphate shunt by primaquine metabolites has been shown to occur independently of glutathione redox equilibrium [111, 114].

CONCLUSIONS

1. The risk of relapse for vivax malaria ranges from 5% to 80%.
2. The standard regimen of 15 mg daily for 14 days is often not effective. A regimen of 30 mg for 14 days should be

Table 6. Summary of reports from India, Pakistan, and Brazil showing the therapeutic efficacy of standard chloroquine in combination with 15 mg of primaquine daily for 5 or 14 days for treatment of *Plasmodium vivax* malaria.

Reference	Geographic location	Primaquine therapy given	No. of patients	Relapse rate, %	Therapeutic efficacy, % (95% CI)
[81]	India	None	222	9	...
[83]	India	None	60	12	...
[84]	India	None	723	9	...
[81]	India	15 mg for 5 days	995	11	-9.7 (-21 to 3)
[83]	India	15 mg for 5 days	62	27	...
[84]	India	15 mg for 5 days	759	7	2.2 (-3 to 7)
[83]	India	15 mg for 14 days	63	0	100 (95 to 100)
[23]	Pakistan	None	350	51	...
[23]	Pakistan	15 mg for 5 days	250	51	0 (-8 to 8)
[23]	Pakistan	15 mg for 14 days	100	32	37 (27 to 48)
[85]	Brazil	15 mg for 5 days	30	27	ND ^a
[85]	Brazil	15 mg for 14 days	31	7	...

^a No data: relapse rate without primaquine therapy unknown.

prescribed, or 0.5 mg/kg daily for 14 days for infants or people weighing >70 kg.

3. Supervised compliance with prescribed therapy is necessary to prove resistance.

4. Taking primaquine with food greatly improves its gastrointestinal tolerability.

5. Primaquine-induced methemoglobinemia is mild and self-limited.

6. Therapeutic doses of primaquine are well tolerated and not toxic in people considered good candidates to receive the treatment.

7. Primaquine is dangerous with G6PD deficiency and should not be used without knowledge of G6PD status or during pregnancy.

8. Standard primaquine therapy rapidly and completely prevents development of *P. vivax* in mosquitoes (i.e., transmission blocking).

9. A single 45-mg dose of primaquine adjunctive to therapy for *P. falciparum* infection may not block transmission.

10. The 5-day, 15-mg regimen of primaquine is not effective against relapse of *P. vivax* malaria.

Acknowledgments

Financial support. This work was supported in part by the US Department of Defense Global Emerging Infections Surveillance Program.

Potential conflict of interest. J.K.B. and S.L.H.: No conflict.

References

- World Health Organization (WHO). Fact sheet no. 94. Geneva: WHO, 1996.
- Zucker JR. 1996. Changing patterns of autochthonous malaria transmission in the United States: a review of recent outbreaks. *Emerging Infect Dis* 1996; 2:37–43.
- Baird JK. Malaria at the millennium: control strategies in crisis. *Drugs* 2000; 59:719–43.
- Centers for Disease Control and Prevention. Local transmission of *Plasmodium vivax* malaria—Houston, Texas, 1994. *MMWR Morb Mortal Wkly Rep* 1995; 44 (15):301–3.
- Layton LM, Parise ME, Campbell CC, et al. Mosquito-transmitted malaria in New York City, 1993. *Lancet* 1995; 346:729–31.
- Brooke JH, Genese CA, Bloland PB, Zucker JR, Spitalny KC. Probable introduced malaria transmission in New Jersey. *N Engl J Med* 1994; 331:22–3.
- Centers for Disease Control and Prevention. Mosquito-transmitted malaria—California and Florida, 1990. *MMWR Morb Mortal Wkly Rep* 1990; 40 (6):106–8.
- Coatney GR, Collins WE, Warren McW, Contacos PG. The primate malarial. Bethesda, MD: US Department of Health, Education and Welfare, National Institutes of Health, 1971.
- Rieckmann KH, McNamara JV, Kass L, Powell RD. Gametocytocidal and sporonticidal effects of primaquine upon two strains of *Plasmodium falciparum*. *Military Med* 1969; 134:802–19.
- Garnham PCC. Hypnozoites and “relapses” in *Plasmodium vivax* and in vivax-like malaria. *Trop Geogr Med* 1988; 40:187–95.
- Most H, London IM, Kane CA, Lavietes PH, Schroeder EF, Hayman JM Jr. Chloroquine for treatment of acute attacks of vivax malaria. *JAMA* 1946; 131:963–7.
- Alving AS, Hankey DD, Coatney GR, et al. Korean vivax malaria. II. Curative treatment with pamaquine and primaquine. *Am J Trop Med Hyg* 1953; 6:970–6.
- Hankey DD, Jones Jr R, Coatney GR, et al. Korean vivax malaria. I. Natural history and response to chloroquine. *Am J Trop Med* 1953; 2: 958–69.
- Coatney GR, Alving AS, Jones R Jr, et al. Korean vivax malaria. V. Cure of the infection by primaquine administered during long-term latency. *Am J Trop Med Hyg* 1953; 6:985–88.
- Adak T, Sharma VP, Orlov VS. Studies on the *Plasmodium vivax* relapse pattern in Delhi, India. *Am J Trop Med Hyg* 1998; 59:175–9.
- Sharma RC, Gautam AS, Orlov V, Sharma VP. Relapse pattern of *Plasmodium vivax* in Kheda District, Gujarat. *Indian J Malariol* 1990; 27:95–9.
- Wiselogle FY. A survey of antimalarial drugs: 1941–1945. 2 vols. Ann Arbor, MI: J.W. Edwards, 1943.
- Edgcomb JH, Arnold J, Yount EH Jr, Alving AS, Eichelberger L. Primaquine, SN 13272: a new curative agent in vivax malaria: a preliminary report. *J National Malaria Soc* 1950; 9:285–92.
- Fairley NH. Chemotherapeutic suppression and prophylaxis in malaria. *Trans Roy Soc Trop Med Hyg* 1945; 38:311–57.
- Most H, London IM, Kane CA, Lavietes PH, Schroeder EF, Hayman JM Jr. Chloroquine for treatment of acute attacks of vivax malaria. *JAMA* 1946; 131:963–7.
- Gordon HH, Dieuaide FR, Marble A, Christianson HB, Dahl LK. Treatment of *Plasmodium vivax* of foreign origin. *Arch Int Med* 1947; 79:365–81.
- Sinha S, Dua VK, Sharma VP. Efficacy of 5 day radical treatment of primaquine in *Plasmodium vivax* cases at BHEL Industrial Complex, Hadwar (U.P.). *Indian J Malariol* 1989; 26:83–6.
- Rowland M, Durrani N. Randomized controlled trials of 5- and 14-days primaquine therapy against relapses of vivax malaria in an Afghan refugee settlement in Pakistan. *Trans Roy Soc Trop Med Hyg* 1999; 93:641–3.
- Luxemburger C, van Vugt M, Jonathan S, et al. Treatment of vivax malaria on the western border of Thailand. *Trans Roy Soc Trop Med Hyg* 1999; 93:433–8.
- Schwartz E, Sidi Y. New aspects of malaria imported from Ethiopia. *Clin Infect Dis* 1998; 26:1089–91.
- Ohtomo H, Hioki A, Tanabe K, Nakabayashi T, Ishizaki T. Clinical evaluation of antimalarial regimens in Japan. *Zbl Bakt Hyg A* 1987; 264:513–20.
- Ehrman FC, Ellis JM, Young MD. *Plasmodium vivax* Chesson strain. *Science* 1945; 101:377.
- Baird JK, Leksana B, Masbar S, et al. Diagnosis of resistance to chloroquine by *Plasmodium vivax*: timing of recurrence and whole blood chloroquine levels. *Am J Trop Med Hyg* 1997; 56:621–6.
- Smoak BL, DeFraités RE, Magill AJ, Kain KC, Welldt BT. *Plasmodium vivax* infections in U.S. Army troops: failure of primaquine to prevent relapse in studies from Somalia. *Am J Trop Med Hyg* 1997; 56:231–4.
- Schwartz E, Parise M, Kozarsky P, Cetron M. Delayed onset of malaria—implications for chemoprophylaxis in travelers. *New Engl J Med* 2003; 349:1501–6.
- Carson PE. 8-Aminoquinolines. In: Peters W, Richards WHG, eds. Antimalarial drugs. II. Current antimalarials and new drug developments. Berlin: Springer-Verlag, 1984:83–121.
- Schmidt LH, Fradkin R, Vaughan D, Rasco J. Radical cure of infections with *Plasmodium cynomolgi*: a function of total 8-aminoquinoline dose. *Am J Trop Med Hyg* 1977 26:1116–28.
- Clyde DF, McCarthy VE. Radical cure of chesson strain vivax malaria in man by 7, not 14, days of treatment with primaquine. *Am J Trop Med Hyg* 1977; 26:562–3.
- Alving AS, Johnson CF, Tarlov AR, Brewer GJ, Kellermeyer RW, Carson PE. Mitigation of the hemolytic effect of primaquine and enhancement of its action against exoerythrocytic forms of the Chesson strain of *Plasmodium vivax* by intermittent regimens of drug administration. *Bull WHO* 1960; 22:621–31.

35. Brueckner RP, Ohrt C, Baird JK, Milhous WK. 8-Aminoquinolines. In: Rosenthal PJ, ed. Antimalarial chemotherapy: mechanisms of action, resistance and new directions in drug discovery. Totowa, NJ: Humana Press, 2000:123–51.
36. Clyde DF. Clinical problems associated with the use of primaquine as a tissue schizonticidal and gametocytocidal drug. Bull WHO 1981; 59:391–5.
37. Palmer RMJ, Weatherall M. The effect of some anti-inflammatory and antimalarial drugs on the migration of horse leukocytes in vitro. Br J Pharmacol 1977; 59:472.
38. Thong YH, Ferrante A, Rowan-Kelly B. Primaquine inhibits mitogen-induced human lymphocyte proliferative responses. Trans Roy Soc Trop Med Hyg 1978; 72:537–9.
39. Thong YH. Immunosuppression caused by primaquine. Trans Roy Soc Trop Med Hyg 1979; 73:474.
40. Thong YH, Ferrante A, Secker LK. Normal immunological responses in mice treated with chloroquine, quinine and primaquine. Trans Roy Soc Trop Med Hyg 1981; 75:108–9.
41. Alving AS, Arnold J, Hockwald RS, et al. Potentiation of the curative action of primaquine in vivax malaria by quinine and chloroquine. J Lab Clin Med 1955; 46:301–6.
42. Baird JK, Basri H, Subianto B, et al. Treatment of chloroquine-resistant *Plasmodium vivax* with chloroquine and primaquine or halofantrine. J Infect Dis 1995; 171:1678–82.
43. Fryauff DJ, Richards AL, Baird JK, et al. Lymphocyte proliferative response and subset profiles during extended periods of chloroquine or primaquine prophylaxis. Antimicrob Agents Chemother 1996; 40: 2737–42.
44. Fryauff DJ, Baird JK, Purnomo, et al. Malaria in a nonimmune population after extended chloroquine or primaquine prophylaxis. Am J Trop Med Hyg 1997; 56:137–40.
45. Kaplan MH, Bernstein LS. Improved therapy for Vietnam acquired vivax malaria. Military Med 1974:444–8.
46. Bunnag D, Karbwang J, Thanavibul A, et al. High dose primaquine in primaquine resistant vivax malaria. Trans Roy Soc Trop Med Hyg 1994; 88:218–9.
47. Pukrittayakamee S, Vanijononta S, Chantra A, Clemens R, White NJ. Blood stage antimalarial efficacy of primaquine in *Plasmodium vivax* malaria. J Infect Dis 1994; 169:932–5.
48. Saint-Yves IFM. Comparison of treatment schedules for *Plasmodium vivax* infections in the Solomon Islands. P N G Med J 1977; 20:62–5.
49. Alving AS, Hankey DD, Coatney R, et al. Korean vivax malaria. II. Curative treatment with pamaquine and primaquine. Am J Trop Med Hyg 1953; 6:970–6.
50. Coatney GR, Alving AS, Jones R Jr, et al. Korean vivax malaria. V. Cure of the infection by primaquine administered during long-term latency. Am J Trop Med Hyg 1953; 6:985–8.
51. Thaeler AD, Arnold JD, Alving AS. A clinical study of primaquine (SN-13272) in the treatment of malaria among the Miskito Indians of Nicaragua. Am J Trop Med Hyg 1953; 2:989–99.
52. Miller LH, Wyler DJ, Glew RH, Collins WE, Contacos PG. Sensitivity of four Central American strains of *Plasmodium vivax* to primaquine. Am J Trop Med Hyg 1974; 23:309–10.
53. Martelo OJ, Smoller M, Saladin TA. Malaria in American soldiers. Arch Intern Med 1969; 123:383–7.
54. Fisher GU, Gordon MP, Lobel HO, Runcik K. Malaria in soldiers returning from Vietnam: epidemiologic, therapeutic and clinical studies. Am J Trop Med Hyg 1970; 19:27–39.
55. Boulos M, Amato Neto V, Dutra AP, Di Santi SM, Shiroma M. Analise da frequencia de recaidas de malaria por *Plasmodium vivax* em regio nao endemica. Rev Inst Med Trop Sao Paulo 1991; 33:143–6.
56. Kass MA, Gordon M, Morley RE, Meltzer DW, Goldberg JJ. Compliance with topical timolol treatment. Am J Ophthalmol 1987; 103: 188–93.
57. Tanabe K, Shimada K. Clinical evaluation of antimalarial drugs [in Japanese]. Kansenshogaku Zasshi 1990; 64:668–73.
58. Roth HP, Caron HS. Accuracy of doctors' estimates and patients' statements on adherence to a drug regimen. Clin Pharmacol Ther 1978; 23:361–70.
59. Jelinek T, Nothdurft HD, Von Sonnenburg F, Loscher T. Long-term efficacy of primaquine in the treatment of vivax malaria in non-immune travelers. Am J Trop Med Hyg 1995; 52:322–4.
60. Ronn AM, Bygbjerg IC. Problemer med primakin- recidivprofylakse hos malariapatienter. Ugeskr Laeger 1993; 155:3901–4.
61. Isaac-Renton JL, Koon ALK, Chan RMT, Chow AW, Sacks SL. Drug resistance in malaria: three cases and a review. Can Med Assoc J 1983; 129:454–7.
62. Henderson A, Simon J, Melia W, Navein J, McCallum J. Polyresistant malaria in Gurka soldiers returning from Papua New Guinea: treatment and prevention. J R Army Med Corps 1986; 132:37–41.
63. Rombo L, Edwards G, Ward SA, et al. Seven patients with relapses of *Plasmodium vivax* or *P. ovale* despite primaquine treatment. Trop Med Parasitol 1987; 38:49–50.
64. Charoenlarp P, Harinasuta T. Relapses of vivax malaria after a conventional course of primaquine and chloroquine: report of 2 cases. Southeast Asian J Trop Med Public Health 1973; 4:135–7.
65. Luzzi GA, Warrell DA, Barnes AJ, Dunbar EM. Treatment of primaquine-resistant *Plasmodium vivax* malaria [letter]. Lancet 1992; 340:310.
66. Lapierre J, Coquelin B, Galal AA, et al. Chimioresistance de souches de *Plasmodium falciparum* et *Plasmodium vivax* au Combodge particulieres morphologiques de *Plasmodium vivax*. Medecine Tropicale 1984; 44:339–49.
67. Arias AE, Corredor RA. Low response of Colombian strains of *Plasmodium vivax* to classical antimalarial therapy. Trop Med Parasitol 1988; 40:21–3.
68. Looareesuwan S, Wilairatana P, Glanarongran R, et al. Atovaquone and proguanil hydrochloride followed by primaquine for treatment of *Plasmodium vivax* in Thailand. Trans Roy Soc Trop Med Hyg 1999; 93:637–40.
69. Duarte EC, Pang LW, Ribeiro LC, Fontes CJ. Association of sub-therapeutic dosages of a standard drug regimen with failures in preventing relapses of vivax malaria. Am J Trop Med Hyg 2001; 65:471–6.
70. Schwartz E, Regev-Yochay G, Kurnik D. Considering primaquine dose adjustment for radical cure of *Plasmodium vivax* malaria. Am J Trop Med Hyg 2000; 62:393–5.
71. Arnold J, Alving AS, Clayman CB, Hochwald RS. Induced primaquine resistance in vivax malaria. Trans Roy Soc Trop Med Hyg 1961; 55: 345–50.
72. Rieckmann KH, Davis DR, Hutton DC. *Plasmodium vivax* resistant to chloroquine? Lancet 1989; 2:1183–4.
73. Baird JK, Basri H, Purnomo, et al. Resistance to chloroquine by *Plasmodium vivax* in Irian Jaya, Indonesia. Am J Trop Med Hyg 1991; 44:547–52.
74. Murphy GS, Basri H, Purnomo, et al. Vivax malaria resistant to treatment and prophylaxis with chloroquine. Lancet 1993; 341:96–100.
75. Phillips EJ, Keystone JS, Kain KC. Failure of combined chloroquine and high-dose primaquine therapy for *Plasmodium vivax* acquired in Guyana, South America. Clin Infect Dis 1996; 23:1171–3.
76. Collins WE, Jeffery GM. Primaquine resistance in *Plasmodium vivax*. Am J Trop Med Hyg 1996; 55:243–9.
77. Singh J, Ray AP, Misra BG, Nair CP. Antirelapse treatment with primaquine and pyrimethamine. Indian J Malariol 1954; 8:127–36.
78. Basavaraj HR. Observations on the treatment of 678 malaria cases with primaquine in an area free from malaria transmission in Mysore State, India. Indian J Malariol 1960; 14:269–81.
79. Contacos PG, Coatney GR, Collins WE, Briesch PE, Jeter MH. Five day primaquine therapy: an evaluation of radical curative activity against vivax malaria infection. Am J Trop Med Hyg 1973; 22:693–5.
80. Cedillos RA, Warren M, Jeffery GM. Field evaluation of primaquine in the control of *Plasmodium vivax*. Am J Trop Med Hyg 1978; 27: 466–72.
81. Roy RG, Shanmugham CAK, Chakrapani KV, Ganeshan AV. Results

- of 5 day course of radical treatment of *Plasmodium vivax* in six districts of Tamil Nadu. Indian J Med Res **1979**; 69:939–43.
82. Singh N, Mishra AK, Sharma VP. Radical treatment of vivax malaria in Madhya Pradesh, India. Indian J Malariol **1990**; 27:55–6.
 83. Gogtay NJ, Desai S, Kamtekar KD, Kadam VS, Dalvi SS, Kshirsagar NA. Efficacies of 5- and 14-day primaquine regimens in the prevention of relapses in *Plasmodium vivax* infections. Ann Trop Med Parasitol **1999**; 93:809–12.
 84. Yadav RS, Ghash SK. Radical curative efficacy of five-day regimen of primaquine for treatment of *Plasmodium vivax* malaria in India. J Parasitol **2002**; 88:1042–4.
 85. Villalobos-Salcedo JM, Tada MS, Kimura E, Menezes MJ, Pereira-da-Silva LH. In vivo sensitivity of *Plasmodium vivax* isolates from Rondonia (western Amazon region, Brazil) to regimens including chloroquine and primaquine. Ann Trop Med Parasitol **2000**; 94:749–58.
 86. Prasad RN, Virk KJ, Sharma VP. Relapse/reinfection patterns of *Plasmodium vivax* infection: a four year study. Southeast Asian J Trop Med Public Health **1991**; 22:499–503.
 87. Dua VK, Sharma VP. *Plasmodium vivax* relapses after 5 days of primaquine treatment in some industrial complexes of India. Ann Trop Med Parasitol **2001**; 95:655–9.
 88. Pukrittayakamee S, Vanijanonta S, Chantra A, Clemens R, White NJ. Blood stage antimalarial efficacy of primaquine in *Plasmodium vivax* malaria. J Infect Dis **1994**; 169:932–5.
 89. Wilairatana P, Silachamroon U, Krudsood S, et al. Efficacy of primaquine regimens for primaquine-resistant *Plasmodium vivax* in Thailand. Am J Trop Med Hyg **1999**; 61:973–7.
 90. Baird JK, Wiady I, Sutanihardja A, et al. Short report: therapeutic efficacy of chloroquine combined with primaquine against *Plasmodium falciparum* in northeastern Papua, Indonesia. Am J Trop Med Hyg **2002**; 66:659–60.
 91. Arnold J, Alving AS, Hockwald RS, et al. The antimalarial action of primaquine against the blood and tissue stages of falciparum malaria. J Lab Clin Med **1955**; 46:391–7.
 92. Rieckmann KH, McNamara JV, Frischer H, Stockert TA, Carson PE, Powell RD. Gametocytocidal and sporonticidal effects of primaquine and of sulfadiazine with pyrimethamine in a chloroquine-resistant strain of *Plasmodium falciparum*. Bull WHO **1968**; 38:625–32.
 93. Rieckmann KH, McNamara JV, Kass L, Powell RD. Gametocytocidal and sporonticidal effects of primaquine upon two strains of *Plasmodium falciparum*. Military Medicine **1969**; 134:802–19.
 94. Hogh B, Gamage-Mendis A, Butcher GA, et al. The differing impact of chloroquine and pyrimethamine/sulfadoxine upon the infectivity of malaria species to the mosquito vector. Am J Trop Med Hyg **1998**; 58:176–82.
 95. Gogtay NJ, Chogle AR, Sorabjee JS, Marathe SN, Kshirsagar NA. Poor gametocytocidal activity of 45 mg primaquine in chloroquine-treated patients with acute, uncomplicated *Plasmodium falciparum* malaria in Mumbai (Bombay): an issue of public health importance. Ann Trop Med Parasitol **1999**; 93:813–6.
 96. Kaneko A, Kamei K, Suzuki T, Ishii A, Siagian R, Panjaitan W. Gametocytocidal effect of primaquine in a chemotherapeutic malaria control trial in North Sumatra, Indonesia. Southeast Asian J Trop Med Public Health **1989**; 20:351–9.
 97. Klein TA, Tada MS, Lima JBP, Tang AT. Infection of *Anopheles darlingi* fed on patients infected with *Plasmodium vivax* before and during treatment with chloroquine plus primaquine in Costa Marques, Rondonia, Brazil. Mem Inst Oswaldo Cruz **1992**; 87:191–5.
 98. Baird JK, Purnomo, Masbar S. *Plasmodium ovale* in Indonesia. Southeast Asian J Trop Med Pub Health **1990**; 21:541–4.
 99. Patterson JE, Bia FJ, Miller K, McPhedran P. Relapsing malaria infection acquired in Kenya. Yale J Biol Med **1987**; 60:245–53.
 100. Purnomo, Solihin A, Gomez-Saladin E, Bangs MJ. Rare quadruple malaria infection in Irian Jaya Indonesia. J Parasitol **1999**; 85:574–9.
 101. Nathwani D, Currie PF, Smith CC, Khaund R. Recurrent *Plasmodium ovale* infection from New Guinea—chloroquine resistance or inadequate primaquine therapy. J Infect **1991**; 23:343–4.
 102. Brewer GJ, Tarlov AR, Alving AS. The toxicity of the 8-aminoquinoline antimalarial drugs. Bull Nat Soc Ind Mal Mosq Dis **1961**; 9:331.
 103. Clayman CB, Arnold J, Hochwald RS, Yount EH, Edgcomb JH, Alving AS. Toxicity of primaquine in Caucasians. JAMA **1952**; 149:1563–8.
 104. Fryauff DJ, Baird JK, Basri H, et al. Randomized placebo-controlled trial of primaquine for prophylaxis of falciparum and vivax malaria. Lancet **1995**; 346:1190–3.
 105. Weiss WR, Johnson A, Oloo AJ, Hoffman SL. Daily primaquine is an effective prophylaxis against falciparum malaria in Kenya. J Infect Dis **1995**; 171:1569–75.
 106. Baird JK, Fryauff DJ, Basri H, et al. Primaquine for prophylaxis against malaria among nonimmune transmigrants in Irian Jaya, Indonesia. Am J Trop Med Hygiene **1995**; 52:479–84.
 107. Fletcher KA, Evans-Price DA, Giles HM, et al. Studies on the pharmacokinetics of primaquine. Bull WHO **1981**; 59:407.
 108. Coleman MD, Coleman NA. Drug-induced methaemoglobinaemia: treatment issues. Drug Safety **1996**; 14:394.
 109. Hall AH, Kulig KW, Rumack BH. Drug- and chemical-induced methaemoglobinaemia: clinical features and management. Medical Toxicol **1986**; 81:579–82.
 110. Beutler E. G6PD deficiency. Blood **1994**; 84:3613–36.
 111. Baird JK, Davidson DE, Decker-Jackson JE. Oxidative activity of hydroxylated primaquine analogs: non-toxicity to glucose-6-phosphate dehydrogenase-deficient human red blood cells in vitro. Biochemical Pharmacology **1986**; 35:1091–8.
 112. Echler G. Determination of glucose-6-phosphate dehydrogenase. Am J Med Technol **1983**; 49:259.
 113. Tantular IS, Iwai K, Lin K, et al. Field trials of a rapid test for G6PD deficiency in combination with a rapid diagnosis of malaria. Trop Med Int Health **1999**; 4:245–50.
 114. Baird JK, McCormick GJ, Canfield CJ. Effects of nine synthetic putative metabolites of primaquine on activity of the hexose monophosphate shunt in human red blood cells in vitro. Biochem Pharmacol **1986**; 35:1099–106.