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Effect of dietary crude protein source on hormone and follicle characteristics in beef heifers

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ABSTRACT: Ground, raw soybeans (SB), or dried distillers grain plus solubles (DDGS) were utilized in heifer development diets to determine the effect of dietary fat and protein source on hormone and follicle characteristics and ADG. The experiment was conducted over 2 yr with 100 June-born heifers (199 ± 2 kg initial BW, n = 50 per yr). The experimental periods were 157 and 207 d in yr 1 and 2, respectively. Heifers were provided a dietary supplement (DM basis) of 1.23 kg of SB and 0.40 kg of corn or 1.65 kg of DDGS between weaning and breeding. Estrus was synchronized with 2 injections of PGF2α 14 d apart. Dominant follicles were measured and aspirated via transvaginal ultrasonography 60 h after the second PGF2α injection. Heifers were exposed to bulls beginning 14 d after aspiration for 45 d. Heifer ADG was greater (P = 0.02) for DDGS heifers in yr 1, but was similar (P = 0.47) in yr 2. However, there was no difference (P = 0.35) in final BW in either year. There was no difference (P ≥ 0.67) in follicle size, follicle hormone concentrations, or pregnancy rate (88%) between yr 1 and 2. Serum estrogen at 48 or 60 h after PGF2α injection were similar (P ≥ 0.91); however, LH at 60 h in yr 2 tended to be greater (P = 0.07) for DDGS heifers. The percentage of heifers experiencing an LH surge 48 and 60 h after PGF2α injection was not affected (P ≥ 0.40) by treatment. Calf production was not affected (P ≥ 0.20) by developmental diet. In summary, DDGS and SB have similar effects on hormone and follicle characteristics at the inclusion rates used in these studies.

Key words: fertility, heifer development, protein supplement

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doi:10.2527/jas.2009-2236

INTRODUCTION

Development of replacement heifers at growth rates sufficient to ensure that a large percentage reach or are near puberty before breeding is important for efficient beef cattle production. Supplementation of low- or medium-quality roughage diets with high-energy sources of protein can provide sufficient nutrients for economic development and successful reproductive function. Soybeans (SB) are a nutrient-dense feedstuff (40% CP; 20% fat, DM basis; NRC, 1996), associated with improved reproductive function in beef heifers (Lammoglia et al., 2000) and mature cows (Wehrman et al., 1991; Ryan et al., 1992). Another economical, high-energy source of CP from the corn milling industry is dried distillers grain plus solubles (DDGS; 30% CP; 11% fat, DM basis; NRC, 1996). Feeding DDGS provides CP, undegradable intake protein, energy, and fat and has been reported to improve conception rates in yearling beef heifers (Martin et al., 2007). Both SB and DDGS may be beneficial to reproduction as sources of dietary fat and CP (Wiley et al., 1991; Williams and Stanko, 2000; Martin et al., 2007). Supplemental fat stimulated programmed growth of preovulatory follicles, increased total number of follicles, and increased the size of preovulatory follicles (Mattos et al., 2000). Increased size of preovulatory follicles may be due, in part, to increased concentrations of plasma LH, which stimulates the later stage of follicular growth. The ovulation of larger follicles may result in the formation of larger corpora lutea with increased steroidogenic capacity and result in greater progesterone production, which has been associated with greater conception rates. Thus, to further investigate the mechanism by which these supplements may influence reproductive outcome, the objective of this study was to determine the effects of supplemental SB or DDGS on hormone and follicle characteristics and BW gain in beef heifers.

1A contribution of the University of Nebraska Agricultural Research Division, supported in part by funds provided through the Hatch Act. Mention of a trade name, proprietary products, or company name is for presentation clarity and does not imply endorsement by the authors of the University of Nebraska.
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Received June 22, 2009.
Accepted November 13, 2009.
MATERIALS AND METHODS

Procedures and facilities for this experiment were approved by the University of Nebraska–Lincoln Institutional Animal Care and Use Committee. This experiment was conducted over 2 yr at the University of Nebraska Gudmunsden Sandhills Laboratory near Whitman.

This experiment was replicated in 2 yr using 100 summer-born heifers (n = 50 each yr). Crossbred heifers (Red Angus × Simmental; 199 ± 2 kg initial BW; 8 mo of age) were allotted by BW and randomly assigned to 1 of 4 pens per year. All heifers had ad libitum access to late-harvested Sandhills meadow hay (CP = 6.2 and 8.8% for yr 1 and 2, respectively). In each year, 2 pens of heifers were supplemented daily with 1.23 kg of ground SB mixed with 0.40 kg of ground corn (31% CP, 17.1% fat, DM basis) per heifer and 2 pens of heifers were supplemented with 1.65 kg of DDGS (31% CP, 11.5% fat, DM basis; Chief Ethanol Fuels, Hastings, NE) per heifer daily. Supplements were approximately isocaloric. Supplement nutrient analysis was performed by standard wet chemistry procedures (Ward Laboratories, Kearney, NE). The experimental supplementation periods for yr 1 and 2 were 157 and 207 d, respectively, continuing through follicle aspiration.

Blood samples (5 mL via coccygeal venipuncture) were collected at 10-d intervals before and during the feeding period to determine pubertal status. Blood samples were stored at 4°C for serum separation by centrifugation (1,300 × g for 20 min at 4°C) within 24 h. Serum samples were stored at −20°C for subsequent analysis. Serum progesterone concentrations of >1 ng/mL were considered indicative of luteal activity. Body weights were measured at the time of blood collection and at the end of the feeding period. Estrus was synchronized with 2 injections of PGF2α, 14 d apart. Sixty hours after the second injection, the largest (dominant) follicles (DF) were measured and aspirated using an ultrasound-guided transvaginal probe (Aloka, UST-981-5). After positioning the ultrasound-guided probe (Aloka 500V ultrasound with a 5.0-MHz probe; Aloka, Wallingford, CT) with needle attachment, Blood samples (5 mL via coccygeal venipuncture) were taken 48 and 60 h after the second injection of PGF2α for serum 17β-estradiol (E2) and LH analysis. Blood samples were stored at 4°C for serum separation by centrifugation (1,300 × g for 20 min at 4°C) within 24 h. Granulosa cells were harvested from aspirates, and follicular fluid (FF) samples were stored at −80°C until subsequent steroid hormone analysis. Heifers were managed on native pastures with bulls stored at −80°C until subsequent steroid hormone analysis. Heifers were managed on native pastures with bulls stored at −80°C until subsequent steroid hormone analysis.

Concentrations of progesterone in serum and FF were determined using a solid-phase RIA kit (Coat-A-Count, Diagnostic Products Corporation, Los Angeles, CA) without extraction as described previously (Stewart et al., 1996; Melvin et al., 1999). Inter- and intraassay CV for serum progesterone were 6.5 and 6.4%, respectively. Inter- and intra-assay CV for FF progesterone were 9.0 and 9.1%, respectively. Concentrations of E2 in extracted serum samples and nonextracted FF samples were determined by RIA as described by Kojima et al. (1992). Inter- and intraassay CV for FF E2 were 4.1 and 4.6% respectively; assay sensitivity was 0.01 pg/mL. Inter- and intraassay CV for serum E2 were 10.0 and 10.0% respectively; assay sensitivity was 0.02 pg/mL. Preovulatory follicles producing more E2 than progesterone were classified as estrogen-active. As granulosa cells luteinize shortly before ovulation, steroid production shifts from E2 to progesterone and the follicle is no longer E2-active. Estrogen-activity of the DF was determined by the estrogen:progesterone ratio and E2 concentrations (ratio ≥1.0 and >100 ng of E2/mL of FF indicates DF is estrogen-active; Roberts and Echternkamp, 2003). Concentrations of LH in serum were determined in all samples by double-antibody RIA (Wolfe et al., 1989; Cupp et al., 1995) using an antisemum made against ovine LH in rabbits (TEA-RaOLH #35), purified iodinated ovine LH (LER-1056-C2) as labeled hormone, and NIH-LH-B7 as the standard. The assay for LH was validated as follows. Serial dilutions of 4 independent bovine serum samples were assayed at volumes ranging from 50 to 200 µL. Assay determinations of these dilutions from each of the 4 independent samples were highly correlated (r = 0.9997). Three independent samples were used to determine the recovery of added LH (15.6, 31.3, 62.5, 125, and 250 pg). Percent recovery from these 3 samples averaged 100.2 ± 5.7%. Intra- and interassay CV were 2.6 and 8.2%, respectively. The limit of detection of LH was 0.088 ng/mL of serum under the conditions utilized. Serum LH concentrations greater than 4 ng/mL at the 48- or 60-h sampling were used as criteria to indicate a preovulatory LH surge had occurred (Cupp et al., 1995).

Follicle Aspiration

Before follicle aspiration, both ovaries were visualized via ultrasonography (Aloka SSD-500V) to determine location and size of the DF. The size of the largest follicle was measured across 2 dimensions and recorded. Follicle aspirations were performed using an 18-gauge stainless-steel needle housed within a transvaginal ultrasound-guided probe (Aloka, UST-981-5). After positioning the DF opposite the vaginal wall from the needle guide, the needle was inserted through the vaginal wall and into the follicle. Follicular fluid and granulosa cells were aspirated by vacuum pressure created by a 60-mL disposable syringe connected by plastic surgical tubing to the needle. A sterile syringe was used for each animal, and the needle and tubing were rinsed 3 times with sterile water between each aspiration. Immediately after collection, follicular aspirates were transferred into 1.5-mL
conical tubes and placed on ice until centrifugation at 14,000 × g for 1 min at 4°C.

**Statistical Analyses**

Data were analyzed using pen as the experimental unit. Data were tested for year × treatment interactions ($P \leq 0.05$). Individual year data will be presented where an interaction exists. Differences in BW, ADG, timing of estrus, DF diameter, and hormone concentrations in serum and FF were analyzed using PROC MIXED (SAS Inst. Inc., Cary, NC). Differences in synchronization, conception and pregnancy rates, cyclicity, and number of E2-active follicles were analyzed using PROC GLIMMIX of SAS. Nonsignificant ($P > 0.10$) covariates, including age and sire breed, were removed from the statistical model, and data were re-analyzed with only treatment and significant covariates remaining in the model. The random effect of pen within year × treatment was included in the model.

**RESULTS**

Midtest BW tended to be greater ($P = 0.08$) for DDGS than SB heifers, but final BW was not different ($P = 0.35$; Table 1). Year of experiment (yr 1 vs. 2) had a significant ($P = 0.03$) effect on ADG. In yr 1, DDGS heifers gained more BW ($P = 0.02$) than SB heifers. However, ADG was not different ($P = 0.47$) in yr 2. Treatment did not affect estrous cyclicity, nor did year ($P > 0.10$). Progesterone analysis of serum indicated 28% of heifers had achieved puberty at treatment initiation ($P = 0.54$), and 94% were pubertal at the end of the feeding period ($P = 0.99$).

There was no difference ($P = 0.69$) in DF diameter between DDGS- and SB-fed heifers (Table 2). Follicular fluid concentration of E2 ($P = 0.87$) and progesterone ($P = 0.96$) were not affected by treatment. There was no difference ($P = 0.67$) in percentage of E2-active follicles between treatment groups. There were also no differences ($P > 0.10$) in FF E2 or progesterone concentrations when only E2-active follicle data were analyzed. Concentrations of E2 and LH in serum collected 48 h after the second injection of PGF2α, were not affected ($P \geq 0.49$) by treatment (Table 3). There was a treatment × year interaction ($P = 0.04$) for LH concentrations at 60 h, where SB and DDGS heifers had similar ($P = 0.35$) serum LH concentrations in yr 1 (5.83 vs. 3.07 ± 1.83 ng/mL; SB and DDGS, respectively). However, SB heifers tended to have less ($P = 0.07$) serum LH concentrations at 60 h in yr 2 (2.18 vs. 8.55 ± 1.83 ng/mL; SB and DDGS, respectively) than DDGS heifers. Serum E2 concentration at 60 h after PGF2α was unaffected ($P = 0.91$) by treatment. The percentage of heifers experiencing an LH surge at 48 h ($P = 0.76$) and 60 h ($P = 0.40$) was not different between treatment groups (Figure 1). Pregnancy rates were not affected ($P = 0.99$) by dietary treatment (88%; Table 2).

Calving data are displayed in Table 4. Neither calf birth date ($P = 0.52$) nor the percentage of heifers giving birth in the first 21 d of the season ($P = 0.84$) was affected by development diet. Percentage of male calves ($P = 0.54$) and calf birth BW ($P = 0.77$) was similar between dietary treatments. The percentage of heifers requiring assistance at calving was not different ($P = 0.20$). Calf weaning BW ($P = 0.22$) and adjusted 205-d calf BW ($P = 0.24$) were also similar between DDGS and SB heifers. After the second breeding season, 95% of SB-developed and 91% of DDGS-developed heifers were pregnant ($P = 0.58$).

**DISCUSSION**

In the current study, heifer development supplementation strategy did not influence DF diameter after estrous synchronization. Similar proportions of heifers experienced an LH surge before aspiration with similar FF E2 and progesterone concentrations. Soybeans are...
a legume and contain phytoestrogens, which have been associated with endocrine disruption in ruminant animals (Adams, 1995). The limited research conducted with SB in heifer development diets suggests an estrogenic feedstuff may affect synchronization rate and the timing of estrus after PGF2α in beef heifers (Funston, 2004). However, SB feeding has no detrimental effects on pregnancy rate (Howlett et al., 2003; Funston, 2004; Harris et al., 2008). In the current study, overall pregnancy rates were not affected by supplement.

The high fat content of SB limits their use in cattle diets to 10 to 15% of total ration DM. Soybeans have elevated linoleic acid content, which has been associated with improved reproductive function in beef heifers (Lammoglia et al., 2000) and mature cows (Wehrman et al., 1991; Ryan et al., 1992). Previous research has demonstrated feeding increased levels of dietary fat results in a larger DF (Lammoglia et al., 1996; De Fries et al., 1998) and is associated with reduced embryonic mortality (Funston, 2004). In the current study, overall pregnancy rates were not affected by supplement.

Table 2. Effects of supplement on follicle data and pregnancy rates

<table>
<thead>
<tr>
<th>Item</th>
<th>Treatment</th>
<th>SB</th>
<th>DDGS</th>
<th>SEM</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td></td>
<td>50</td>
<td>50</td>
<td>0.4</td>
<td>0.69</td>
</tr>
<tr>
<td>DF diameter, mm</td>
<td></td>
<td>12.4</td>
<td>12.2</td>
<td>0.4</td>
<td>0.69</td>
</tr>
<tr>
<td>FF E2, pg/mL</td>
<td></td>
<td>1,831.6</td>
<td>1,924.3</td>
<td>369.4</td>
<td>0.87</td>
</tr>
<tr>
<td>FF P4, ng/mL</td>
<td></td>
<td>118.1</td>
<td>116.8</td>
<td>20.1</td>
<td>0.96</td>
</tr>
<tr>
<td>E2-active, %</td>
<td></td>
<td>76</td>
<td>70</td>
<td>7</td>
<td>0.67</td>
</tr>
<tr>
<td>Pregnant, %</td>
<td></td>
<td>88</td>
<td>88</td>
<td>5</td>
<td>0.99</td>
</tr>
</tbody>
</table>

1SB = supplemented with 1.23 kg of raw, ground soybeans and 0.40 kg of corn during development; DDGS = supplemented with 1.65 kg of dried distillers grain plus solubles during postweaning development.
2Dominant follicle (DF) diameter.
3Follicular fluid (FF) estradiol (E2) concentrations (pg/mL) for dominant follicle.
4FF progesterone (P4) concentrations (ng/mL) for dominant follicle.
5Percent of dominant follicles with E2:P4 ratio >1.0 and E2/mL of FF >100 ng.

Table 3. Serum LH and estradiol (E2) concentrations taken at 48 and 60 h after second PGF2α injection

<table>
<thead>
<tr>
<th>Item</th>
<th>Treatment</th>
<th>SB</th>
<th>DDGS</th>
<th>SEM</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>LH, ng/mL</td>
<td></td>
<td>2.85</td>
<td>2.22</td>
<td>0.61</td>
<td>0.49</td>
</tr>
<tr>
<td>48 h^2</td>
<td></td>
<td>5.83</td>
<td>3.07</td>
<td>1.83</td>
<td>0.35</td>
</tr>
<tr>
<td>60 h^3</td>
<td></td>
<td>2.18</td>
<td>8.55</td>
<td>1.83</td>
<td>0.07</td>
</tr>
<tr>
<td>Yr 1</td>
<td></td>
<td>15.97</td>
<td>15.88</td>
<td>1.57</td>
<td>0.97</td>
</tr>
<tr>
<td>Yr 2</td>
<td></td>
<td>16.56</td>
<td>16.32</td>
<td>1.50</td>
<td>0.91</td>
</tr>
</tbody>
</table>

1SB = supplemented with 1.23 kg of raw, ground soybeans and 0.40 kg of corn during postweaning development; DDGS = supplemented with 1.65 kg of dried distillers grain plus solubles during postweaning development.
2Assessed in serum harvested 48 h after PGF2α injection.
3Assessed in serum harvested 60 h after PGF2α injection.

Figure 1. Percentage of heifers experiencing an LH surge at 48 and 60 h after PGF2α injection. No difference (P > 0.40) was detected between heifers offered a raw, ground soybean (SB), or dried distillers grain plus solubles (DDGS) supplement.
collection of reproductive data was delayed by follicle aspiration.

Previous data indicate spring-born heifers fed diets containing SB, after a large percentage attained puberty, exhibited decreased response to synchronization and delayed estrus (Harris et al., 2008). If spring-born heifers consumed diets containing SB and heifers were prepubertal, no difference was seen in reproductive responses. Conception and pregnancy to AI and final pregnancy rates were similar across diets regardless of when SB diets were initiated. In the current experiment, DF characteristics were similar in summer-born heifers supplemented with SB or DDGS during postweaning development, and pregnancy rates did not differ throughout the 2-yr experiment. These data, and previous work, indicate SB and DDGS provided at these levels of supplementation are acceptable feedstuffs for heifers in the prebreeding period.

LITERATURE CITED


hormone and IGF-I receptors, and steroids in dominant follicles during the first follicular wave in cattle exhibiting regular estrous cycles. Endocrinology 137:2842–2850.


