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Influence of heterocyclic and oxime-containing farnesol analogs on quorum sensing and pathogenicity in *Candida albicans*

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Abstract

A series of synthetic molecules combining a geranyl backbone with a heterocyclic or oxime head group are quorum-sensing molecules that block the yeast to mycelium transition in the dimorphic fungus *Candida albicans*. A number of the analogs have an $IC_{50} \leq 10 \mu M$, a level of potency essentially identical to the natural quorum sensing signal, the sesquiterpene farnesol. Two of the most potent analogs, neither toxic toward healthy mice, display remarkably different effects when co-administered with *C. albicans*. While neither offers protection from candidiasis, one analog mimics farnesol in acting as a virulence factor, whereas the other has no effect. The results offer the first example of highly potent synthetic fungal quorum-sensing molecules, and provide the first evidence for the ability to decouple quorum sensing and virulence.

Graphical abstract

A series of farnesol analogs are potent quorum-sensing molecules for the human pathogen *Candida albicans*. Two of the most potent analogs display very different effects in a mammalian model of systemic candidiasis.

Keywords: Farnesol, Quorum sensing, *Candida albicans*, Virulence, Candidiasis, Oxime, Tetrazole, Triazole, Thiotetrazole, Thiotriazole, Thioimidazole

1. Introduction

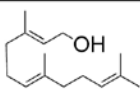
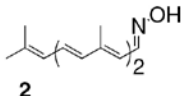
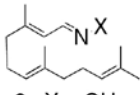
Candida albicans normally resides as a harmless yeast in the GI and urinary tracts, and to a lesser extent on the skin, of most humans. However, for patients with weakened immune systems (e.g., AIDS patients, transplant recipients, and premature infants) *C. albicans* is an opportunistic and often deadly pathogen that will invade host tissues, undergo a dimorphic shift, and then grow as a fungal mass in the kidney, heart, or brain. *Candida albicans* is the fourth leading cause of hospital-acquired infection in the United States and over 95% of AIDS patients will suffer from infections by *C. albicans*.^[1] and [2] There is a pressing need for new treatments for fungal infections.

Candida albicans is a polymorphic fungus able to grow as yeasts, hyphae, and pseudohyphae, and a great many chemical and environmental factors influence the relative populations of these morphologies. In liquid culture, *C. albicans* develops as either yeasts or hyphae, depending in part on whether the cell densities are above or below 10^6 cells per ml.³ Our research revealed that the population dependence of *C. albicans* morphology results from the continuous production and detection

of the sesquiterpene farnesol (**1**, Table 1),^[3] and [4] which acts to block the yeast to mycelium transition at concentrations as low as 1–2 μM .^[5] and [6] In accordance with the precedent established for homoserine lactones in Gram-negative bacteria,⁷ the phenomenon was termed quorum sensing. Farnesol was the first quorum-sensing molecule (QSM) to be discovered in an eukaryotic system.^[3] and [4]

The dimorphic nature of *C. albicans* is essential for pathogenicity.⁸ The discovery of a QSM able to direct growth in the yeast phase therefore suggested the possibility of a “single morphology therapy” which would force this opportunistic pathogen to remain in a monomorphic, nonpathogenic lifestyle. However, three separate experiments have shown that farnesol actually increases fungal pathogenicity. First, cultures of *C. albicans* pretreated with subinhibitory doses of fluconazole, a protocol that increases the levels of secreted farnesol 8- to 10-fold,⁹ were 4- to 5-fold more pathogenic to mice.¹⁰ Second, mice which received farnesol either in their drinking water or via intraperitoneal injection showed ca. 4-fold higher lethality from candidiasis than mice receiving control IP injections.¹¹ Third, a knockout mutant of *C. albicans* lacking the

Table 1. QSM activity of oximes

Analog	IC ₅₀ (μM) ^a	Toxicity ^b
 1 (farnesol)	1–2	None
 2	10	ND ^c
 3: X = OH 4: X = OMe	3: 5 4: ≥ 100	3: High ^d 4: ND

^a Inhibition of germ tube formation.^b Healthy mice observed over 14 d following injection (IP) with 1 ml of 20 mM solution (0.5% Tween 80 v/v in sterile, nonpyrogenic saline).^c Not determined.^d Mice died within 4–5 days.

DPP3 gene encoding an isoprenoid pyrophosphatase for conversion of farnesyl pyrophosphate to farnesol, and thus producing only 15% as much farnesol as the wild-type, was 4- to 5-fold less pathogenic. Reconstitution of *DPP3* restored both farnesol production and pathogenicity.¹¹ Taken together, these observations show that farnesol can act as a virulence factor for *C. albicans*. We have subsequently discovered a potential basis for this behavior, in that farnesol interferes with the normal progression of cytokine induction in mice infected with *C. albicans*.¹²

We are interested in the relationships between quorum sensing and virulence and, in particular, whether it is possible to decouple the two. Specifically, we were interested in the possibility of designing analogs that retain farnesol's ability to block hyphal development but lack farnesol's activity as a virulence factor. In order to explore this problem, we required potent QSMs structurally distinct from farnesol. We now report the discovery of a series of highly potent synthetic quorum-sensing molecules, as well as preliminary investigations of their activity in a mouse model of systemic candidiasis.

2. Results

When we initiated these studies, there were no synthetic molecules possessing a significant fraction of farnesol's biological activity as a quorum-sensing molecule. Previously, we had probed farnesol's mode of action with over 40 natural and synthetic farnesol analogs.⁶ Modified structural features included the head group, chain length, presence or absence of the three double bonds, substitution of a backbone carbon by S, O, N, and Se heteroatoms, presence or absence of a 3-methyl branch, and bulkiness of the hydrophobic tail. Although, more than half the compounds showed measurable quorum-sensing activity (inhibition of hyphal development), the best displayed only 7% of the activity of *E,E*-farnesol.⁶

2.1. Oxime leads for new QSMs

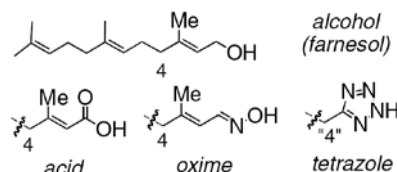
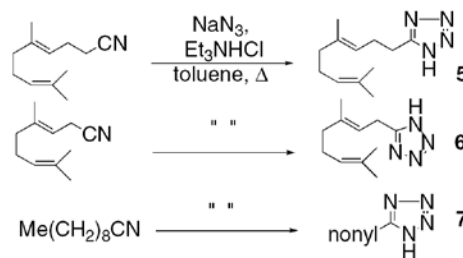
While developing a series of fluorescent farnesol analogs,¹³ we discovered a pentaene oxime **2** with QSM activity (IC₅₀ = 10 μM) superior to any previously investigated analog (Table 1; see the experimental section for a description of the QSM assay). Based on this discovery, we prepared the oxime of farnesaldehyde (**3**) and found it was also a potent QSM (IC₅₀ = 5 μM, Table 1). The corresponding *O*-methyl ether, farnesaldehyde methoxyoxime **4**, was inactive (IC₅₀ > 100 μM), consistent with the results of earlier studies pointing to a requirement for an acidic head group.⁶ However, oxime **3** was found to exhibit significant toxicity (IP) in healthy mice and this class of molecules was not pursued further.

2.2. Tetrazole QSMs

The high quorum-sensing activity of oximes **2** and **3** led us to explore replacement of the primary alcohol of farnesol with other acidic functional groups (Fig. 1). Our earlier work had found farnesoic acid to be a weakly active QSM (IC₅₀ = 36 μM).⁶ 5-Substituted-1H-tetrazoles are weak acids (pK_a ~ 5) often used as lipophilic isosteres for carboxylic acids,^{[14] and [15]} and therefore we investigated several analogs incorporating a tetrazole in place of C₁–C₃ of farnesol.

Analog **5**, **6**, and **7** were synthesized via cycloaddition of the appropriate nitrile with Et₃NHN₃, generated in situ from NaN₃ (Scheme 1).¹⁶

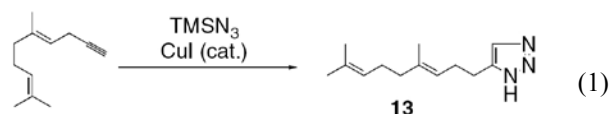
Analog **5**, which incorporates a tetrazole in place of the C₁–C₃ region of farnesol, was a potent QSM with IC₅₀ = 7 μM (Table 2). However, in agreement with conclusions from our earlier research,⁶ chain length was clearly important. Tetrazole **6**, although possessing only one fewer backbone carbon than **5**, was considerably less active, while commercially available 5-methyl-1-tetrazole (not shown) was inactive as a QSM at concentrations up to 3 mM. Nonyl tetrazole **7** and commercially

**Figure 1.** Design concept for analogs.**Scheme 1.** Synthesis of Tetrazoles.

available styrenyl tetrazole **8** were also highly active; however, **7** was only marginally soluble in water. The strong quorum-sensing activity observed for analogs **5** and **8** implies a functional equivalency between styrene and 4,8-dimethyl-3,7-nonadiene sidechains, suggesting that 3-methyl-5-phenyl-2,4-pentadienol or similar structures might represent productive future targets as synthetic QSMs.¹⁷ Along these lines, it is interesting to note a report indicating that dodecanol, an analog of the 12-carbon homoserine lactone secreted as a QSM by *Pseudomonas aeruginosa*, is a weak QSM for *C. albicans*.¹⁸

2.3. Other heterocyclic QSMs

The potency of the tetrazoles as quorum-sensing molecules led us to investigate additional examples of five-membered heterocyclic analogs possessing the same approximate shape, crosssection, lipophilicity, and molecular length (Scheme 2). Our previous studies had found the replacement of a methylene with a sulfur atom (thioether) to be a well-tolerated structural modification,⁶ and we therefore mainly focused on sulfur-linked heterocycles, which are readily available via nucleophilic displacement (Scheme 2).¹⁹ A similar approach has been employed for synthesis of a thiotriazole from farnesyl chloride.²⁰ The substrates tested grafted a series of heterocyclic head groups onto a geranyl (3,7-dimethyl-2,6-octadienyl) tail: thio-1,2,3,4-tetrazole (**9**), thio-1,2,4-triazole (**10**), thio-1,2,3-triazole (**11**), and thioimidazole (**12**). We also investigated a carbon-linked 1,2,3-triazole (**13**) prepared through a dipolar cycloaddition (Eq. 1).²¹



Although thiotetrazole **9** and thioimidazole **12** were only minimally active ($IC_{50} > 50 \mu M$), triazoles **10**, **11**, and **13** were potent QSMs (Table 3). It is notable that two of the analogs (**10** and **13**) exhibit QSM activity equal to *E,E*-farnesol, with three other analogs (**5**, **7**, and **11**) displaying activities nearly as great. It is important to note that none of the compounds listed in Table 1 and Table 2 had any inhibitory effects on the rate of cell growth, even at the highest concentrations tested ($\geq 100 \mu M$).

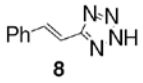
2.4. Effect of heterocyclic QSMs on candidiasis

In selecting analogs for mouse assays, we eliminated the nonyl tetrazole **7** due to limited water solubility. Three of the most potent synthetic QSMs (**5**, **10**, and **11**) were tested next for their potential toxicity to mice following IP injection of 1 ml of a 20 mM solution (ca. 4.4 mg of analog in 0.5% v/v Tween 80 in sterile, nonpyrogenic saline). Attempts to use 0.1% Triton X-100 in place of Tween 80 were precluded by toxicity. In an initial experiment with 3 mice per group, no toxicity was observed for at least 14 days following injection. In a follow-up experiment with 15 mice per group, also observed for at least 14 days, injection of **11** caused a slight peri-

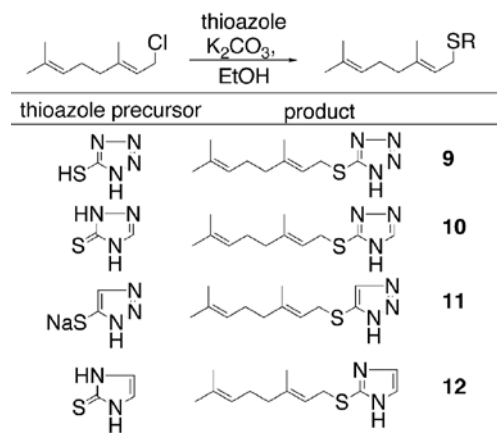
tonitis and mild hepatomegaly in some mice. Therefore, analogs **5** and **10** were chosen for further testing with a standard mouse model of candidiasis,^{[10] and [11]} employing tail vein injection of *C. albicans* cells and IP injection of the analogs.

Forty-five mice were inoculated with 5×10^6 cells of *C. albicans* A72 through tail vein injection; an additional 45 mice were inoculated with 2×10^6 cells. Each batch of mice was divided into 3 groups of 15 mice each. The chosen doses provide ca. 50% mortality after 3 days with strain *C. albicans* A72;¹⁰ virtually identical results were obtained with each dose. Group 1 was given a control injection (IP, 1 ml of 0.5% Tween 80 in saline) while groups 2 and 3 were injected with

Table 2. Quorum sensing and toxicity of tetrazoles

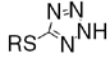
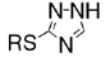
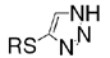
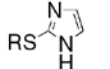
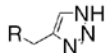
Tetrazole	IC_{50} (μM) ^a	Toxicity ^b
5	7	Non-toxic, virulence factor
6	>50	ND
7	3	Non-toxic
	10	ND

^{a,b} See Table 1.



Scheme 2. Additional heterocyclic analogs.

Table 3. Quorum sensing and toxicity in **9–13**

	Azole ^c	IC_{50} (μM) ^a	Toxicity ^b
9		>50	ND
10		1	None in ≥ 14 d; not a virulence factor
11		3	Slight
12		>50	ND
13		1–2	ND

^{a,b} See Table 1; ^c R = geranyl.

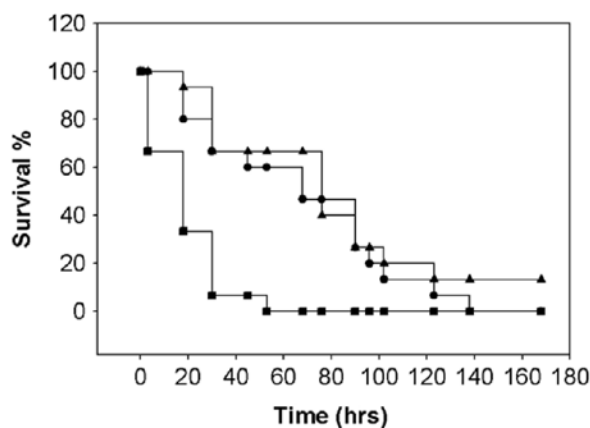


Figure 2. Influence of **5** and **10** on systemic candidiasis. Mouse mortality in the presence and absence of tetrazole **5** and triazole **10** following inoculation with *C. albicans* A72 (5×10^6 cells) and IP injection of 1 ml of: • (saline); ▲ (20 mM thio-1,2,4-triazole **10** in 0.5% Tween 80); ■ (20 mM tetrazole **5** in 0.5% Tween 80).

analogs **5** and **10**, respectively. Neither analog protected the mice from candidiasis (Fig. 2). In the case of the thiotriazole **10**, mouse lethality closely paralleled that observed for the control Tween 80 injections. In contrast, coadministration of tetrazole **5** enhanced the pathogenicity of *C. albicans* A72 ca. 4- to 5-fold, almost exactly duplicating the effects of farnesol.^{[10] and [11]} The effect is sufficiently pronounced that the entire group of 15 mice treated with **5** died within 60 h of infection. It is noteworthy that **5** and **10** display opposing effects in vitro (QSM) and in vivo (pathogenicity). Thiotriazole **10** is ca. 7-fold more potent than **5** as a QSM but, unlike the tetrazole, it does not act as a farnesol-mimic in promoting virulence. Thus, the two analogs uncouple the in vitro and in vivo effects of farnesol.

3. Discussion

Farnesol is a biologically active, lipid-soluble molecule. We have shown that it blocks the yeast to mycelium transition in *C. albicans*,³ acts as a virulence factor in a mouse model of systemic candidiasis,^{[10] and [11]} probably by interfering with the normal progression of cytokine expression in their immune response,¹² and triggers apoptosis in the fungus *Aspergillus nidulans*.²² Others have shown that farnesol alters circadian rhythms in *Neurospora crassa* and targets HMG CoA reductase for proteolysis in both *Saccharomyces cerevisiae* and Chinese hamster ovary cells.^{[23], [24] and [25]} Farnesol has also been shown to block calcium channels, stimulating cell differentiation, and triggering apoptosis in other mammalian cell lines.²⁶ Farnesol-induced apoptosis has also been reported in tobacco cells.²⁷ In addition, farnesol has recently been found to be a potent and selective inhibitor of one class of monoamine oxidases (MAO-B).²⁸ The family of heterocyclic farnesol analogs described above now provides a tool to discriminate between yeast-hyphal quorum sensing and pathogenicity. In particular, the availability of molecules such as tetrazole **5**

(a potent quorum-sensing molecule which enhances pathogenicity) and **10** (a potent quorum-sensing molecule which does not enhance pathogenicity) may provide a potent tool in understanding how farnesol functions as a virulence factor in interactions with cellular macrophages and CD4 T cells to alter adaptive immunity.¹² The analogs also provide a means of establishing differences and similarities between farnesol's effect on virulence and its ability to block both hyphal development and biofilm formation.^{[3] and [29]} Along these lines, it is interesting to note that an analog of triazole **10** bearing five additional carbons has been reported to be a potent activator of protein kinase C.²⁰ Of course, the differences in the pathogenicity of *C. albicans* observed in the presence of farnesol, **5**, and **10** could also be due to differing pharmacokinetics or their differing influence on mouse cytokine production.

It is also intriguing to consider the basis for the failure of farnesol, tetrazole **5**, or thiotriazole **10** to exert any protective role on the course of systemic candidiasis. One possibility is that the conditions of the experiment were not well suited to test the potential of quorum-sensing molecules as fungistatic therapeutics. Mycelia, the invasive form of the fungi capable of penetrating host tissues to enter the bloodstream, are disfavored by the addition of exogenous farnesol or synthetic QSMs. However, the standard experimental protocol calls for injection of *C. albicans* directly into the bloodstream (tail vein), potentially minimizing the influence of any regimen that disfavors the mycelial morphology. Three other clinical models of candidiasis, oral,³⁰ vaginal,³¹ and gastrointestinal,³² may provide a more relevant test of the potential of a 'single morphology' therapy for control of fungal infections.

4. Conclusion

We have discovered a family of synthetic quorum-sensing molecules for *C. albicans* possessing in vitro potency comparable to *E,E*-farnesol, the natural signal. Investigations of several of the analogs in a mouse model reveals no protection against systemic candidiasis, but do demonstrate the ability to decouple quorum sensing and virulence, suggesting this family of farnesol analogs may prove extremely useful in dissecting the basis for farnesol's many biological activities.

5. Experimental

5.1. General procedures

Tetrahydrofuran (THF) was distilled from sodium/benzophenone. Other solvents were used as received. All reactions were performed under N₂ in oven-dried or flame-dried glassware. All new compounds were characterized by ¹H, ¹³C, IR, and HRMS. NMR spectra were recorded at 400 MHz (¹H) or 100 MHz (¹³C) in CDCl₃ unless otherwise noted. IR spectra were collected on NaCl plates. Melting points are uncorrected. Mass spectra were acquired at the Nebraska Center for Mass Spectrometry (Lincoln, NE). TLC analyses were conducted on Analtech 0.25 mm GHLF plates, with analytes vi-

sualized by UV, by dipping in 1% KMnO₄ (specific for alkenes), or by charring after exposure to an aqueous solution of ceric sulfate and ammonium molybdate (general indicator). (*E*)-5-(2-phenylethynyl)-1*H*-tetrazole (**8**) was purchased from Alfa Aesar ("listed as 5-styryl-1*H*-1,2,3,4-tetrazole") and used without purification. Abbreviations: EA, ethyl acetate; hex; hexane; HOAc, acetic acid; NBA, 3-nitrobenzylalcohol.

5.2. Assay of quorum sensing

Assays for quorum sensing,³ toxicity to mice,^{[10] and [11]} and mortality have been described;^{[10] and [11]} a brief overview follows: undifferentiated cells are inoculated to a final density of $1-2 \times 10^7$ cells/ml into a solution of *N*-acetyl glucosamine (2.5 mM, buffered to pH 7) and incubated for 4 h while shaking (200 rpm, 37 °C), after which the relative percentage of cells with germ tube formation (GTF) is determined. Values from triplicate readings are always within 10% of the mean value. The relative percentage of cells displaying GTF, 95–98% for cells grown on unsupplemented media, is reduced considerably in the presence of farnesol or related QSMs.⁵

5.3. Preparation of Substrates

The preparation of oximes **2** (*anti*, 2*E*,4*E*,6*E*,8*E*)-3,7,11-trimethyldodeca-2,4,6,8,10-pentaenal oxime)¹³ and **3** (*anti*, 2*E*,6*E*)-3,7,11-trimethyldodeca-2,6,10-trienal oxime)³³ has been described.

5.3.1. *anti*- and *syn*-(2*E*,6*E*)-3,7,11-Trimethyldodeca-2,6,10-trienal, *O*-methyl oxime [*anti*-4 and *syn*-4]

Into a pyridine solution (5 ml) of methoxylamine hydrochloride (0.109 g, 1.3 mmol) was added (2*E*,6*E*)-3,7,11-trimethyldodeca-2,6,10-trienal (0.115 g, 0.52 mmol; available through oxidation of commercially available farnesol with MnO₂ or chromic acid). After 4 h the reaction mixture was diluted with water (50 ml) and extracted with 30% Et₂O/Hex (100 ml). The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure to afford a mixture which was separated by flash chromatography (2% Et₂O/Hex) to afford a mixture of *anti*-4 (0.080 g, 62%) and *syn*-4 (0.045 g, 38%).

anti-4. $R_f = 0.3$ (5% EA/Hex); ¹H δ 8.01 (d, 1H, *J* = 10.4), 5.92 (d, 1H, *J* = 9.6), 5.09 (m, 2H), 3.86 (s, 3H), 2.15 (m, 4H), 2.05 (m, 2H), 1.97 (m, 2H), 1.82 (d, 3H, *J* = 1.2), 1.68 (s, 3H), 1.60 (s, 6H); ¹³C δ 147.7, 147.5, 135.8, 131.3, 124.2, 123.2, 117.8, 61.5, 40.0, 39.6, 26.7, 26.0, 25.7, 17.7, 17.2, 16.0; IR 2965, 2932, 2855, 1443, 1051; MS (HR FAB, 3-NBA) calcd for C₁₆H₂₈NO (MH⁺): 250.2171; found 250.2160 (4.4 ppm).

syn-4. $R_f = 0.2$ (5% EA/Hex) ¹H δ 7.27 (d, 1H, *J* = 10), 6.41 (dd, 1H, *J* = 10, 1.2), 5.09 (m, 2H), 3.90 (s, 3H), 2.17 (m, 4H), 2.06 (m, 2H), 1.98 (m, 2H), 1.87 (d, 3H, *J* = 1.2), 1.69 (d, 3H, *J* = 0.8), 1.61 (s, 6H); ¹³C δ 149.8, 144.8, 135.9, 131.4, 124.2, 123.2, 113.3, 61.8, 40.2, 39.7, 26.7, 26.1, 25.7, 17.7, 17.1, 16.0; IR 2964, 2932, 2855, 1639, 1441, 1056; MS (HR FAB, 3-NBA) calcd for C₁₆H₂₈NO (MH⁺): 250.2171; found 250.2166 (1.8 ppm).

5.3.2. (*E*)-5,9-Dimethyldeca-4,8-dienitrile

This compound was prepared by a variant of a known procedure.³⁴ Into a –78 °C solution of acetonitrile (0.52 ml, 10 mmol) in dry THF (10 ml) was added dropwise *n*-BuLi (5 mmol, 2.0 ml, nominally 2.5 M in hexane), followed by geranyl chloride (0.37 ml, 2.0 mmol). The reaction was allowed to warm to rt, and, after 30 min, quenched with water. The mixture was extracted with 10% EA/hex (150 ml) and the organic layer was dried over Na₂SO₄. The crude residue obtained upon concentration was purified by flash chromatography (5% EA/Hex) to afford 0.28 g (79%) of the nitrile as a colorless oil: $R_f = 0.2$ (5% EA/Hex).

5.3.3. (*E*)-5-(4,8-Dimethylnona-3,7-dienyl)-1*H*-tetrazole (**5**)

Into a suspension of NaN₃ (0.455 g, 7.00 mmol) in toluene (5 ml) was added Et₃NHCl (Fluka, 1.10 g, 8.00 mmol) followed by a solution of (*E*)-5,9-dimethyldeca-4,8-dienitrile (see above, 2.00 mmol, 0.354 g) in 2 ml of toluene. The reaction mixture was held at reflux for 24 h and then allowed to cool to rt. The mixture was acidified to pH ~ 1 with 10% aq HCl and extracted with 20% EA/Hex (3 × 100 ml). The combined organic layers were concentrated under reduced pressure and the residue purified by flash chromatography (Hex/EA/AcOH 10:2:1) to furnish 0.41 g (93%) of tetrazole **5** as a colorless oil: $R_f = 0.3$ (5:1:1, Hex/AcOH/EA); ¹H δ 13.55 (br s, 1H), 5.18 (t, 1H, *J* = 6.8), 5.04 (t, 1H, *J* = 6.5), 3.14 (t, 2H, *J* = 7.6), 2.56 (q, 2H, *J* = 7.2), 2.03 (m, 2H), 1.97 (m, 2H), 1.66 (s, 3H), 1.66 (s, 3H), 1.58 (s, 3H), 1.52 (s, 3H); ¹³C δ 156.5, 138.4, 132.0, 124.0, 121.3, 39.5, 26.4, 26.1, 25.6, 23.7, 17.7, 15.9; IR 2917, 2625, 1558, 1441, 1376, 1253; HR-FABMS (3-NBA) calcd for C₁₂H₂₁N₄ (MH⁺): 221.1766; found: 221.1771 (2.1 ppm).

5.3.4. (*E*)-4,8-Dimethylnona-3,7-dienitrile³⁵

Into a solution of NaCN (0.196 g, 4.00 mmol) in ethanol (10 ml) was added geranyl chloride (0.37 ml, 2.0 mmol). The reaction mixture was stirred for 2 h, diluted with water, and with 5% EA/Hex (twice). The separated organic layer was dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography (5% EA/Hex) to furnish 0.313 g (96%) of the known nitrile: $R_f = 0.3$ (5% EA/Hex).

5.3.5. (*E*)-5-(3,7-Dimethylocta-2,6-dienyl)-1*H*-tetrazole (**6**)

This compound was prepared from (*E*)-5-(3,7-dimethylocta-2,6-dienitrile) (0.31 g, 1.9 mmol) by a similar approach as for **5**. The product (0.37 g, 95%) was a colorless crystal (mp = 52–54 °C, uncorrected) used without further purification: $R_f = 0.3$ (5:1:1, Hex/AcOH/EA); ¹H δ 12.8 (br s, 1H), 5.45 (dt, 1H, *J* = 7.2, 1.2), 5.05 (m, 1H), 3.82 (d, 2H, *J* = 7.6), 2.05 (m, 4H), 1.69 (s, 3H), 1.68 (s, 3H), 1.57 (s, 3H); ¹³C δ 156.0, 141.5, 132.2, 123.9, 115.7, 39.3, 26.2, 25.6, 22.5, 17.6, 16.2; IR 3417, 2971, 2916, 2732, 2253; HR-FABMS (3-NBA) calcd for C₁₁H₁₉N₄ (MH⁺): 207.1610; found: 207.1612 (1.3 ppm).

5.3.6. 5-Nonyl-1*H*-tetrazole (**7**)

This compound was prepared from decanonitrile (0.38 ml, 2.0 mmol) by a similar method as employed for synthesis of

5 and 6. The crude material (0.359 g, 91%) displayed spectra identical to literature reports³⁶ and was used without further purification: $R_f = 0.2$ (5:1:1, Hex/AcOH/EA).

5.3.7. (*E*)-5-(3,7-Dimethylocta-2,6-dienylthio)-1H-tetrazole (9)

Into a solution of 5-mercapto-1H-tetrazole (0.20 g, 2.0 mmol) in ethyl alcohol (10 ml) were added K_2CO_3 (0.55 g, 4.0 mmol) and geranyl chloride (0.19 ml, 1.0 mmol). After 12 h, the reaction mixture was acidified to pH 7 and extracted with 50% EA/Hex (3 × 50 ml). The combined organic extracts were dried with Na_2SO_4 . The residue obtained upon concentration in vacuo was purified by flash chromatography (5:1:1 Hex/AcOH/EA) to furnish 0.213 g (89%) of mercaptotetrazole **9** as a colorless oil: $R_f = 0.2$ (5:1:1, Hex/AcOH/EA); 1H δ 13.0 (br s, 1H), 5.37 (t, 1H, $J = 7.6$), 5.03 (t, 1H, $J = 6.4$), 3.97 (d, 2H, $J = 8$), 2.03 (m, 4H), 1.68 (s, 3H), 1.66 (s, 3H), 1.56 (s, 3H); ^{13}C δ 155.1, 143.4, 132.1, 123.5, 116.8, 39.4, 31.4, 26.2, 26.6, 17.7, 16.2; IR 3055, 2917, 2727, 2560, 1504, 1447, 1038; HR-FABMS (3-NBA) calcd for $C_{11}H_{19}N_4$ (MH^+): 239.1330; found 239.1336 (2.2 ppm).

5.3.8. (*E*)-3-(3,7-Dimethylocta-2,6-dienylthio)-1H-1,2,4-triazole (10)

Into a solution of 1H-1,2,4-triazole-3-thiol (0.152 g, 1.50 mmol) in ethyl alcohol (10 ml) were added K_2CO_3 (0.276 g, 2.00 mmol) and geranyl chloride (0.19 ml, 1.0 mmol). After 12 h, the reaction mixture was worked up as for **9** and the residue purified by flash chromatography (40% EA/Hex) to furnish 0.235 g (99%) of thiotriazole **10**: $R_f = 0.3$ (50% EA/Hex); 1H δ 13.70 (br s, 1H), 8.18 (s, 1H), 5.33 (dt, 1H, $J = 1.2, 2$), 5.02 (m, 1H), 3.80 (d, 2H, $J = 8$), 2.00 (m, 4H), 1.64 (s, 3H), 1.56 (s, 3H); ^{13}C δ 156.4, 147.7, 141.5, 131.8, 131.8, 123.6, 118.1, 39.4, 31.2, 26.2, 25.6, 17.6, 16.0; IR 3111, 2924, 1449, 1276, 910, 734; MS (HR-FABMS, 3-NBA) calcd for $C_{12}H_{20}N_3S$ (MH^+): 238.1378; found: 238.1383 (2.3 ppm).

5.3.9. (*E*)-4-(3,7-Dimethylocta-2,6-dienylthio)-1H-1,2,3-triazole (11)

Into a 0 °C solution of the sodium salt of 4-mercapto-1H-1,2,3-triazole (0.262 g, 2.2 mmol) in ethanol (10 ml) was added geranyl chloride (0.19 ml, 1.0 mmol). The reaction mixture was stirred for 2 h and then allowed to warm to RT over the course of 1 h. The reaction mixture was diluted with water and the mixture extracted with 20% EA/Hex (3 × 50 ml). The combined organic extracts were dried with Na_2SO_4 and concentrated, with the residue purified by flash chromatography (20% EA/Hex) to furnish 0.218 g (92%) of thiotriazole **11**. $R_f = 0.2$ (20% EA/Hex); 1H δ 14.23 (br s, 1H), 7.72 (s, 1H), 5.28 (d of t, 1H, $J = 1.2, 8$), 5.02 (m, 1H), 3.54 (d, 2H, $J = 7.6$), 1.97 (m, 4H), 1.64 (d, 3H, $J = 0.8$), 1.55 (s, 3H), 1.47 (s, 3H); ^{13}C δ 140.7, 139.3, 133.5, 131.6, 123.7, 118.8, 39.3, 33.0, 26.2, 25.5, 17.6, 15.7; IR 3130, 2916, 1661, 1448, 1228, 1109, 839; HR-FABMS (3-NBA) calcd for $C_{12}H_{20}N_3S$, MH^+ : 238.1378; found: 238.1369 (3.8 ppm).

5.3.10. (*E*)-2-(3,7-Dimethylocta-2,6-dienylthio)-1H-imidazole (12)

This compound was prepared from 2-mercaptoimidazole (0.12 g, 1.2 mmol) and geranyl chloride (0.19 ml, 1.0 mmol) in 96% yield (0.227 g) by a similar procedure as for **9**. The crude product was pure by NMR and was utilized without further purification: $R_f = 0.4$ (50% EA/Hex); 1H δ 12.06 (br s, 1H), 7.17 (s, 1H), 5.29 (dt, 1H, $J = 1.2, 7.6$), 5.03 (m, 1H), 3.62 (d, 2H, $J = 8$), 2.05–1.92 (m, 4H), 1.65 (s, 3H), 1.56 (s, 3H), 1.46 (s, 3H); ^{13}C δ 140.5, 139.6, 131.6, 123.9, 123.8, 119.0, 39.4, 33.4, 26.2, 25.6, 17.6, 15.6; IR 3094, 2916, 2637, 1447, 1328, 1098; HR-FABMS (3-NBA) calcd for $C_{13}H_{20}N_2S$ (MH^+): 237.1313; found 237.1310 (1.1 ppm).

5.3.11. (*E*)-4-(4,8-Dimethylnona-3,7-dienyl)-1H-1,2,3-triazole (13)

Into a solution of (*E*)-6,10-dimethylundeca-5,9-dien-1-yne³⁷ (0.176 g, 1.00 mmol) in 9:1 DMF/MeOH (10 ml) was added $TMSN_3$ (1.3 ml, 10 mmol) followed by CuI (0.1 g, 0.5 mmol).²¹ The mixture was held at 100 °C for 12 h and then allowed to cool to rt. The reaction mixture was diluted with water and then extracted with 20% EA/Hex (3 × 70 ml). The combined organic layers were dried with Na_2SO_4 and concentrated. The residue was purified by flash chromatography (20% EA/Hex) to furnish 0.168 g (77%) of triazole **13**: $R_f = 0.2$ (20% EA/Hex); 1H (CD_3OD) δ 7.53 (s, 1H), 5.15 (dt, 1H, $J = 1.2, 7.2$), 5.05 (m, 1H), 2.76 (t, 2H, $J = 7.6$), 2.35 (q, 2H, $J = 7.2$), 2.04 (m, 2H), 1.98 (m, 2H), 1.65 (s, 3H), 1.57 (s, 3H), 1.54 (s, 3H); ^{13}C (CD_3OD , 100 MHz) δ 144.2, 137.7, 132.3, 128.7, 125.4, 124.2, 40.9, 28.8, 27.8, 26.1, 24.2, 17.9, 16.3; IR 3140, 2919, 2856, 1448, 1377, 1108, 983, 838; HR-FABMS (3-NBA) calcd for $C_{13}H_{21}N_3$ (MH^+): 220.1814; found: 220.1811 (1.1 ppm).

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References and notes

- O. Gudlaugsson, S. Gillespie, K. Lee, J. Vande Berg, J. Hu, S. Messer, L. Herwaldt, M. Pfaller and D. Diekema, *Clin. Infect. Dis.* **37** (2003), pp. 1172–1177.
- G.L. Macphail, G.D. Taylor, M. Buchanan-Chell, C. Ross, S. Wilson and A. Kureishi, *Mycoses* **45** (2002), pp. 141–145.
- J.M. Hornby, E.C. Jensen, A.D. Lise, J.J. Tasto, B. Jahnke, R. Shoemaker, P. Dussault and K.W. Nickerson, *Appl. Environ. Microbiol.* **67** (2001), pp. 2982–2992.

- 4 K.W. Nickerson, A.L. Atkin and J.M. Hornby, *Appl. Environ. Microbiol.* **72** (2006), pp. 3805–3813.
- 5 D.D. Mosel, R. Dumitru, J.M. Hornby, A.L. Atkin and K.W. Nickerson, *Appl. Environ. Microbiol.* **71** (2005), pp. 4938–4940.
- 6 R. Shchepin, J.M. Hornby, E. Burger, T. Niessen, P. Dussault and K.W. Nickerson, *Chem. Biol.* **10** (2003), pp. 743–750.
- 7 W.C. Fuqua, S.C. Winans and E.P. Greenberg, *J. Bacteriol.* **176** (1994), pp. 269–275.
- 8 S.P. Saville, A.L. Lazzell, C. Monteagudo and J.L. Lopez-Ribot, *Eukaryot. Cell* **2** (2003), pp. 1053–1060.
- 9 J.M. Hornby and K.W. Nickerson, *Antimicrob. Agents Chemother.* **48** (2004), pp. 2305–2307.
- 10 D.H. Navarathna, J.M. Hornby, N. Hoerrmann, A.M. Parkhurst, G.E. Duhamel and K.W. Nickerson, *J. Antimicrob. Chemother.* **56** (2005), pp. 1156–1159.
- 11 D.H. Navarathna, J.M. Hornby, N. Krishnan, A. Parkhurst, G.E. Duhamel and K.W. Nickerson, *Infect. Immun.* **75** (2007), pp. 1609–1618.
- 12 D.H. Navarathna, K.W. Nickerson, G.E. Duhamel, T.R. Jerrels and T.M. Petro, *Infect. Immun.* **75** (2007), pp. 4006–4011.
- 13 R. Shchepin, R. Dumitru, K.W. Nickerson, M. Lund and P.H. Dussault, *Chem. Biol.* **12** (2005), pp. 639–641.
- 14 R.J. Herr, *Bioorg. Med. Chem.* **10** (2002), pp. 3379–3393.
- 15 C.W. Thornber, *Chem. Soc. Rev.* **8** (1979), pp. 563–580.
- 16 S. Fürmeier and J.O. Metzger, *Eur. J. Org. Chem.* (2003), pp. 885–893.
- 17 We thank a reviewer for this suggestion.
- 18 D.A. Hogan, A. Vik and R. Kolter, *Mol. Microbiol.* **54** (2004), pp. 1212–1223.
- 19 M. Kume, T. Kubota, Y. Kimura, H. Nakashimizu and K. Motokawa, *J. Antibiot.* **46** (1993), pp. 316–330.
- 20 B.A. Gilbert, Y.-H. Lim, J. Ding, J.A. Badwey and R.R. Rando, *Biochemistry* **34** (1995), pp. 3916–3920.
- 21 T. Jin, S. Kamijo and Y. Yamamoto, *Eur. J. Org. Chem.* (2004), pp. 3789–3791.
- 22 C.P. Semighini, J.M. Hornby, R. Dumitru, K.W. Nickerson and S.D. Harris, *Mol. Microbiol.* **59** (2006), pp. 753–764.
- 23 T. Granshaw, M. Tsukamoto and S. Brody, *J. Biol. Rhythms* **18** (2003), pp. 287–296.
- 24 A.G. Shearer and R.Y. Hampton, *EMBO J.* **24** (2005), pp. 149–159.
- 25 T.E. Meigs and R.D. Simoni, *Arch. Biochem. Biophys.* **345** (1997), pp. 1–9.
- 26 P.A. Edwards and J. Ericsson, *Ann. Rev. Biochem.* **68** (1999), pp. 157–185.
- 27 A. Hemmerlin and T.J. Bach, *Plant Physiol.* **123** (2000), pp. 1257–1268.
- 28 (a)C. Binda, F. Hubálek, M. Li, N. Castagnoli, D.E. Edmondson and A. Mattevi, *Neurology* **67** (Suppl. 2) (2006), pp. S5–S7.
(b)A.A. Khalil, B. Davies and N. Castagnoli, *Bioorg. Med. Chem.* **14** (2006), pp. 3392–3398.
(c)F. Hubálek, C. Binda, A. Khalil, M. Li, A. Mattevi, N. Castagnoli and D.E. Edmondson, *J. Biol. Chem.* **280** (2005), pp. 15761–15766.
- 29 G. Ramage, S.P. Saville, B.L. Wickes and J.L. Lopez-Ribot, *Appl. Environ. Microbiol.* **68** (2002), pp. 5459–5463.
- 30 S. Elahi, G. Pang, R. Clancy and R.B. Ashman, *Infect. Immun.* **68** (2000), pp. 5771–5777.
- 31 G. Cheng, K. Woznaik, M.A. Wallig, P.L. Fidel Jr., S.R. Trupin and L.L. Hoyer, *Infect. Immun.* **73** (2005), pp. 1656–1663.
- 32 G. Samonis, E.J. Anaissie, B. Rosenbaum and G.P. Bodey, *Infect. Immun.* **58** (1990), pp. 1514–1517.
- 33 G.C. Zheng and H. Kakisawa, *Bull. Chem. Soc. Jpn.* **62** (1989), pp. 602–604.
- 34 D.S. Shin, *J. Kor. Chem. Soc.* **36** (1992), pp. 579–583.
- 35 (a)K. Mori and Y. Funaki, *Tetrahedron* **41** (1985), pp. 2369–2377.
(b)T. Takahashi, S. Tomida and T. Doi, *Synlett* (1999), pp. 644–646.
- 36 F. Kiuchi, N. Miyashita, Y. Tsuda, K. Kondo and H. Yoshimura, *Chem. Pharm. Bull.* **35** (1987), pp. 2880–2886.
- 37 B.H. Lipshutz, G. Bulow, F. Fernandez-Lazaro, S.K. Kim, R. Lowe, P. Mollard and K.L. Stevens, *J. Am. Chem. Soc.* **121** (1999), pp. 11664–11673.