

University of Nebraska - Lincoln

DigitalCommons@University of Nebraska - Lincoln

Public Health Resources

Public Health Resources

12-2004

West Nile Virus Blood Transfusion-Related Infection Despite Nucleic Acid Testing

Alexandre Macedo de Oliveira

Brady D. Beecham

Susan P. Montgomery

Robert S. Lanciotti

Jeffrey M. Linnen

See next page for additional authors

Follow this and additional works at: <https://digitalcommons.unl.edu/publichealthresources>



Part of the [Public Health Commons](#)

de Oliveira, Alexandre Macedo; Beecham, Brady D.; Montgomery, Susan P.; Lanciotti, Robert S.; Linnen, Jeffrey M.; Giachetti, Cristina; Pietrelli, Larry A.; Stramer, Susan L.; and Safraneck, Thomas J., "West Nile Virus Blood Transfusion-Related Infection Despite Nucleic Acid Testing" (2004). *Public Health Resources*. 30.

<https://digitalcommons.unl.edu/publichealthresources/30>

This Article is brought to you for free and open access by the Public Health Resources at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Public Health Resources by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

Authors

Alexandre Macedo de Oliveira, Brady D. Beecham, Susan P. Montgomery, Robert S. Lanciotti, Jeffrey M. Linnen, Cristina Giachetti, Larry A. Pietrelli, Susan L. Stramer, and Thomas J. Safraneck

West Nile virus blood transfusion-related infection despite nucleic acid testing

Alexandre Macedo de Oliveira, Brady D. Beecham, Susan P. Montgomery, Robert S. Lanciotti, Jeffrey M. Linnen, Cristina Giachetti, Larry A. Pietrelli, Susan L. Stramer, and Thomas J. Safraneck

BACKGROUND: A case of West Nile virus (WNV) encephalitis associated with transfusion of blood that did not react when tested for WNV by minipool (MP) nucleic acid testing (NAT) is described. A Nebraska man developed clinical encephalitis 13 days after surgery and transfusion of 26 blood components. Antibody testing confirmed WNV infection. An investigation was initiated to determine the source of this infection.

STUDY DESIGN AND METHODS: The patient's family members were interviewed to identify risk factors for WNV infection. Residual samples were retested for WNV RNA using transcription-mediated amplification (TMA) assay and two polymerase chain reaction (PCR) assays. Blood donors' follow-up serum samples were collected. All samples were tested for WNV-specific immunoglobulin M antibodies.

RESULTS: The patient's family denied recent mosquito exposure. The 20 blood components collected after July 2003 did not react when tested for WNV in a six-member MP-NAT at the time of donation. Retrospective individual testing identified one sample as WNV-reactive by the TMA assay and one of the PCR assays. Seroconversion was demonstrated in the donor associated with this sample.

CONCLUSION: WNV RNA detection by individual donation NAT demonstrates viremic blood escaping MP-NAT and supports transfusion-related WNV transmission. MP-NAT may not detect all WNV-infected blood donors, allowing WNV transmission to continue at low levels. WNV NAT assays might vary in sensitivity and pooling donations could further impact test performance. Understanding MP NAT limitations can improve strategies to maintain safety of the blood supply in the United States.

West Nile virus (WNV), a mosquito-borne flavivirus, was initially seen in the US in 1999 and first reported among Nebraska residents in 2002.^{1,2} Humans serve as incidental hosts, and most infections are asymptomatic; approximately 30 percent of infections result in a non-neuroinvasive disease known as West Nile fever, and less than 1 percent of infected individuals develop severe diseases such as meningitis and/or encephalitis.³⁻⁵ In the US, a total of 9862 human cases of WNV disease were reported in 45 states and the District of Columbia in 2003. Nebraska reported more than 1900 human WNV cases in 2003, ranking second only to Colorado.⁶ Blood transfusion-related transmission of WNV infection during the 2002 US epidemic prompted rapid development of two investigational nucleic acid testing (NAT) assays to screen donated blood for WNV viremia: the TaqScreen WNV test (Roche

ABBREVIATIONS: IDT = individual donation testing; IND = investigational new drug; MP(s) = minipool(s); PRNT = plaque reduction neutralization test; SLE = St Louis encephalitis; WNV = West Nile virus.

From the Epidemic Intelligence Service, Division of Applied Public Health Training, Epidemiology Program Office, Centers for Disease Control and Prevention, Atlanta, Georgia; the Office of Epidemiology, Nebraska Health and Human Services System, Lincoln, Nebraska; the Division of Vector-Borne Infectious Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Fort Collins, Colorado; Gen-Probe Inc., San Diego, California; Roche Molecular Systems Inc., Pleasanton, California; and the American Red Cross, Gaithersburg, Maryland.

Address reprint requests to: Thomas J. Safraneck, Nebraska Health and Human Services, 301 Centennial Mall South, PO Box 95007, Lincoln, NE 68509-5007; e-mail: tom.safraneck@hhss.state.ne.us.

Received for publication April 9, 2004; revision received June 14, 2004, and accepted June 30, 2004.

TRANSFUSION 2004;44:1695-1699.

Molecular Systems Inc., Pleasanton, CA) and the Procleix WNV assay (Gen-Probe Inc., San Diego, CA).^{7,8} (Use of trade names is for identification purposes only and does not constitute endorsement by the US Public Health Service or the Centers for Disease Control and Prevention [CDC].) The tests were implemented between June and August 2003.^{9,10} Because of automation constraints, the tests are performed with minipools (MPs) of either six (TaqScreen assay) or 16 (Procleix assay) donations. A limited number of blood centers have the capacity to routinely screen by individual donation testing (IDT); in addition, some programs converted to IDT for limited periods during the 2003 epidemic. Donated blood from reactive pools is individually retested to identify the WNV-reactive donation. Donors associated with WNV-reactive individual tests are deferred from donation for at least 4 weeks. The WNV-reactive donation and any of that donor's unexpired blood components obtained in the 28 days before the reactive tests are discarded. In this report, we describe the investigation of a patient who developed WNV encephalitis after receiving blood that had did not react with NAT for WNV in a six-member MP.

CASE REPORT

On August 4, 2003, an 80-year-old Nebraska man underwent open-heart surgery. During surgery, the patient received 26 blood components because of massive bleeding: 8 units of red blood cells, 6 units of fresh frozen plasma (FFP), 2 units of single-donor platelets (PLTs), and 10 units of random-donor PLTs. The patient recovered and was discharged to home 10 days later.

On August 19, 2003, 15 days after transfusion, the patient was rehospitalized after experiencing mental confusion and fever for 2 days. The patient's condition worsened, and he was placed on ventilatory support for 3 weeks. His condition gradually improved, and he was subsequently transferred to a rehabilitation facility.

WNV encephalitis was suspected, and serum and cerebrospinal fluid were collected during the second hospitalization. These tested positive for the presence of WNV-specific immunoglobulin M (IgM) antibodies at the Nebraska Public Health Laboratory (Table 1).

On September 10, 2003, 37 days after transfusion, a convalescent serum sample was collected from the patient. This sample tested positive for the presence of WNV by WNV-specific IgM enzyme-linked immunosorbent assay (ELISA) and WNV plaque reduction

neutralization test (PRNT) and negative for the presence of St Louis encephalitis (SLE) by PRNT at the CDC (Table 1).

Upon receipt of the first positive WNV IgM test, the hospital infection-control practitioners notified the Office of Epidemiology at the Nebraska Health and Human Services System and an investigation was launched to determine the source of this infection.

MATERIALS AND METHODS

Family members of the infected patient were interviewed in August 2003 to evaluate the potential for mosquito exposure after surgery. In addition, all transfused blood components were identified and traced to their donors. Residual samples from the donation testing tubes and unused co-components were located and stored frozen (-20°C). Implicated donors were interviewed about symptoms consistent with WNV infection, and a follow-up blood sample was collected in September 2003.

Quarantined residual samples were tested individually for the presence of WNV RNA with three different laboratory tests: the two investigational assays used for WNV blood donor screening and an in-house assay. The tests were a transcription-mediated amplification assay (Procleix WNV assay, Gen-Probe Inc.) and two different dynamic quantitative polymerase chain reaction (PCR) assays: TaqScreen WNV test (Roche Molecular Systems Inc.) and an in-house PCR (CDC, Fort Collins, CO).¹¹⁻¹³ Reactive specimens were further tested with the commercial assays: 10 replicates with undiluted samples and 10

TABLE 1. Laboratory tests performed on the index patient at time of readmission and convalescence, Nebraska, 2003

Laboratory study	Collection date	Result	Normal values
Hemoglobin (g/100 mL)	8/20/2003	11	13.5-17.0
WBC count ($\times 10^3$ cells/mm ³)	8/20/2003	9	4-10
Differential (%)			
Neutrophils		57	40-80
Lymphocytes		39	20-45
Monocytes		3	0-10
Eosinophils		1	0-7
Basophils		0	0-2
Serum			
Glucose (mg/dL)	8/20/2003	103	65-120
Protein (g/dL)	8/20/2003	6.1	6.0-8.3
CSF*			
Glucose (mg/dL)	8/20/2003	58	<105
Protein (mg/dL)	8/20/2003	98.4	<42
WBC count (cells/mm ³)	8/20/2003	155	<3
Mononuclear (%)		39	
Segmented neutrophils (%)		61	
Arbovirus studies			
WNV IgM (serum)	8/20/2003	Positive	Negative
WNV IgM (CSF)	8/20/2003	Positive	Negative
WNV IgM (serum)	9/10/2003	Positive	Negative
WNV PRNT	9/10/2003	Positive	Negative
SLE PRNT	9/10/2003	Negative	Negative

* CSF = cerebrospinal fluid.

with MP-dilution levels (1:6 for the TaqScreen assay and 1:16 for the Procleix assay). Viral load of reactive specimens was determined by quantitative PCR (SuperQuant for WNV, National Genetics Institute, Los Angeles, CA) and through dilutional studies at CDC and Gen-Probe Inc.

Residual samples from donations and follow-up samples were tested with WNV IgM antibody-capture ELISA. WNV infection was confirmed with PRNT for WNV and SLE.¹⁴

RESULTS

Exposure assessment

Family members denied recent mosquito exposure, reporting that the patient was hospitalized or bedridden at home during the 2 weeks before symptom onset. The infected patient received 26 blood components from 26 different donors. These donations generated 41 additional blood components: 18 were transfused to other patients, 17 were quarantined, and 6 were discarded before our investigation began.

Six of the 26 donations took place in February 2003 and had no residual samples available at the time of investigation. The remaining 20 donations were made in July and August 2003. Thirteen of these had approximately 200 mL of quarantined FFP available for testing; the other 7 had approximately 2 mL of serum remaining.

Results from donations

The CDC PCR, the investigational TaqScreen assay, and the investigational Procleix assay were all carried out on the 13 high-volume specimens by the developers of each test. One specimen reacted by the Procleix assay, equivocal by the CDC PCR, and did not react by the TaqScreen assay. A subsequent CDC PCR assay with an increased RNA extraction volume reacted. The viral load in this sample was estimated to be 560 copies per mL by the Super-Quant assay and 40 and 30 copies per mL by CDC and

Gen-Probe Inc., respectively. Replicate results with the Procleix assay showed reactivity in all 10 replicates with undiluted samples and in 3 of 10 at 1:16 dilution. The TaqScreen assay exhibited reactivity in 5 of 10 and 2 of 10 replicates at undiluted and 1:6-diluted samples, respectively. This donation generated 1 unit of PLTs, which was transfused to the index patient. The other 12 high-volume samples did not react in all NAT assays.

The seven 2-mL specimens were aliquotted and sent to CDC and the other test developers' laboratories. They all did not react by the CDC PCR and the Procleix assay. Volume was insufficient for the TaqScreen assay.

WNV-specific IgM was negative for all 20 donations that had residual samples available for testing. Test results are shown in Table 2.

Donor follow-up

We obtained follow-up samples from 24 of the 26 donors in mid-September 2003 and tested these for WNV-specific IgM antibodies. The median interval between donation and follow-up sample collection was 44 days (range, 38-219 days). The donor associated with the NAT-positive sample (Donor A) and two other donors tested positive for IgM antibodies (Table 2). Confirmatory PRNT tests were positive for the presence of WNV and negative for the presence of SLE on these three samples. These three donors denied WNV symptoms during the 30 days before and after donation.

Blood co-components from WNV-specific IgM-positive donors

The two additional blood components derived from Donor A's donation were not transfused. Units from the other two IgM-positive donors were transfused to two other patients. Both recipients were asymptomatic for WNV. One tested negative for the presence of WNV IgM, and the other declined testing.

TABLE 2. Test results obtained on donated blood and follow-up samples for the 26 donors, Nebraska, 2003

Donor	Original donation				Follow-up sample		
	WNV IgM	CDC PCR	Procleix assay	TaqScreen assay	WNV IgM	WNV PRNT	SLE PRNT
A	Negative	Reactive*	Reactive	Did not react	Positive	Positive	Negative
B	Negative	Did not react	Did not react	Did not react	Positive	Positive	Negative
C	Negative	Did not react	Did not react	NA†	Positive	Positive	Negative
D-M	Negative	Did not react	Did not react	Did not react	Negative	NA	NA
N	Negative	Did not react	Did not react	Did not react	NA	NA	NA
O-T	Negative	Did not react	Did not react	NA	Negative	NA	NA
U-Y‡	NA	NA	NA	NA	Negative	NA	NA
Z‡	NA	NA	NA	NA	NA	NA	NA

* Test performed using high extraction volume.

† NA = not available.

‡ Donations from February 2003.

DISCUSSION

This investigation describes a patient with WNV encephalitis acquired through transfusion of WNV viremic blood that went undetected by MP-NAT screening but reacted for WNV by IDT. Our conclusion is supported by the detection of WNV RNA in one of the donations, followed by seroconversion in the corresponding donor. The implicated donor never developed symptomatic WNV infection. WNV RNA likely was present at levels below the detection limit of the MP format.

Although this investigation suggests WNV blood transfusion-related infection, it has three main limitations. First, the infected patient could have contracted WNV before surgery. Because no presurgical samples from the infected patient were available for testing, excluding existing infection was impossible. Second, it was not possible to rule out mosquito exposure as the mode of transmission. The patient denied symptoms before surgery, however, and his convalescent condition kept him hospitalized or indoors at home, making this other source of infection unlikely. Finally, because two other seroconversions were documented among these donors, we cannot exclude the possibility of an alternate-source donor. Recipients of co-components, however, from donated blood from the seroconverters did not experience WNV symptoms, nor was WNV RNA identified in residual samples from their donations. The high incidence of WNV infection in Nebraska during summer 2003 makes it possible that these persons were infected after donating blood.

Although the six donations from February 2003 were not available for NAT, these donations occurred before WNV was circulating in Nebraska. In addition, follow-up samples from five of these donors were negative for the presence of WNV-specific IgM antibodies, making them an unlikely source of infection.

WNV blood transfusion-related infections were first described during the 2002 epidemic.^{7,8} To minimize this risk, the Food and Drug Administration (FDA) recommended deferring donors reporting fever with headache in the week before donation.¹⁵ In addition to the clinical screening, beginning June 2003, NAT was introduced as a blood-screening tool with protocols conducted under FDA's investigational new drug (IND) mechanisms.^{9,10} Recognizing technology and resource constraints, the IND protocols allowed for blood donation screening in MPs of 6 (TaqScreen assay) or 16 (Procleix assay) members. Pooling donations may reduce assay sensitivity and increase the possibility of missing donors with low-level viremia.

In this report, the six-member MP TaqScreen assay performed on the donations did not react, and the blood products were released for use in accordance with the IND protocol. A patient developed WNV encephalitis 13 days after transfusion with these products, a time frame com-

patible with WNV's incubation period. Residual samples from these donations were subsequently retested at three laboratories. For one donation, samples tested with replicate tests inconsistently reacted by the TaqScreen assay but reacted by the Procleix assay. Viral quantitation of that sample detected low levels of WNV RNA, although variability among quantitative results was observed. Dilutional testing showed that the MP format compromised test performance compared with IDT. The replicate testing also indicated that differences in sensitivity might exist between the investigational NAT assays. Further evaluation is necessary to confirm these conclusions.

The timely development of WNV NAT and its implementation under the IND mechanism are the result of an extraordinary effort by commercial laboratories, CDC, FDA, and the blood-collection community to enhance the safety of the blood supply in the US. In 2003, a total of 818 US blood donations were identified as "presumed viremic," which is defined as repeat reactivity on the same or on a different NAT assay.¹⁶ The high number of viremic donors with asymptomatic WNV infection highlights the need for highly sensitive WNV-specific NAT to ensure blood supply safety. In addition, strategies such as the implementation of IDT in regions with high WNV infection rates should be considered.

NAT for human immunodeficiency virus-1 and hepatitis C virus has improved the identification of infected donors and is universally performed by the US blood industry, although it has been demonstrated to be cost-ineffective by traditional expenditure versus benefit estimates for medical interventions.¹⁷⁻¹⁹ NAT could be cost-effective for WNV screening of the blood supply, because seasonal incidence is high in certain geographic areas, infection is asymptomatic in more than 80 percent of cases, and long periods of viremia can occur.^{20,21}

WNV can be effectively transmitted through blood transfusions. The burden of WNV blood transfusion-related disease is probably underestimated owing to the low rate of symptomatic disease among recipients and the difficulty in establishing blood transfusion as the source of WNV infection.⁸ The presence of infectious WNV in donated blood necessitates the application of viral detection testing such as NAT. Additional studies are needed to define the sensitivity of current NAT assays, their ability to detect low-level viremic donations, and their cost-effectiveness. This would allow optimization of testing protocols.

Emerging infectious diseases pose risks to blood safety.²² Health-care workers and public-health authorities should maintain surveillance for the possibility of WNV and other arboviral transmission through blood transfusions. The development and implementation of WNV NAT for blood screening has contributed to a decreased risk of blood-borne transmission in the US. Nevertheless, low-viremic donors may go undetected by MP-NAT.

ACKNOWLEDGMENTS

We are indebted to Michael P. Busch, MD, PhD, Blood Systems Research Institute, San Francisco, California, for his assistance in facilitating laboratory testing. We thank Amy Broulik and Stephanie Miller, both from Gen-Probe Inc., and Lawrence Cheng, from Roche Molecular Systems Inc., for their excellent technical assistance.

REFERENCES

1. Nash D, Mostashari F, Fine A, et al. The outbreak of West Nile virus infection in the New York City area in 1999. *N Engl J Med* 2001;344:1807-14.
2. West Nile virus activity—United States, August 21-28, 2002, and Illinois, January 1-August 27, 2002. *MMWR Morb Mortal Wkly Rep* 2002;51:764-6.
3. Petersen LR, Marfin AA. West Nile virus: a primer for the clinician. *Ann Intern Med* 2002;137:173-9.
4. Weiss D, Carr D, Kellachan J, et al. Clinical findings of West Nile virus infection in hospitalized patients, New York and New Jersey, 2000. *Emerg Infect Dis* 2001;7:654-8.
5. Chowers MY, Lang R, Nassar F, et al. Clinical characteristics of the West Nile virus fever outbreak, Israel, 2000. *Emerg Infect Dis* 2001;7:675-8.
6. 2003 West Nile virus activity in the United States (Internet). Atlanta: Centers for Disease Control and Prevention; 2004 [accessed 2004 Jun 2]. Available from: http://www.cdc.gov/ncidod/dvbid/westnile/surv&controlCaseCount03_detailed.htm
7. Harrington T, Kuehnert MJ, Kamel H, et al. West Nile virus infection transmitted by blood transfusion. *Transfusion* 2003;43:1018-22.
8. Pealer LN, Marfin AA, Petersen LR, et al. Transmission of West Nile virus through blood transfusion in the United States in 2002. *N Engl J Med* 2003;349:1236-45.
9. Detection of West Nile virus in blood donation—United States, 2003. *MMWR Morb Mortal Wkly Rep* 2003;52:769-72.
10. Blood centers begin implementing WNV donor screening tests. *ABC Newslett* 2003;1-3.
11. Linnen JM, Lewis M, Broulik A, et al. Procleix® WNV assay: a transcription-mediated amplification based assay for screening blood donations for West Nile virus RNA. *Transfusion* 2003;43S:129A.
12. Gallarda JL, Cheng L, Pai A, et al. Non-clinical performance of the Roche TaqScreen™ West Nile virus system. *Transfusion* 2003;43S:129A.
13. Lanciotti RS, Kerst AJ, Nasci RS, et al. Rapid detection of West Nile virus from human clinical specimens, field-collected mosquitoes, and avian samples by a TaqMan reverse transcriptase-PCR assay. *J Clin Microbiol* 2000;38:4066-71.
14. Martin DA, Muth DA, Brown T, et al. Standardization of immunoglobulin M capture enzyme-linked immunosorbent assays for routine diagnosis of arboviral infections. *J Clin Microbiol* 2000;38:1823-6.
15. Guidance for industry: revised recommendations for the assessment of donor suitability and blood and blood product safety in cases of known or suspected West Nile virus infection (Internet). Rockville (MD): Center for Biologics Evaluation and Research; 2003 [accessed 2003 Nov 1]. Available from: <http://www.fda.gov/cber/gdlns/wnvguid.htm>
16. Update: West Nile virus screening of blood donations and transfusion-associated transmission—United States, 2003. *MMWR Morb Mortal Wkly Rep* 2004;53:281-4.
17. Schreiber GB, Busch MP, Kleiman SH, et al. The risk of transfusion-transmitted viral infections. *N Engl J Med* 1996;334:1685-90.
18. Stramer SL. US NAT yield: where are we after 2 years? *Transfus Med* 2002;12:243-53.
19. Jackson BR, Busch MP, Stramer SL, Aubuchon JP. The cost-effectiveness of NAT for HIV, HCV, and HBV in whole-blood donations. *Transfusion* 2003;43:721-9.
20. Mostashari F, Bunning ML, Kitsutani PT, et al. Epidemic West Nile encephalitis, New York, 1999: results of a household-based seroepidemiological survey. *Lancet* 2001;358:261-4.
21. Biggerstaff BJ, Petersen LR. Estimated risk of West Nile virus transmission through blood transfusion during an epidemic in Queens, New York City. *Transfusion* 2002;42:1019-26.
22. Chamberland ME. Emerging infectious agents: do they pose a risk to the safety of transfused blood and blood products? *Clin Infect Dis* 2002;34:797-805. ■